

BGGN 213
Structural Bioinformatics II
Lecture 12
Barry Grant
UC San Diego
<http://thegrantlab.org/bggn213>

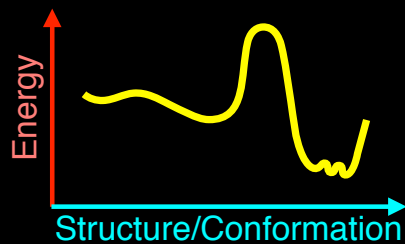
Download MGL Tools: See class website!

Next Up:

- Overview of structural bioinformatics
 - Motivations, goals and challenges
- Fundamentals of protein structure
 - Structure composition, form and forces
- Representing, interpreting & modeling protein structure
 - Visualizing and interpreting protein structures
 - Analyzing protein structures
 - Modeling energy as a function of structure
 - Drug discovery & Predicting functional dynamics

Key concept:

Potential functions describe a systems energy as a function of its **structure**



Two main approaches:

- (1). Physics-Based
- (2). Knowledge-Based

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(1). Physics-Based

(2). Knowledge-Based

For **physics** based potentials
energy terms come from physical theory

$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

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Sum of **bonded** and **non-bonded**
atom-type and position based terms

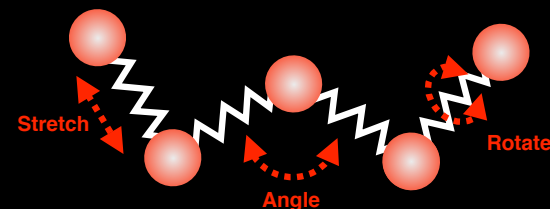
$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

E_{bonded} is itself a sum of three terms:

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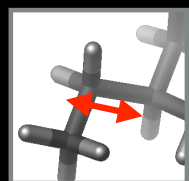
$$E_{\text{bond.stretch}} + E_{\text{bond.angle}} + E_{\text{bond.rotate}}$$



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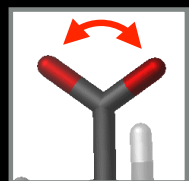
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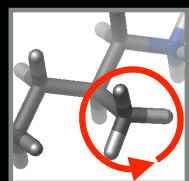
Bond Stretch

$$E_{\text{bond.stretch}}$$



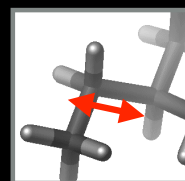
Bond Angle

$$E_{\text{bond.angle}}$$



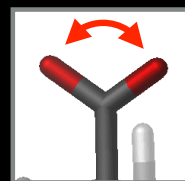
Bond Rotate

$$E_{\text{bond.rotate}}$$



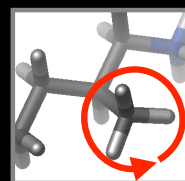
Bond Stretch

$$\sum_{\text{bonds}} K_i^{bs} (b_i - b_o)$$



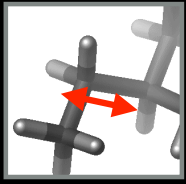
Bond Angle

$$\sum_{\text{angles}} K_i^{ba} (\theta_i - \theta_o)$$



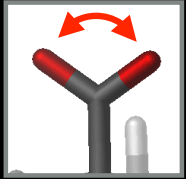
Bond Rotate

$$\sum_{\text{dihedrals}} K_i^{br} [1 - \cos(n_i \phi_i - \phi_o)]$$



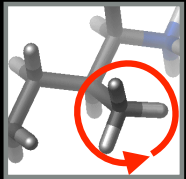
Bond Stretch

$$\sum_{\text{bonds}} K_i^{bs} (b_i - b_o)$$



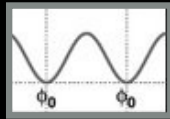
Bond Angle

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Bond Rotate

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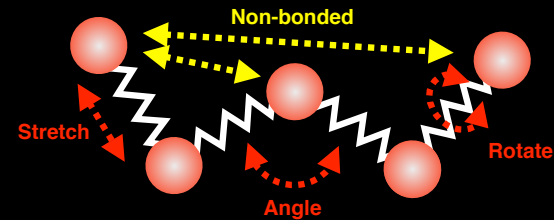
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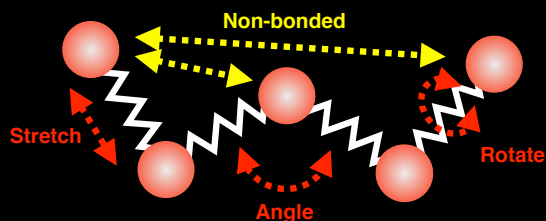
$$E_{\text{van.der.Waals}} + E_{\text{electrostatic}}$$



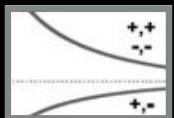
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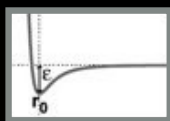
$$E_{\text{van.der.Waals}} + E_{\text{electrostatic}}$$



$$E_{electrostatic} = \sum_{pairs.i,j} \frac{q_i q_j}{\epsilon r_{ij}}$$



$$E_{van.der.Waals} = \sum_{pairs.i,j} \left[\epsilon_{ij} \left(\frac{r_{o,ij}}{r_{ij}} \right)^{12} - 2\epsilon_{ij} \left(\frac{r_{o,ij}}{r_{ij}} \right)^6 \right]$$



Total potential energy

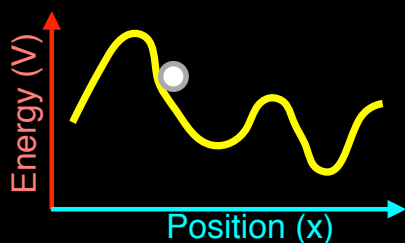
The potential energy can be given as a sum of terms for: Bond stretching, Bond angles, Bond rotations, van der Waals and Electrostatic interactions between atom pairs

$$V(R) = \begin{matrix} E_{bond.stretch} \\ + E_{bond.angle} \\ + E_{bond.rotate} \end{matrix} \left. \vphantom{\begin{matrix} E_{bond.stretch} \\ + E_{bond.angle} \\ + E_{bond.rotate} \end{matrix}} \right\} E_{bonded}$$

$$+ \begin{matrix} E_{van.der.Waals} \\ + E_{electrostatic} \end{matrix} \left. \vphantom{\begin{matrix} E_{van.der.Waals} \\ + E_{electrostatic} \end{matrix}} \right\} E_{non.bonded}$$

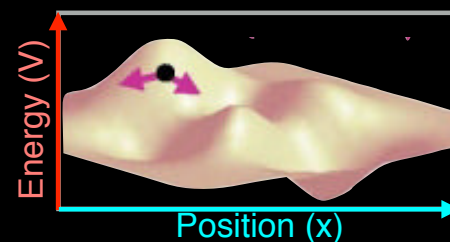
Potential energy surface

Now we can calculate the **potential energy surface** that fully describes the energy of a molecular system as a function of its geometry



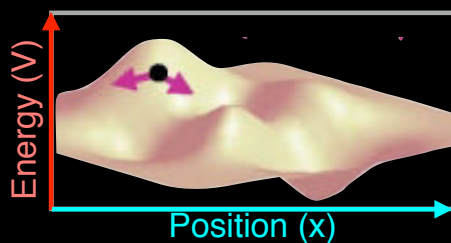
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Key concept:

Now we can calculate the **potential energy surface** that fully describes the energy of a molecular system as a function of its geometry



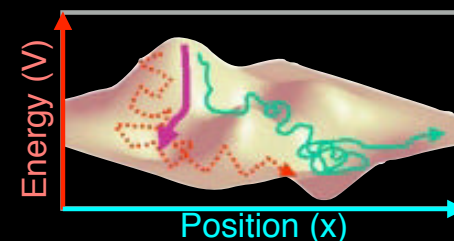
- The **forces** are the gradients of the energy

$$F(x) = -dV/dx$$

Moving Over The Energy Surface

- Energy Minimization** drops into local minimum
- Molecular Dynamics** uses thermal energy to move smoothly over surface
- Monte Carlo Moves** are random. Accept with probability:

$$\exp(-\Delta V/dx)$$



PHYSICS-ORIENTED APPROACHES

Weaknesses

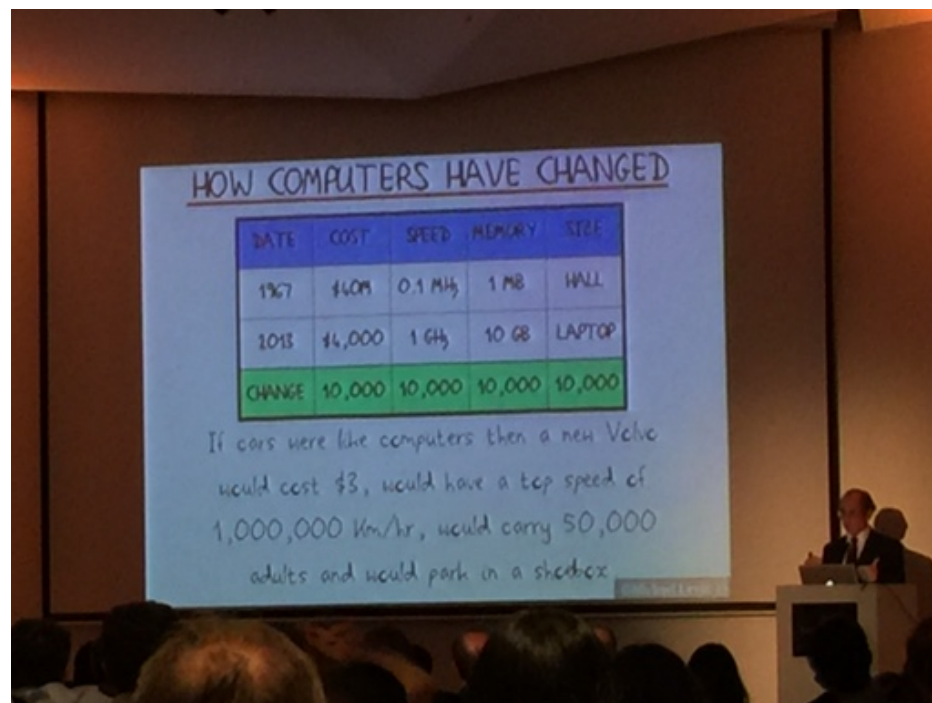
Fully physical detail becomes computationally intractable
Approximations are unavoidable
(Quantum effects approximated classically, water may be treated crudely)
Parameterization still required

Strengths

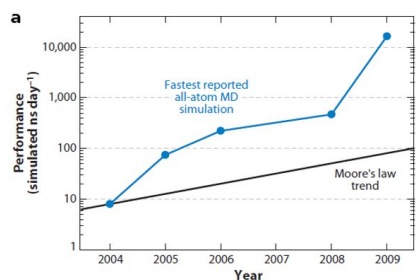
Interpretable, provides guides to design
Broadly applicable, in principle at least
Clear pathways to improving accuracy

Status

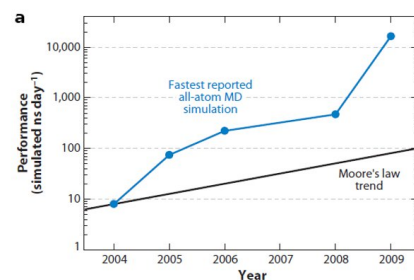
Useful, widely adopted but far from perfect
Multiple groups working on fewer, better approx
Force fields, quantum entropy, water effects
Moore's law: hardware improving



SIDE-NOTE: GPUS AND ANTON SUPERCOMPUTER



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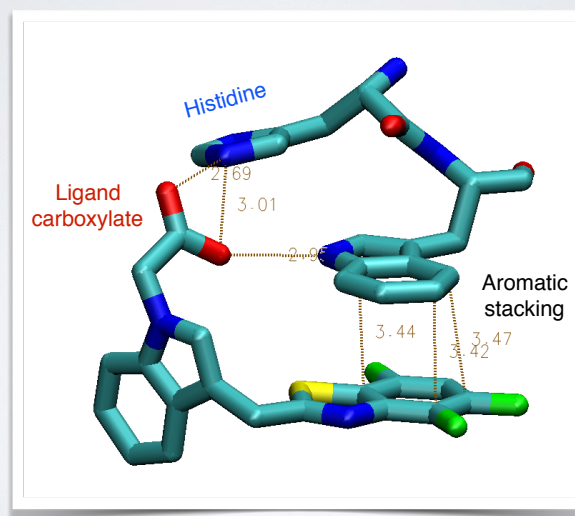
POTENTIAL FUNCTIONS DESCRIBE A SYSTEMS **ENERGY** AS A FUNCTION OF ITS **STRUCTURE**

Two main approaches:

(1). Physics-Based

(2). Knowledge-Based

KNOWLEDGE-BASED DOCKING POTENTIALS



ENERGY DETERMINES **PROBABILITY** (STABILITY)

Basic idea: Use probability as a proxy for energy



Boltzmann:
$$p(r) \propto e^{-E(r)/RT}$$



Inverse Boltzmann:
$$E(r) = -RT \ln[p(r)]$$

Example: ligand **carboxylate O** to protein **histidine N**

- Find all protein-ligand structures in the PDB with a ligand carboxylate **O**
1. For each structure, histogram the distances from **O** to every histidine **N**
 2. Sum the histograms over all structures to obtain $p(r_{O-N})$
 3. Compute $E(r_{O-N})$ from $p(r_{O-N})$

KNOWLEDGE-BASED POTENTIALS

Weaknesses

Accuracy limited by availability of data

Strengths

Relatively easy to implement
Computationally fast

Status

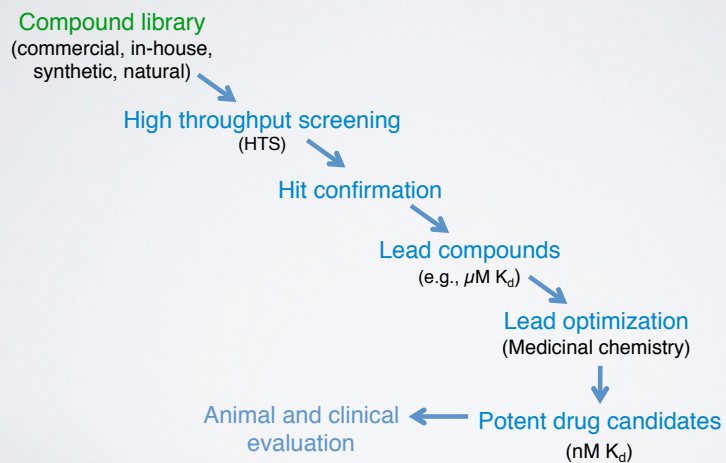
Useful, far from perfect
May be at point of diminishing returns
(not always clear how to make improvements)

Computer Aided Drug Discovery

Next Up:

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THE TRADITIONAL EMPIRICAL PATH TO DRUG DISCOVERY



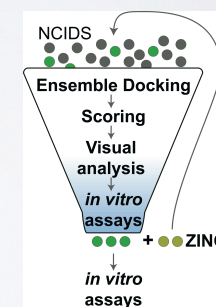
COMPUTER-AIDED LIGAND DESIGN

Aims to reduce number of compounds synthesized and assayed

Lower costs

Reduce chemical waste

Facilitate faster progress



Two main approaches:

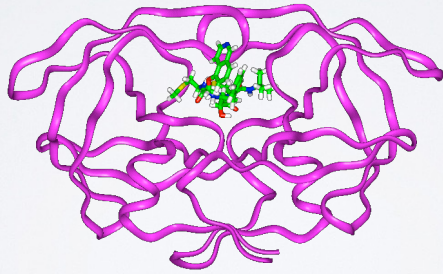
- (1). Receptor/Target-Based
- (2). Ligand/Drug-Based

Two main approaches:

- (1). Receptor/Target-Based
- (2). Ligand/Drug-Based

SCENARIO I: RECEPTOR-BASED DRUG DISCOVERY

Structure of Targeted Protein Known: **Structure-Based Drug Discovery**

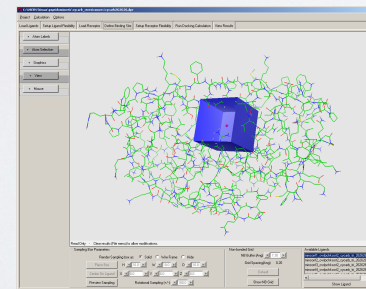


HIV Protease/KNI-272 complex

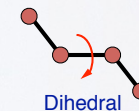
PROTEIN-LIGAND DOCKING

Structure-Based Ligand Design

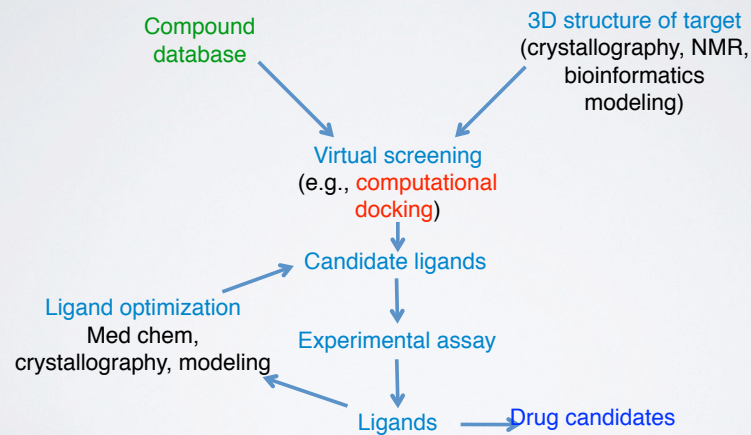
Docking software
Search for structure of lowest energy



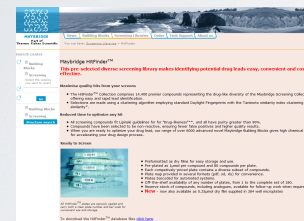
Potential function
Energy as function of structure



STRUCTURE-BASED VIRTUAL SCREENING



COMPOUND LIBRARIES



Commercial
(in-house pharma)



Government (NIH)



Academia

COMMON SIMPLIFICATIONS USED IN PHYSICS-BASED DOCKING

Quantum effects approximated classically

Protein often held rigid

Configurational entropy neglected

Influence of water treated crudely

Do it Yourself!

Hand-on time!

https://bioboot.github.io/bggn213_S19/lectures/#12

You can use the classroom computers or your own laptops. If you are using your laptops then you will need to install **MGLTools**

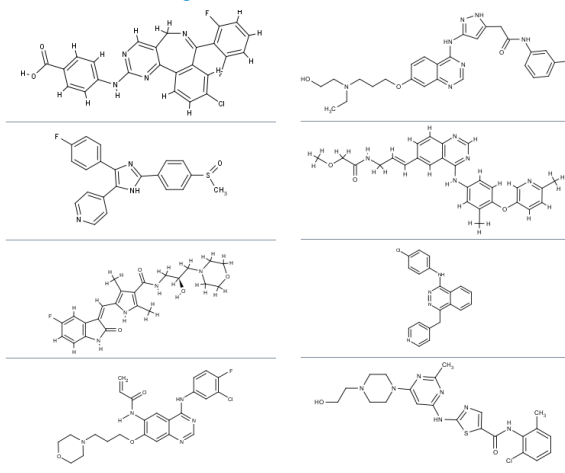
Two main approaches:

- (1). Receptor/Target-Based
- (2). Ligand/Drug-Based

Scenario 2

Structure of Targeted Protein Unknown:
Ligand-Based Drug Discovery

e.g. MAP Kinase Inhibitors



Using knowledge of existing inhibitors to discover more

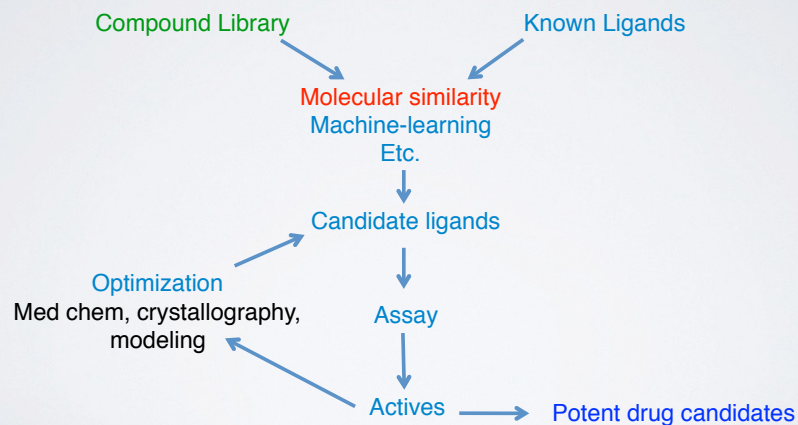
Why Look for Another Ligand if You Already Have Some?

Experimental screening generated some ligands, but they don't bind tightly enough

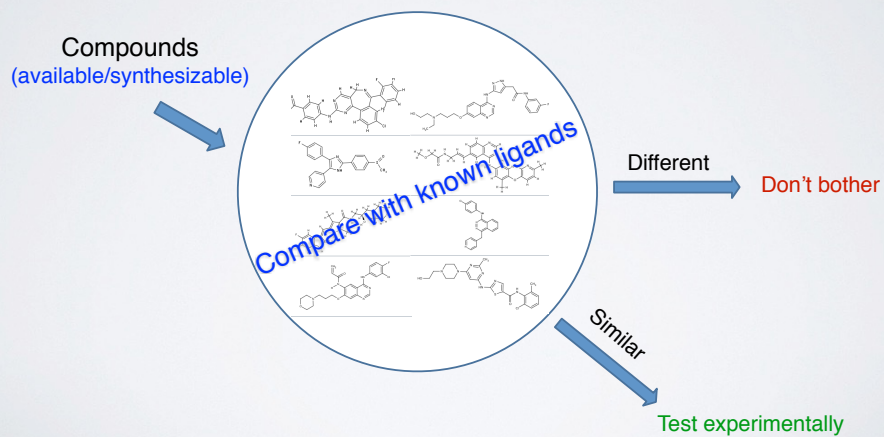
A company wants to work around another company's chemical patents

An high-affinity ligand is toxic, is not well-absorbed, difficult to synthesize etc.

LIGAND-BASED VIRTUAL SCREENING



CHEMICAL SIMILARITY LIGAND-BASED DRUG-DISCOVERY



CHEMICAL FINGERPRINTS BINARY STRUCTURE KEYS

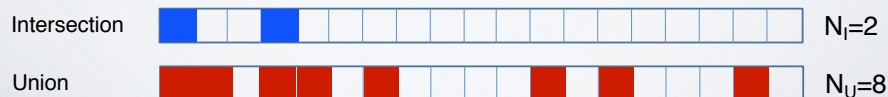


CHEMICAL SIMILARITY FROM FINGERPRINTS



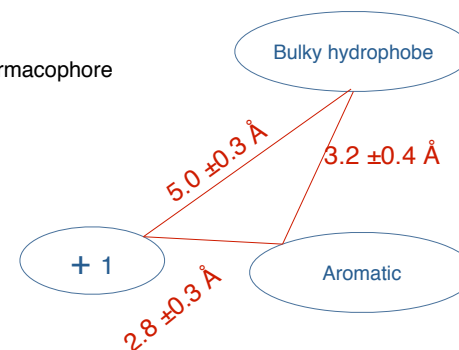
Tanimoto Similarity
(or Jaccard Index), T

$$T \equiv \frac{N_I}{N_U} = 0.25$$



Pharmacophore Models Φάρμακο (drug) + Φορά (carry)

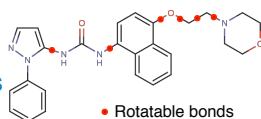
A 3-point pharmacophore



Molecular Descriptors More abstract than chemical fingerprints

Physical descriptors

- molecular weight
- charge
- dipole moment
- number of H-bond donors/acceptors
- number of rotatable bonds
- hydrophobicity (log P and clogP)



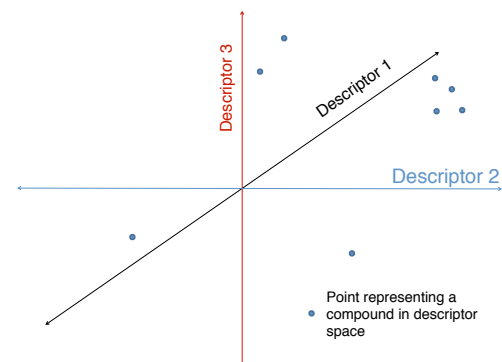
Topological

- branching index
- measures of linearity vs interconnectedness

Etc. etc.

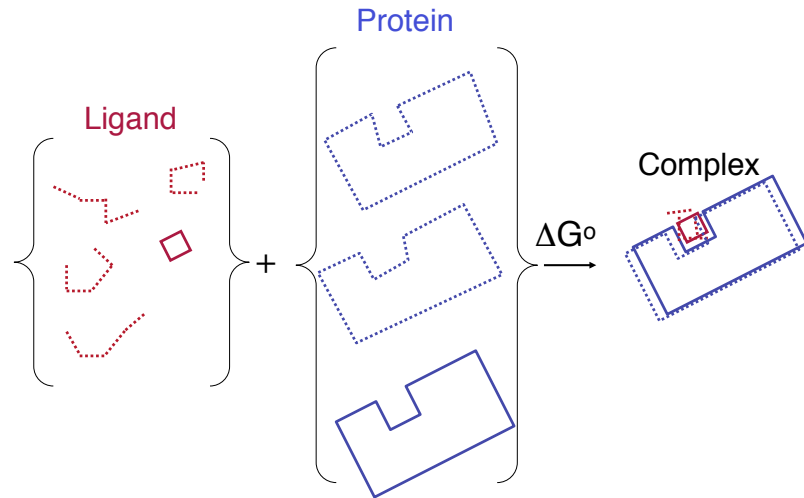
A High-Dimensional “Chemical Space”

Each compound is a point in an n-dimensional space
Compounds with similar properties are near each other



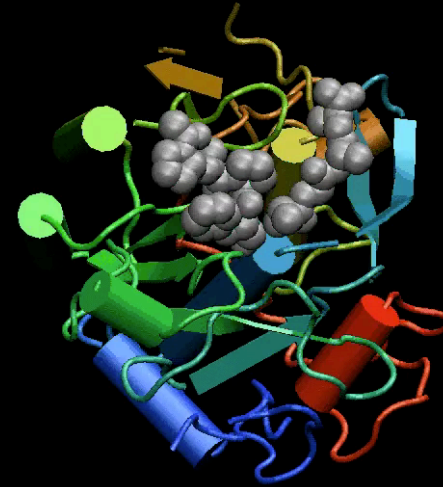
Apply multivariate statistics and machine learning for descriptor-selection. (e.g. partial least squares, PCA, support vector machines, random forest, deep learning etc.)

Proteins and Ligand are Flexible



NMA (Normal Mode Analysis) is a bioinformatics method to predict the intrinsic dynamics of biomolecules

Do it Yourself!



https://bioboot.github.io/bggn213_S19/lectures/#12

NMA in Bio3D

- Normal Mode Analysis (NMA) is a bioinformatics method that can predict the major motions of biomolecules.

```
pdb <- read.pdb("1hel")
modes <- nma(pdb)
m7 <- mktmj(modes, mode=7, file="mode_7.pdb")
```

Then you can open the resulting `mode_7.pdb` file in **VMD**
- Use "TUBE" representation and hit the play button...

Or use the `bio3d.view view()` function

```
library("bio3d.view")
view(m7, col=vec2color(rmsf(m7)))
```

Reference Slides

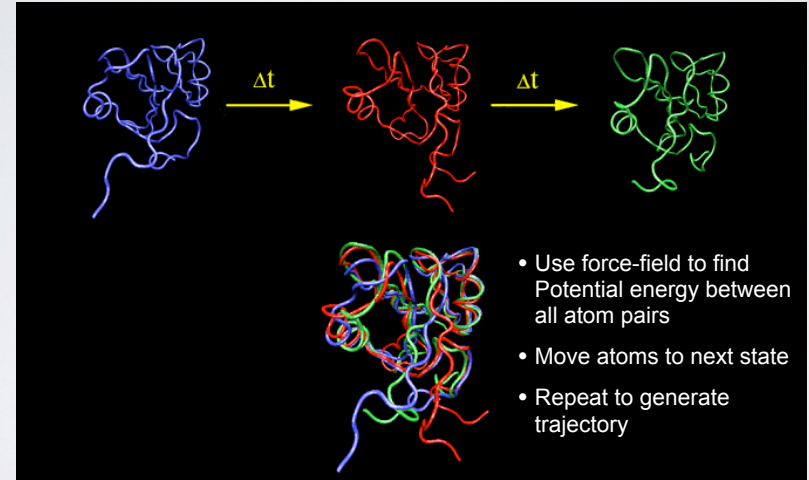
Molecular Dynamics (MD) and Normal Mode Analysis (NMA) Background and Cautionary Notes

[[Muddy Point Assessment](#)]

PREDICTING FUNCTIONAL DYNAMICS

- Proteins are intrinsically flexible molecules with internal motions that are often intimately coupled to their biochemical function
 - E.g. ligand and substrate binding, conformational activation, allosteric regulation, etc.
- Thus knowledge of dynamics can provide a deeper understanding of the mapping of structure to function
 - Molecular dynamics (MD) and normal mode analysis (NMA) are two major methods for predicting and characterizing molecular motions and their properties

MOLECULAR DYNAMICS SIMULATION



McCammon, Gelin & Karplus, *Nature* (1977)

[See: <https://www.youtube.com/watch?v=ui1ZysMFcKk>]

- ▶ Divide **time** into discrete (~1fs) **time steps** (Δt)
(for integrating equations of motion, see below)



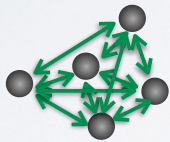
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- ▶ At each time step calculate pair-wise atomic **forces** ($F(t)$) (by evaluating force-field gradient)



Nucleic motion described classically

$$m_i \frac{d^2}{dt^2} \vec{R}_i = -\vec{\nabla}_i E(\vec{R})$$

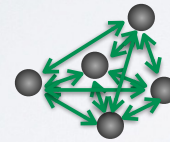
Empirical force field

$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_j(\vec{R})$$

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$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_j(\vec{R})$$

- ▶ Use the forces to calculate velocities and move atoms to new positions (by integrating numerically via the “leapfrog” scheme)



$$v\left(t + \frac{\Delta t}{2}\right) = v\left(t - \frac{\Delta t}{2}\right) + \frac{F(t)}{m} \Delta t$$

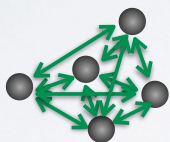
$$r(t + \Delta t) = r(t) + v\left(t + \frac{\Delta t}{2}\right) \Delta t$$

BASIC ANATOMY OF A MD SIMULATION

- ▶ Divide **time** into discrete (~1fs) **time steps** (Δt) (for integrating equations of motion, see below)



- ▶ At each time step calculate pair-wise atomic **forces** ($F(t)$) (by evaluating force-field gradient)



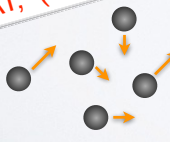
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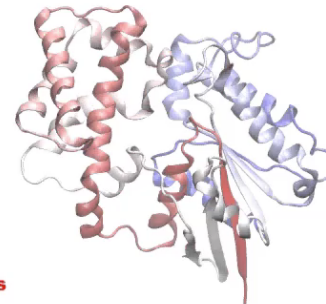
$$v\left(t + \frac{\Delta t}{2}\right) = v\left(t - \frac{\Delta t}{2}\right) + \frac{F(t)}{m} \Delta t$$

$$r(t + \Delta t) = r(t) + v\left(t + \frac{\Delta t}{2}\right) \Delta t$$

REPEAT, (iterate many, many times... 1ms = 10¹² time steps)

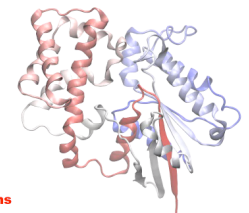
MD Prediction of Functional Motions

Accelerated MD simulation of nucleotide-free transducin alpha subunit



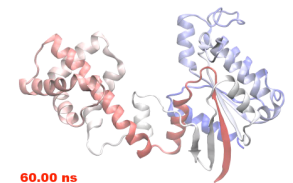
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0.00 ns

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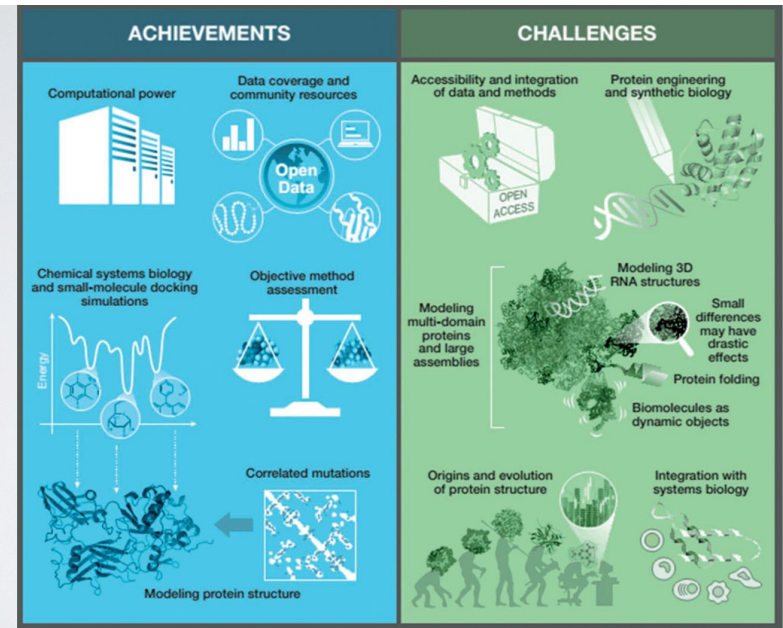
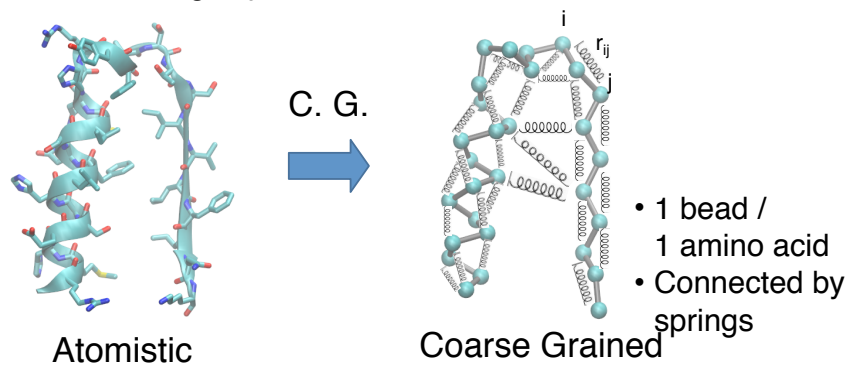


60.00 ns

Yao and Grant, Biophys J. (2013)

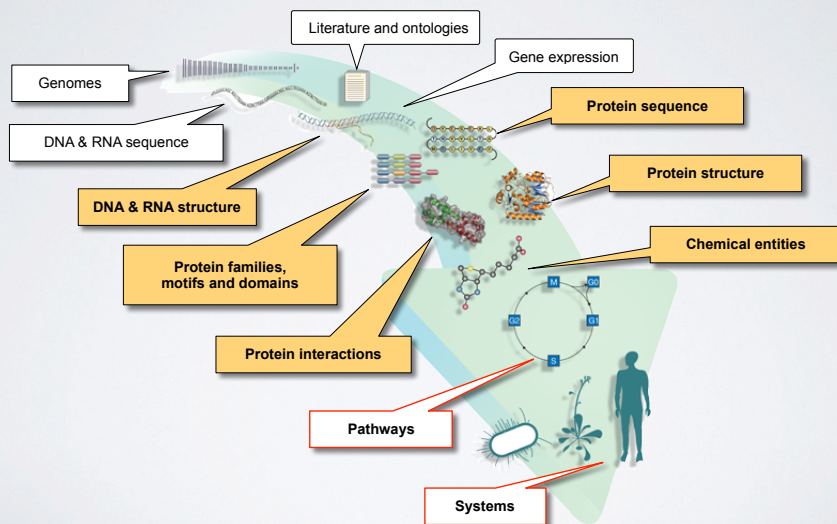
COARSE GRAINING: **NORMAL MODE ANALYSIS** (NMA)

- MD is still time-consuming for large systems
- Elastic network model NMA (ENM-NMA) is an example of a lower resolution approach that finishes in seconds even for large systems.



Ilan Samish et al. *Bioinformatics* 2015;31:146-150

INFORMING SYSTEMS BIOLOGY?



SUMMARY

- Structural bioinformatics is computer aided structural biology
- Described major motivations, goals and challenges of structural bioinformatics
- Reviewed the fundamentals of protein structure
- Explored how to use R to perform structural bioinformatics analysis!
- Introduced both physics and knowledge based modeling approaches for describing the structure, energetics and dynamics of proteins computationally
- Introduced both structure and ligand based bioinformatics approaches for drug discovery and design

[[Muddy Point Assessment](#)]

CAUTIONARY NOTES

- **A model is never perfect**

A model that is not quantitatively accurate in every respect does not preclude one from establishing results relevant to our understanding of biomolecules as long as the biophysics of the model are properly understood and explored.

- **Calibration of parameters is an ongoing imperfect process**

Questions and hypotheses should always be designed such that they do not depend crucially on the precise numbers used for the various parameters.

- **A computational model is rarely universally right or wrong**

A model may be accurate in some regards, inaccurate in others. These subtleties can only be uncovered by comparing to all available experimental data.