



BIMM 143
Unsupervised Learning
Lecture 8
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Recap of Lecture 7

- Reviewed the major steps for R function writing
- Introduced the CRAN & Bioconductor repositories for packages that extend R functionality
- Explored installing and using the ggplot2, bio3d, rgl, rentrez, igraph, blogdown, shiny, and other packages
- Discussed how to judge the utility, usability and development status of R packages.

[\[Muddy Point Feedback Link\]](#)

Today's Menu

- Introduction to machine learning
 - Unsupervised, supervised and reinforcement learning
- Clustering
 - K-means clustering
 - Hierarchical clustering
 - Heatmap representations
- Dimensionality reduction, visualization and 'structure' analysis
 - Principal Component Analysis (PCA)
- Hands-on application to cell classification

Types of machine learning

- Unsupervised learning
 - Finding structure in unlabeled data
- Supervised learning
 - Making predictions based on labeled data
 - Predictions like regression or classification
- Reinforcement learning
 - Making decisions based on past experience

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 - ➔ Imagine you had data that you could plot along a line and you knew you had to put them into $k=3$ "clusters" (e.g. data from three types of tumor cells)



k-means clustering algorithm

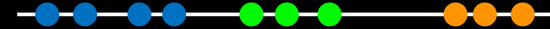
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- You define k the number of clusters!
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Here your eyes can clearly see 3 natural groupings

k-means clustering algorithm

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- You define k the number of clusters!
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Here your eyes can clearly see 3 natural groupings
How does k-means attempt to define this grouping?

Step 1.

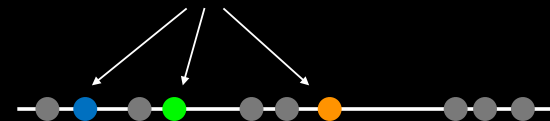
Select k (the number of clusters)



Step 2.

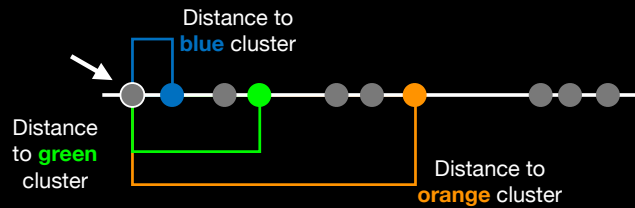
Select $k=3$ distant data points at random

These are the initial clusters



Step 3.

Measure distance between the 1st point and the $k=3$ initial clusters



Step 4.

Assign the 1st point to the nearest cluster



Step 5.

Update cluster centers

Calculate the mean value for the blue cluster including the new point



Step 6.

Assign next point to closest cluster

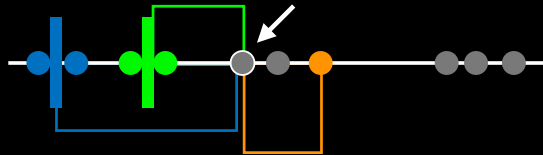
Use updated cluster centers for distance calculation



Step 7.

Update cluster centers and move to next point

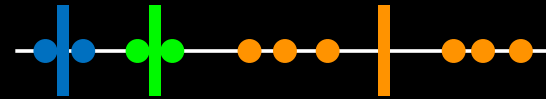
Use updated cluster centers for distance calculation



Step 8.

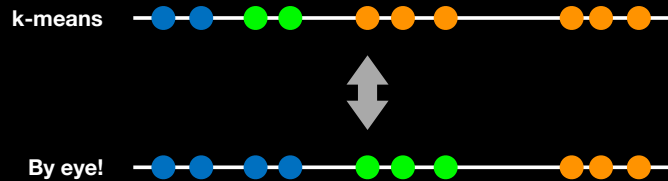
Repeat for each point

Each time updating cluster centers



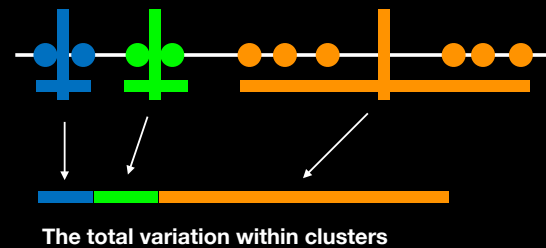
Hmm....

Here the k-means result does not look as good as what we were able to do by eye!



Step 9.

Assess the quality of the clustering by adding up the variation within each cluster

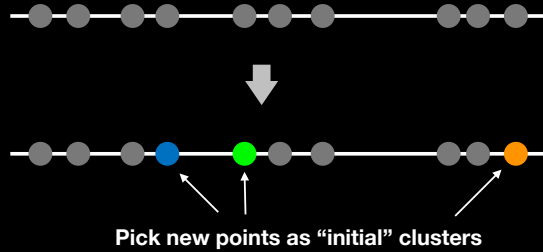


K-means keeps track of these clusters and their total **variance** and then does the whole thing over again with different starting points

Step 10.

Repeat with different starting points

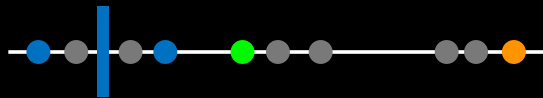
Back to the beginning and do all steps over again...



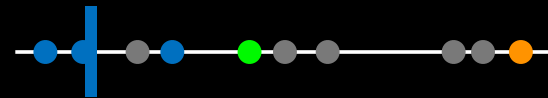
...Pick $k=3$ initial clusters and add the remaining points to the cluster with the nearest mean, recalculating the mean each time a new point is added...



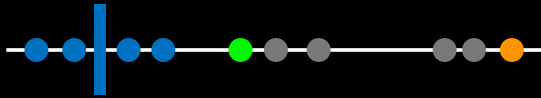
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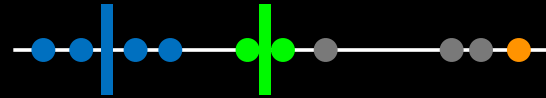
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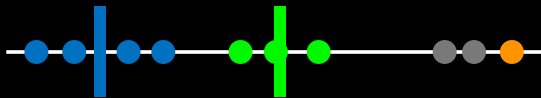
...Pick $k=3$ initial clusters and add the remaining points to the cluster with the nearest mean, recalculating the mean each time a new point is added...



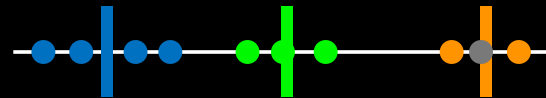
...Pick $k=3$ initial clusters and add the remaining points to the cluster with the nearest mean, recalculating the mean each time a new point is added...



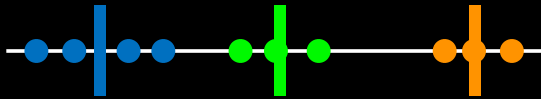
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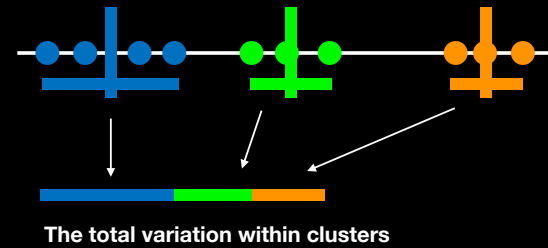
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Now the data are all assigned to clusters, we again sum the variation within each cluster



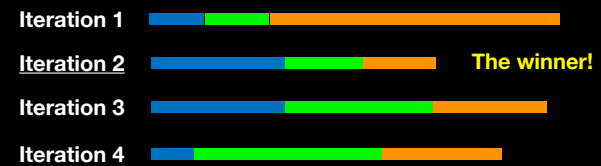
Step 10.

Repeat again with different starting points



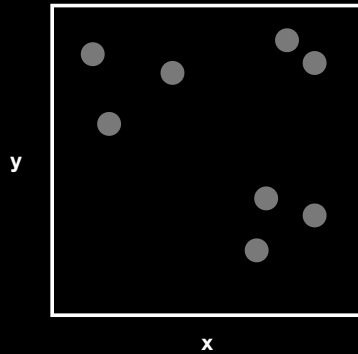
After several iterations k-means clustering knows it has the *best clustering so-far* based on the smallest total variation with clusters.

However, it does not know if it has found the *best overall*. So it will perform several more iterations with different starting points...

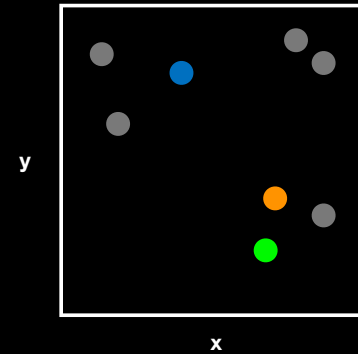


The total variation within clusters

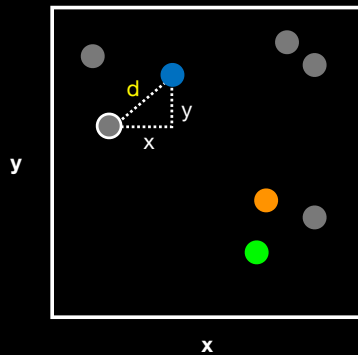
What if we have more dimensions?



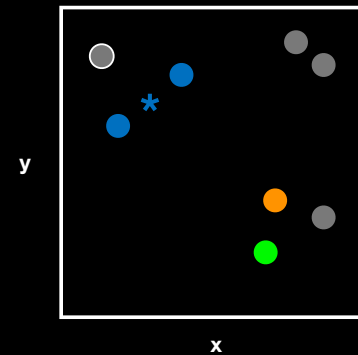
Just like before, we pick 3 random points...



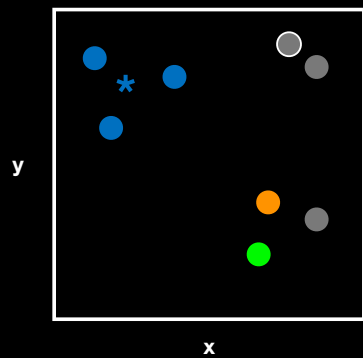
...and use the **Euclidean distance**.
In 2 dimensions the Euclidean distance is the same as the Pythagorean theorem $d = \sqrt{x^2 + y^2}$



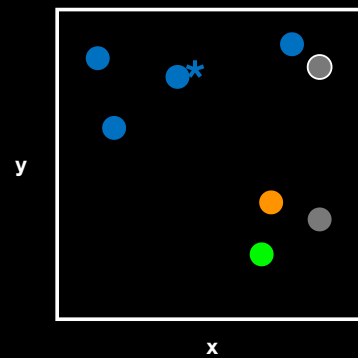
...assign point to nearest cluster and update cluster center *



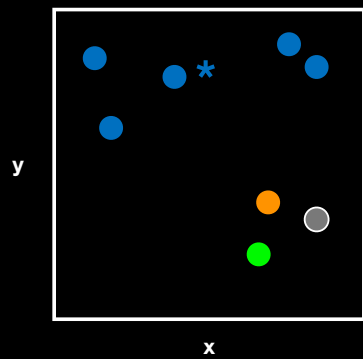
...and continue



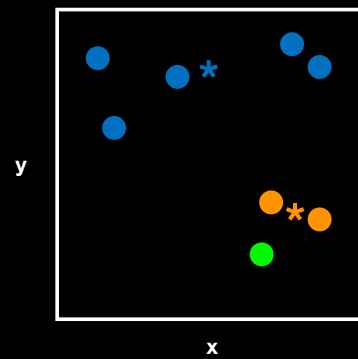
...and continue



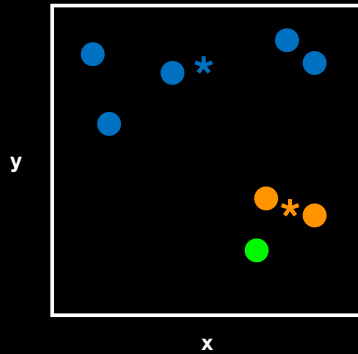
...and continue



...and continue



Again we have to use a number of different starting conditions before deciding on a good clustering!



What if we have even more dimensions?

Cell Samples

	#1	#2	#3
Gene 1	12	6	-13
Gene 2	-7	13	10
Gene 3	8	6	-9
Gene 4	9	5	-11
Gene 5	-3	1	6
Gene 6	10	4	-8

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x **y** **z**

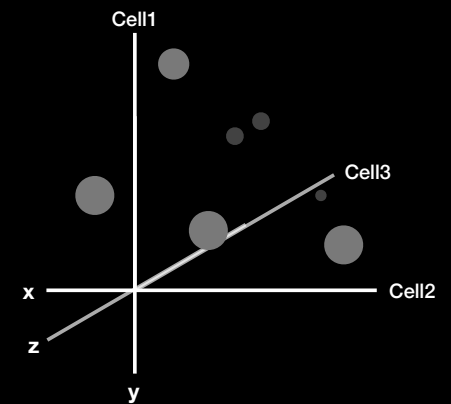
We could simply plot them by relabeling the cell samples as **x**, **y**, and **z** (i.e. a 3D plot)

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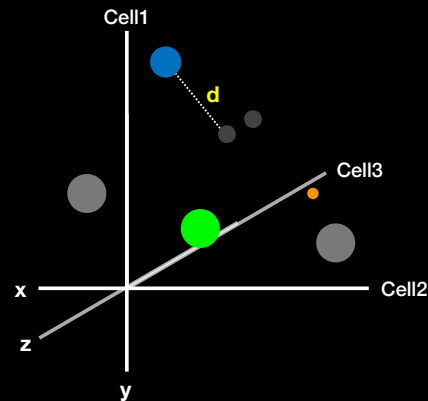


...and go through exactly the same procedure with initial cluster assignment followed by distance calculation etc...

$$d = \sqrt{x^2 + y^2 + z^2}$$

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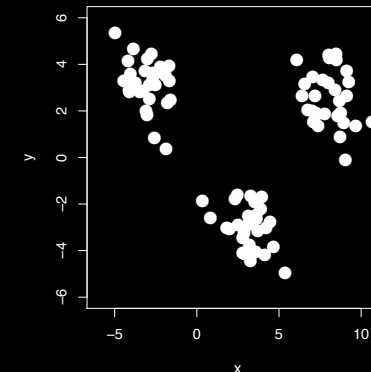
Of course we don't actually need to plot anything.

We can just calculate the Euclidean distance along any number of dimensions and perform our k-means clustering in the same way.

k-means in R

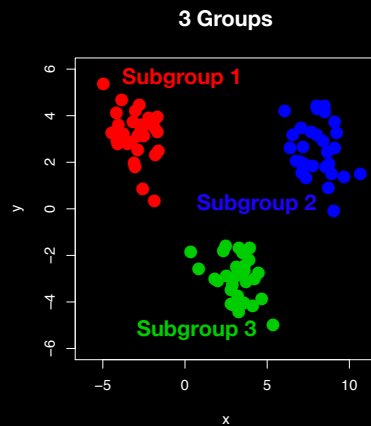
```
# k-means algorithm with 3 centers, run 20 times  
kmeans(x, centers= 3, nstart= 20)
```

- Input **x** is a numeric matrix, or data.frame, with one observation per row, one feature per column
- k-means has a random component
- Run algorithm multiple times to improve odds of the best model



Model selection

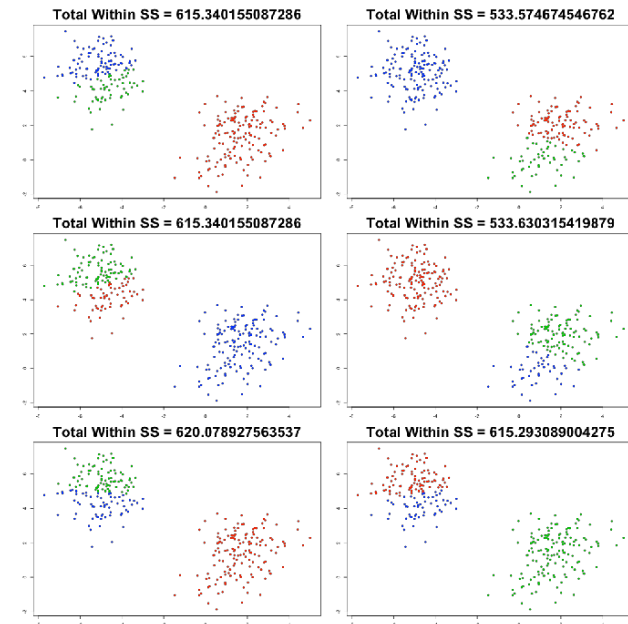
- Recall k-means has a random component
- Best outcome is based on total within cluster sum of squares:
 - For each cluster
 - For each observation in the cluster
 - Determine squared distance from observation to cluster center
 - Sum all of them together

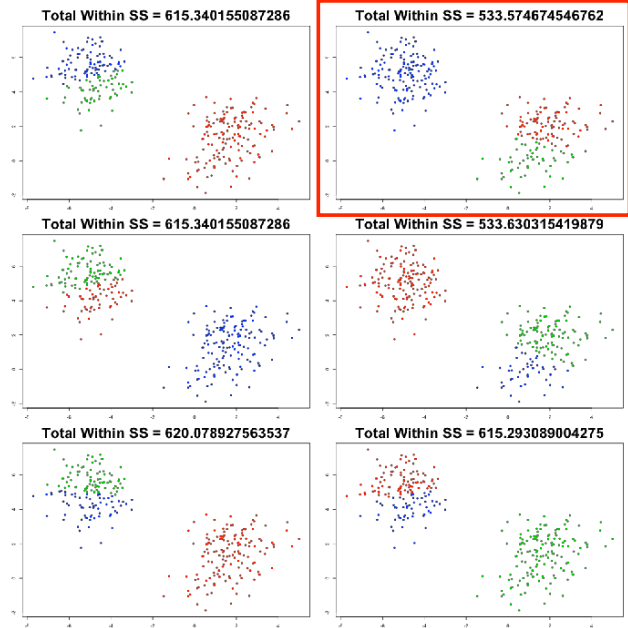


Model selection

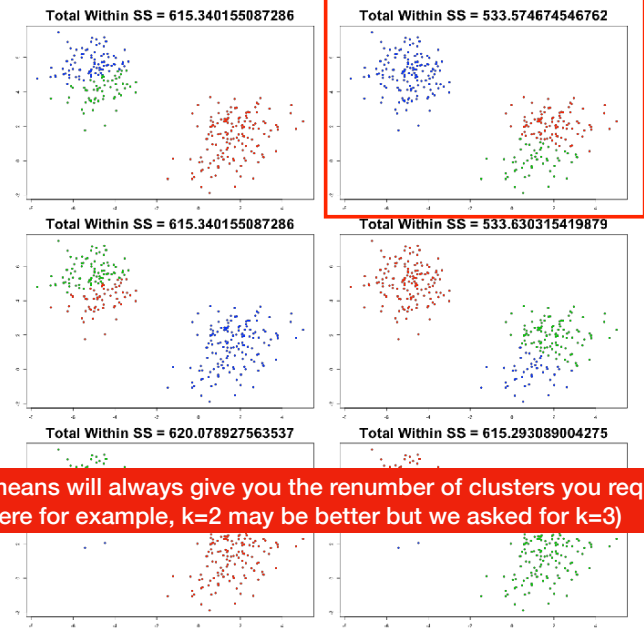
```
# k-means algorithm with 5 centers, run 20 times  
kmeans(x, centers=5, nstart=20)
```

- Running algorithm multiple times (i.e. setting `nstart`) helps find the global minimum total within cluster sum of squares
- Increasing the default value of `nstart` is often sensible



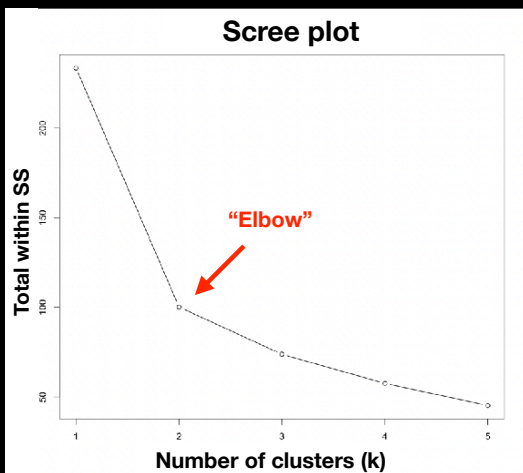


Winner has the smallest within cluster SS



Note. k-means will always give you the renumber of clusters you request! (Here for example, k=2 may be better but we asked for k=3)

Determining number of clusters



Trial and error is not the best approach

Systematically try a range of different k values and plot a “scree plot”.

Here there is a large reduction in SS with **k=2** but after that the values do not go down as quickly!

Your Turn!

```
# Generate some example data for clustering
tmp <- c(rnorm(30,-3), rnorm(30,3))
x <- cbind(x=tmp, y=rev(tmp))

plot(x)

# Use the kmeans() function setting k to 2 and nstart=20

# Inspect/print the results

# Q. How many points are in each cluster?
# Q. What 'component' of your result object details
#   - cluster size?
#   - cluster assignment/membership?
#   - cluster center?

# Plot x colored by the kmeans cluster assignment and
#   add cluster centers as blue points
# Q. Repeat for k=3, which has the lower tot.withinss?
```

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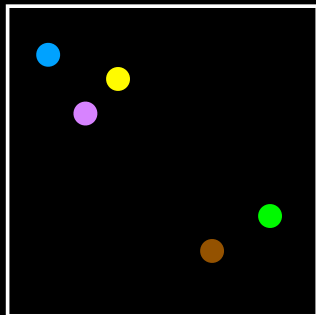
Hierarchical clustering

- Number of clusters is not known ahead of time
- Two kinds of hierarchical clustering:
 - bottom-up
 - top-down

Hierarchical clustering

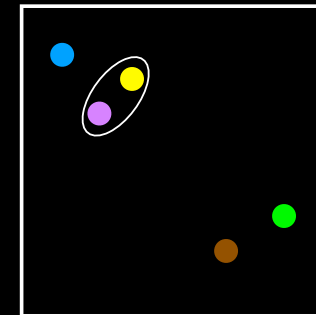
Simple example:

5 clusters: Each point starts as it's own "cluster"!



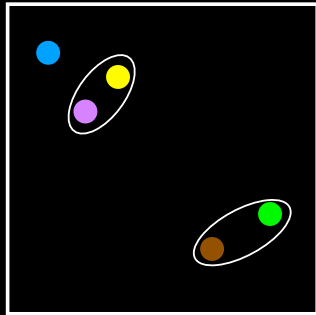
Hierarchical clustering

4 clusters



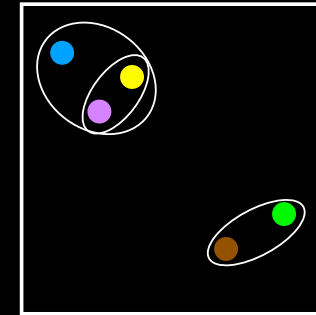
Hierarchical clustering

3 clusters



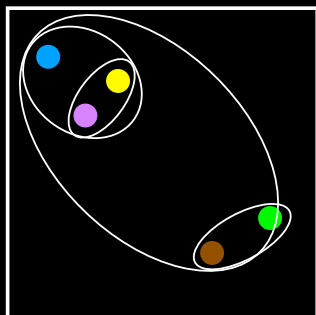
Hierarchical clustering

2 clusters



Hierarchical clustering

End: 1 cluster



Hierarchical clustering in R

```
# First we need to calculate point (dis)similarity
# as the Euclidean distance between observations
dist_matrix <- dist(x)

# The hclust() function returns a hierarchical
# clustering model
hc <- hclust(d = dist_matrix)

# the print method is not so useful here
hc

Call:
hclust(d = dist_matrix)

Cluster method : complete
Distance       : euclidean
Number of objects: 60
```


Lets have a closer look...

```
# Our input is a distance matrix from the dist()
# function. Lets make sure we understand it first
dist_matrix <- dist(x)

dim(dist_matrix)
NULL

View( as.matrix(dist_matrix) )

dim(x)
[1] 60 2

dim( as.matrix(dist_matrix) )
[1] 60 60

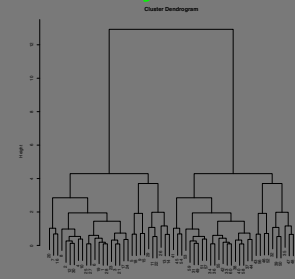
# Note, symmetrical pairwise distance matrix
```

Interpreting results

```
# Create hierarchical cluster model: hc
hc <- hclust(dist(x))

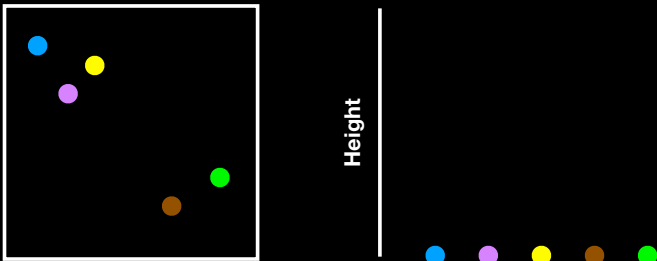
# We can plot the results as a dendrogram
plot(hc)
```

```
# What do you notice?
# Does the dendrogram
# make sense based on
# your knowledge of x?
```



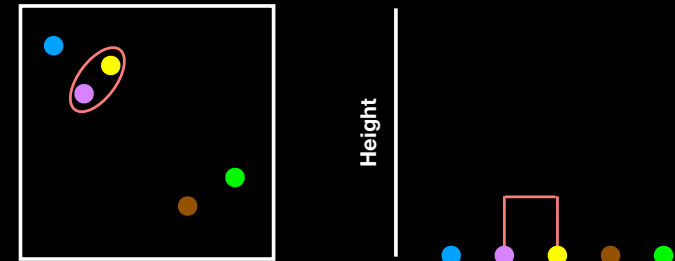
Dendrogram

- Tree shaped structure used to interpret hierarchical clustering models



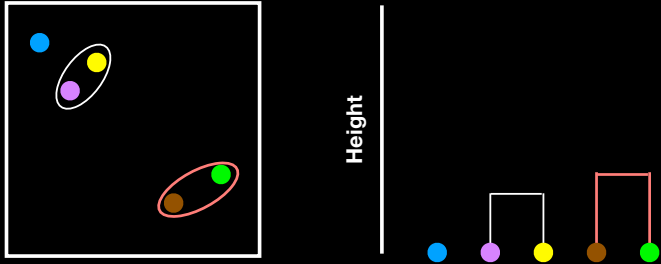
Dendrogram

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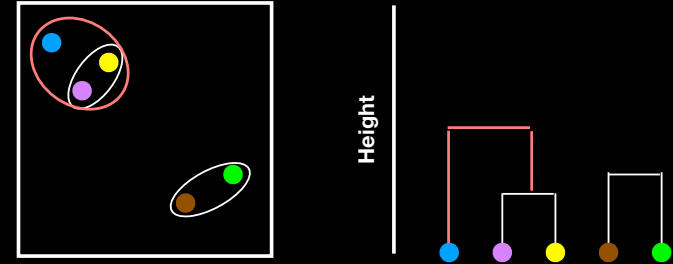
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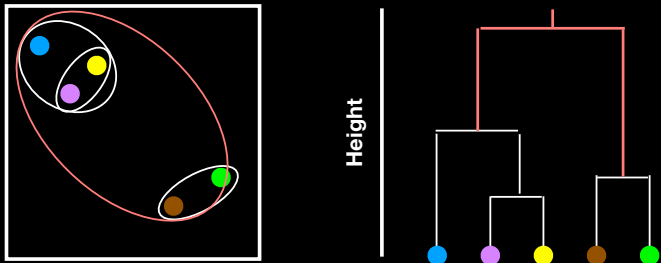
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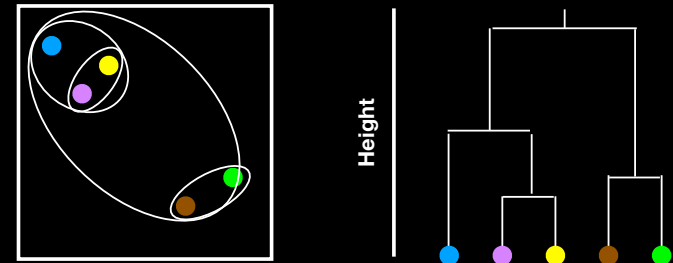
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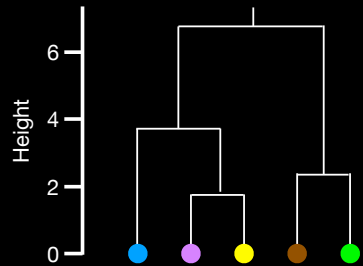
Dendrogram

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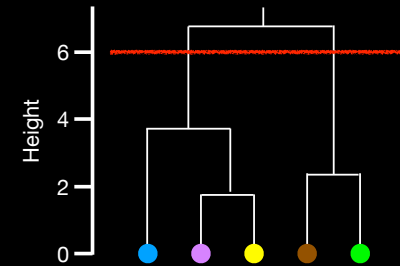
Dendrogram plotting in R

```
# Draws a dendrogram  
plot(hc)
```



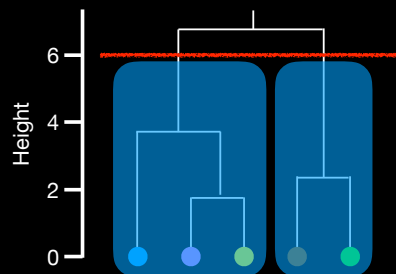
Dendrogram plotting in R

```
# Draws a dendrogram  
plot(hc)  
abline(h=6, col="red")
```



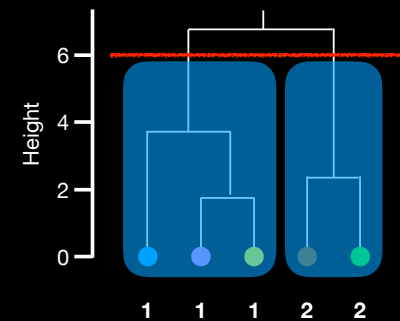
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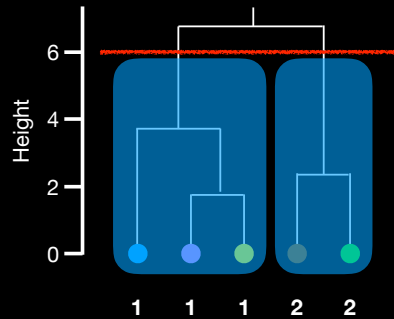
Dendrogram plotting in R

```
# Draws a dendrogram  
plot(hc)  
abline(h=6, col="red")  
cutree(hc, h=6) # Cut by height h  
[1] 1,1,1,2,2
```



Dendrogram plotting in R

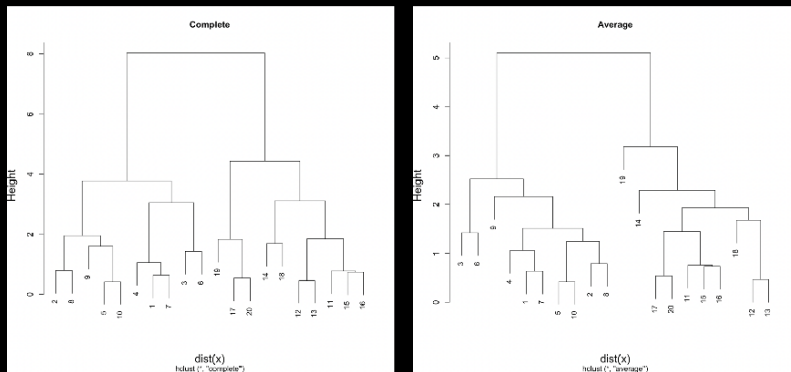
```
# Draws a dendrogram
plot(hc)
abline(h=6, col="red")
cutree(hc, k=2) # Cut into k grps
[1] 1,1,1,2,2
```



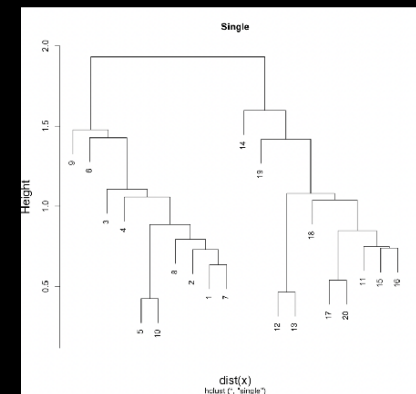
Linking clusters in hierarchical clustering

- How is distance between clusters determined?
- There are four main methods to determine which cluster should be linked:
 - **Complete:** pairwise similarity between all observations in cluster 1 and cluster 2, and uses largest of similarities
 - **Single:** same as above but uses smallest of similarities
 - **Average:** same as above but uses average of similarities
 - **Centroid:** finds centroid of cluster 1 and centroid of cluster 2, and uses similarity between two centroids

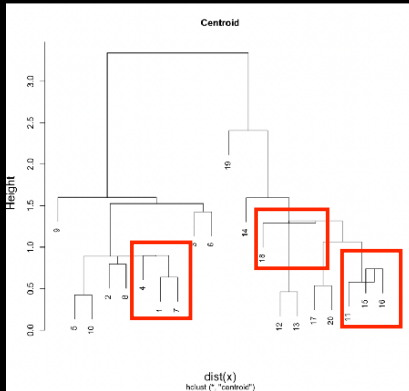
Linking methods: complete and average



Linking method: single



Linking method: centroid



Linkage in R

```
# Using different hierarchical clustering methods
hc.complete <- hclust(d, method="complete")

hc.average <- hclust(d, method="average")

hc.single <- hclust(d, method="single")
```

Side-Note: Practical matters

- Data on different scales can cause undesirable results in clustering methods
- Solution is to scale data so that features have same mean and standard deviation
 - ➔ Subtract mean of a feature from all observations
 - ➔ Divide each feature by the standard deviation of the feature
 - ➔ Normalized features have a mean of zero and a standard deviation of one

Side-Note: Practical matters

```
# Check if scaling is necessary
colMeans(x) # x is a data matrix
[1] -0.1337828  0.0594019

apply(x, 2, sd)
[1] 1.974376 2.112357

# New matrix with col mean of 0 and sd of 1
scaled_x <- scale(x)

colMeans(scaled_x)
[1] 2.775558e-17 3.330669e-17

apply(scaled_x, 2, sd)
[1] 1 1
```

Your Turn!

```
# Step 1. Generate some example data for clustering
x <- rbind(
  matrix(rnorm(100, mean=0, sd = 0.3), ncol = 2), # c1
  matrix(rnorm(100, mean = 1, sd = 0.3), ncol = 2), # c2
  matrix(c(rnorm(50, mean = 1, sd = 0.3),
           rnorm(50, mean = 0, sd = 0.3)), ncol = 2))
colnames(x) <- c("x", "y")

# Step 2. Plot the data without clustering
plot(x)

# Step 3. Generate colors for known clusters
# (just so we can compare to hclust results)
col <- as.factor( rep(c("c1", "c2", "c3"), each=50) )

plot(x, col=col)

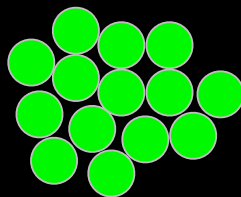
# Q. Use the dist(), hclust(), plot() and outtree()
#     functions to return 2 and 3 clusters
# Q. How does this compare to your known 'col' groups?
```

Today's Menu

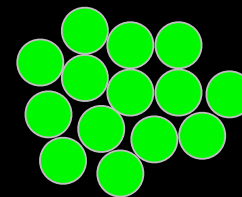
- Introduction to machine learning
 - Unsupervised, supervised and reinforcement learning
- Clustering
 - K-means clustering
 - Hierarchical clustering
 - Heatmap representations
- Dimensionality reduction, visualization and 'structure' analysis
 - Principal Component Analysis (PCA)
- Hands-on application to cell classification

PCA: The absolute basics

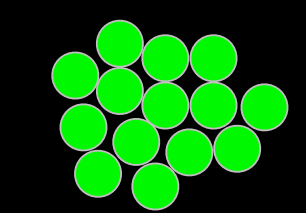
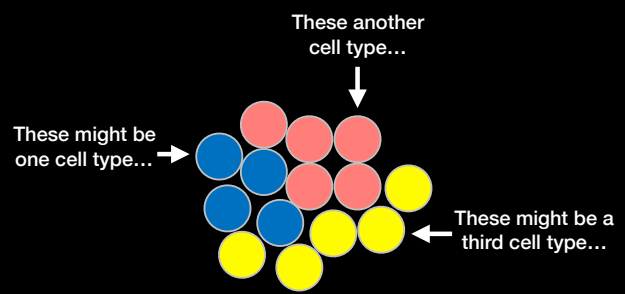
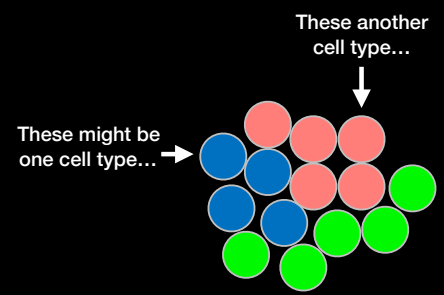
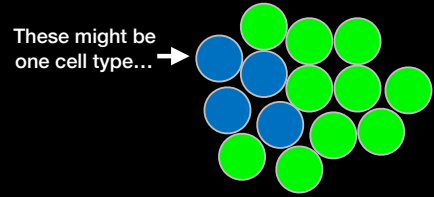
Bunch of "normal" cells



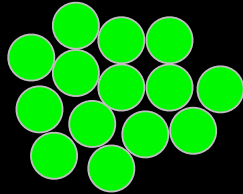
Bunch of "normal" cells



Even though they look the same we suspect that there are differences...



Unfortunately we can't observe
the differences visually



Unfortunately we can't observe the differences visually

So we sequence the mRNA in each cell to identify which genes are active and at what levels.

Here is the data...

	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Each column shows how much each gene is transcribed in each cell

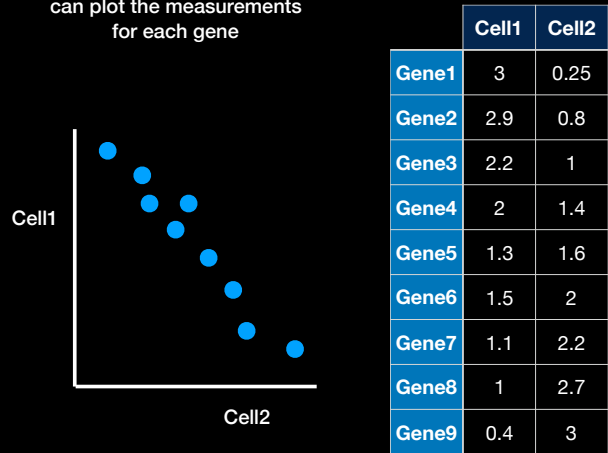
Here is the data...

	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

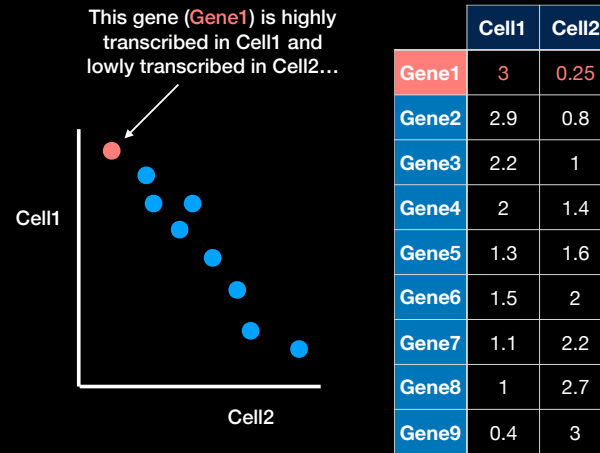
For now lets consider only two cells

	Cell1	Cell2
Gene1	3	0.25
Gene2	2.9	0.8
Gene3	2.2	1
Gene4	2	1.4
Gene5	1.3	1.6
Gene6	1.5	2
Gene7	1.1	2.2
Gene8	1	2.7
Gene9	0.4	3

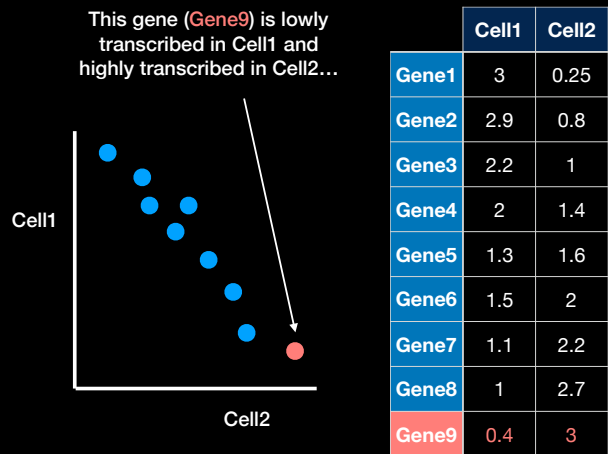
We have just 2 cells so we can plot the measurements for each gene



This gene (**Gene1**) is highly transcribed in Cell1 and lowly transcribed in Cell2...

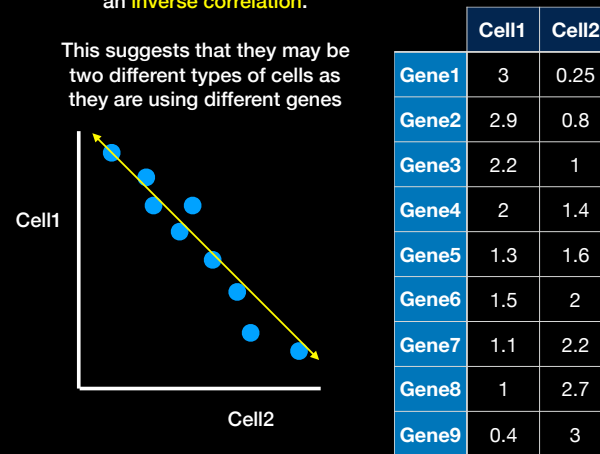


This gene (**Gene9**) is lowly transcribed in Cell1 and highly transcribed in Cell2...



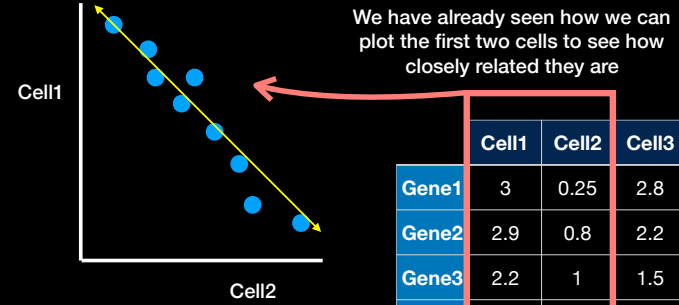
In general, Cell1 and Cell2 have an **inverse correlation**.

This suggests that they may be two different types of cells as they are using different genes

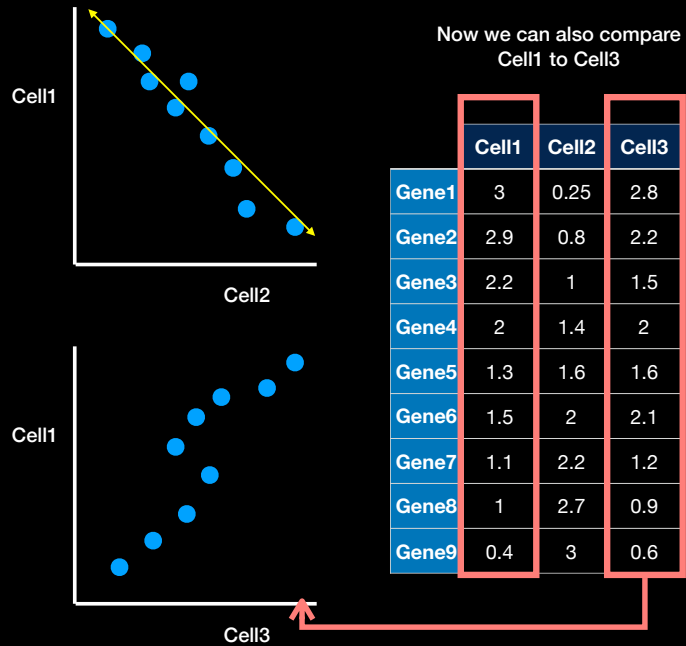


Now lets imagine there are three cells

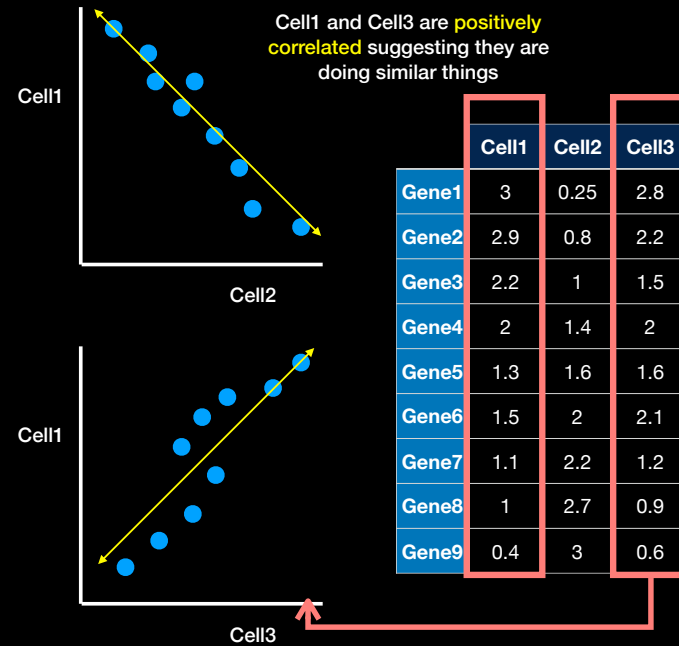
	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6



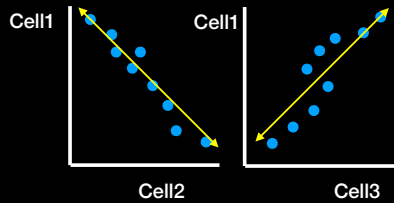
	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6



	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6



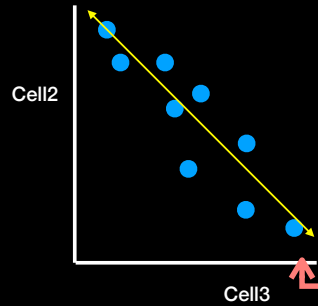
	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6



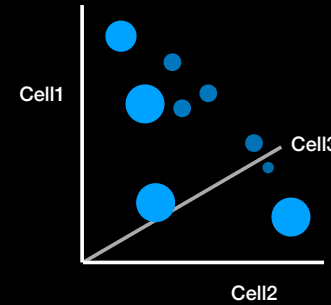
We can also compare Cell2 to Cell3...

	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6

The **inverse correlation** suggests that Cell2 is doing something different from Cell3



Alternatively, we could try to plot all 3 cells at once on a 3-dimensional graph.

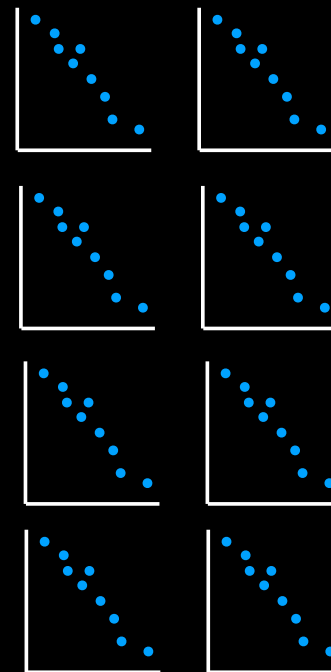


	Cell1	Cell2	Cell3
Gene1	3	0.25	2.8
Gene2	2.9	0.8	2.2
Gene3	2.2	1	1.5
Gene4	2	1.4	2
Gene5	1.3	1.6	1.6
Gene6	1.5	2	2.1
Gene7	1.1	2.2	1.2
Gene8	1	2.7	0.9
Gene9	0.4	3	0.6

But what if we have 4 or more Cells?

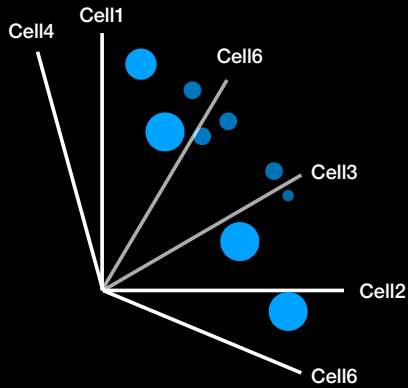
	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
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Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Draw lots of 2 cell plots and try to make sense of them all?

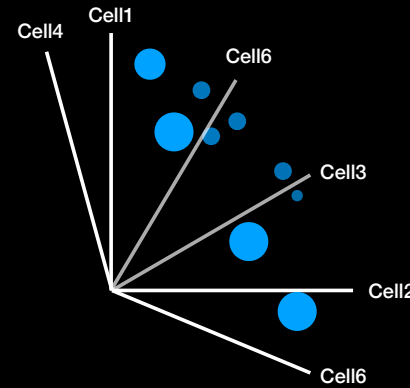


	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Or draw some crazy graph that has an axis for each cell and makes our brains hurt!

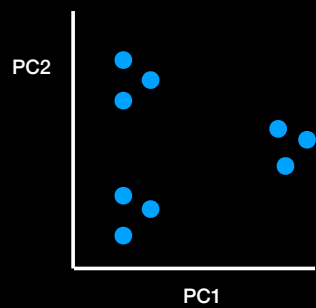


	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...



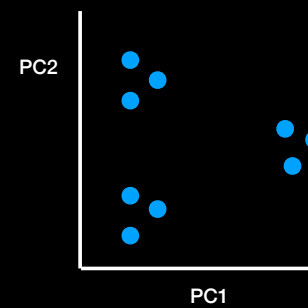
	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Enter Principal Component Analysis (PCA)



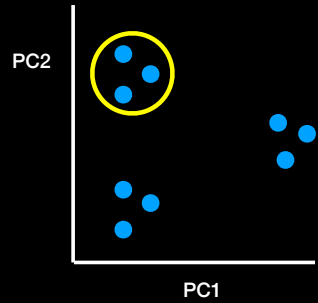
	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

PCA converts the correlations (or lack thereof) among all cells into a representation we can more readily interpret (e.g. a 2D graph!)



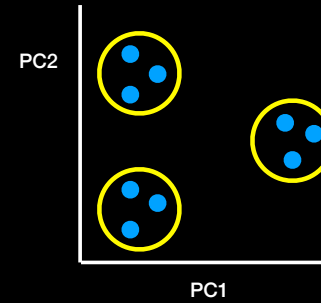
	Cell1	Cell2	Cell3	Cell4	...
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Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Cells that are highly **correlated** cluster together



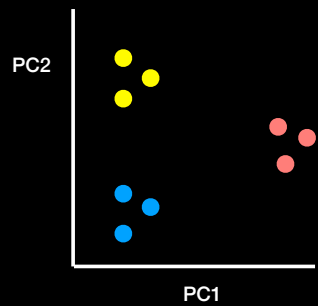
	Cell1	Cell2	Cell3	Cell4	...
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Gene2	2.9	0.8	2.2	1.8	...
Gene3	2.2	1	1.5	3.2	...
Gene4	2	1.4	2	0.3	...
Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

Cells that are highly correlated cluster together



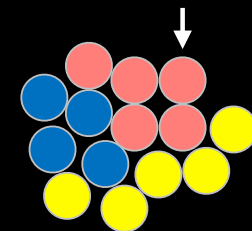
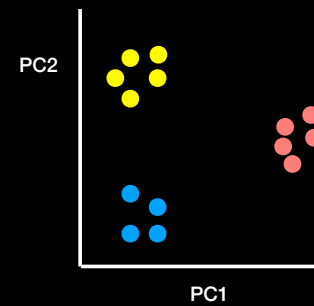
	Cell1	Cell2	Cell3	Cell4	...
Gene1	3	0.25	2.8	0.1	...
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Gene5	1.3	1.6	1.6	0	...
Gene6	1.5	2	2.1	3	...
Gene7	1.1	2.2	1.2	2.8	...
Gene8	1	2.7	0.9	0.3	...
Gene9	0.4	3	0.6	0.1	...

To make the clusters easier to see we can color code them...

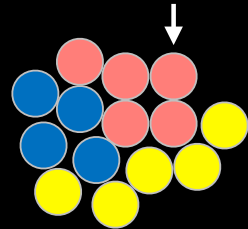
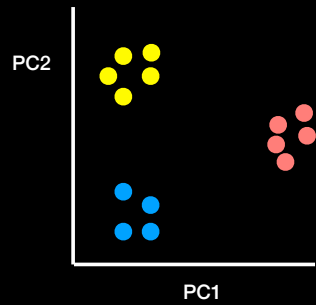


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Once we have identified the clusters from our PCA results, we can go back to our original cells...



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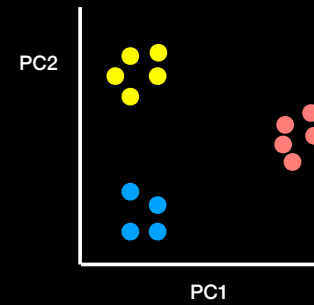


...and see they represent three different types of cells doing three different things with their genes!

Some key points:

The PCs (i.e. new plot axis) are ranked by their importance

So PC1 is more important than PC2 which in turn is more important than PC3 etc.

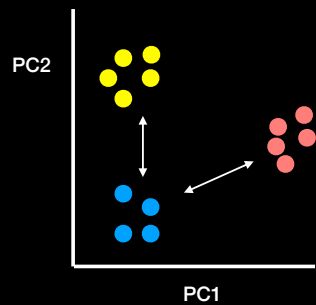


Some key points:

The PCs (i.e. new plot axis) are ranked by their importance

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So the red and blue cluster are more dissimilar than the yellow and blue clusters



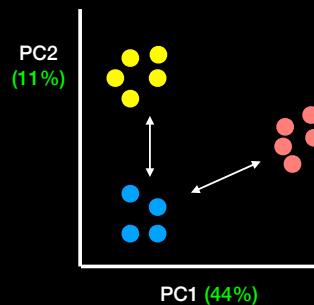
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The PCs (i.e. new plot axis) are ranked by the amount of variance in the original data (i.e. gene expression values) that they "capture"



Some key points:

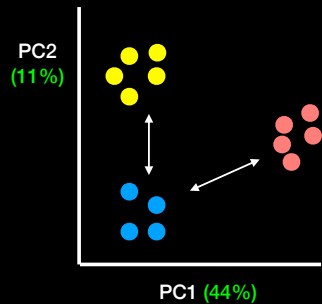
The PCs (i.e. new plot axis) are ranked by their importance

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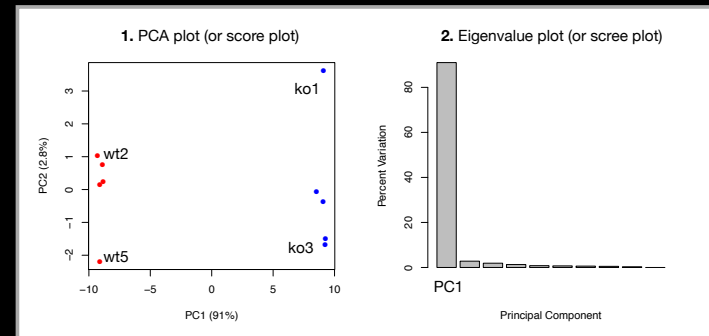
So the red and blue cluster are more dissimilar than the yellow and blue clusters

The PCs (i.e. new plot axis) are ranked by the amount of variance in the original data (i.e. gene expression values) that they "capture"

In this example PC1 'captures' 4x more of the original variance than PC2 ($44/11 = 4$)



- We actually get two main things out of a typical PCA
 - The new axis (called PCs or **Eigenvectors**) and
 - **Eigenvalues** that detail the amount of variance captured by each PC



- Another cool thing we can get out of PCA is a quantitative report on how the original variables contributed to each PC
 - In other words, which were the most important genes that lead to the observed clustering in PC-space
 - These are often called the **loadings** and we can plot them to see which are the most important genes for the observed separation as well as outputting ranked lists of genes that act to discriminate the samples

gene64	gene39
0.1047968	0.1047629

gene7	gene65
-0.1047629	-0.1047443

Hands-on time!

<http://setosa.io/ev/principal-component-analysis/>

Do it Yourself!

Outline: How to do PCA in R

- How to use the **prcomp()** function to do PCA.
- How to draw and interpret PCA plots
- How to determine how much variation each principal component accounts for and the the “intrinsic dimensionality” useful for further analysis
- How to examine the **loadings** (or loading scores) to determine what variables have the largest effect on the graph and are thus important for discriminating samples.

- First lets generate some example data to work with.

```
## Initialize a blank 100 row by 10 column matrix  
mydata <- matrix(nrow=100, ncol=10)
```

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```
## Initialize a blank 100 row by 10 column matrix  
mydata <- matrix(nrow=100, ncol=10)  
  
## Lets label the rows gene1, gene2 etc. to gene100  
rownames(mydata) <- paste("gene", 1:100, sep="")
```

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```
## Initialize a blank 100 row by 10 column matrix  
mydata <- matrix(nrow=100, ncol=10)  
  
## Lets label the rows gene1, gene2 etc. to gene100  
rownames(mydata) <- paste("gene", 1:100, sep="")  
  
## Lets label the first 5 columns wt1, wt2, wt3, wt4 and wt5  
## and the last 5 ko1, ko2 etc. to ko5 (for "knock-out")  
colnames(mydata) <- c( paste("wt", 1:5, sep=""),  
  paste("ko", 1:5, sep="") )
```


- First lets generate some example data to work with.

```
## Initialize a blank 100 row by 10 column matrix
mydata <- matrix(nrow=100, ncol=10)

## Lets label the rows gene1, gene2 etc. to gene100
rownames(mydata) <- paste("gene", 1:100, sep="")

## Lets label the first 5 columns wt1, wt2, wt3, wt4 and wt5
## and the last 5 ko1, ko2 etc. to ko5 (for "knock-out")
colnames(mydata) <- c( paste("wt", 1:5, sep=""),
                      paste("ko", 1:5, sep="") )

## Fill in some fake read counts
for(i in 1:nrow(mydata)) {
  wt.values <- rpois(5, lambda=sample(x=10:1000, size=1))
  ko.values <- rpois(5, lambda=sample(x=10:1000, size=1))

  mydata[i,] <- c(wt.values, ko.values)
}

head(mydata)
```

- **NOTE:** the samples are columns, and the genes are rows!

```
mydata <- matrix(nrow=100, ncol=10)

rownames(mydata) <- paste("gene", 1:100, sep="")

colnames(mydata) <- c( paste("wt", 1:5, sep=""),
                      paste("ko", 1:5, sep="") )

for(i in 1:nrow(mydata)) {
  wt.values <- rpois(5, lambda=sample(x=10:1000, size=1))
  ko.values <- rpois(5, lambda=sample(x=10:1000, size=1))
  mydata[i,] <- c(wt.values, ko.values)
}

head(mydata)
```

	wt1	wt2	wt3	wt4	wt5	ko1	ko2	ko3	ko4	ko5
gene1	147	171	160	175	187	63	57	58	55	59
gene2	151	134	148	126	134	838	831	894	847	830
gene3	702	672	653	681	701	593	579	644	596	610
gene4	319	297	310	296	304	754	807	734	750	774

- Now we have our data we call `prcomp()` to do PCA
- **NOTE:** `prcomp()` expects the samples to be rows and genes to be columns so we need to first transpose the matrix with the `t()` function!

```
## lets do PCA
pca <- prcomp(t(mydata), scale=TRUE)
```

- Now we have our data we call `prcomp()` to do PCA

- **NOTE:** `prcomp()` expects the samples to be rows and genes to be columns so we need to first transpose the matrix with the `t()` function!

```
## lets do PCA
pca <- prcomp(t(mydata), scale=TRUE)

## See what is returned by the prcomp() function
attributes(pca)

# $names
#[1] "sdev"      "rotation" "center"   "scale"    "x"
#
# $class
#[1] "prcomp"
```

- The returned `pca$x` here contains the principal components (PCs) for drawing our first graph.
- Here we will take the first two columns in `pca$x` (corresponding to PC1 and PC2) to draw a 2-D plot

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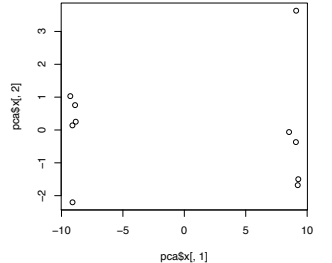
```
## lets do PCA
pca <- prcomp(t(mydata), scale=TRUE)

## A basic PC1 vs PC2 2-D plot
plot(pca$x[,1], pca$x[,2])
```

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```

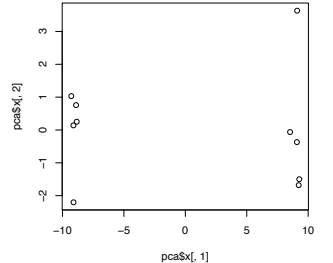


- Looks interesting with a nice separation of samples into two groups of 5 samples each
- Now we can use the square of `pca$sdev`, which stands for “standard deviation”, to calculate how much variation in the original data each PC accounts for

```
## lets do PCA
pca <- prcomp(t(mydata), scale=TRUE)

## A basic PC1 vs PC2 2-D plot
plot(pca$x[,1], pca$x[,2])

## Variance captured per PC
pca.var <- pca$sdev^2
```



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## lets do PCA
pca <- prcomp(t(mydata), scale=TRUE)

## A basic PC1 vs PC2 2-D plot
plot(pca$x[,1], pca$x[,2])

## Percent variance is often more informative to look at
pca.var <- pca$sdev^2
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)
```

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pca.var.per <- round(pca.var/sum(pca.var)*100, 1)

pca.var.per

[1] 91.0 2.8 1.9 1.3 0.8 0.7 0.6 0.5 0.3 0.0
```

- Looks interesting with a nice separation of samples into two groups of 5 samples each
- Now we can use the square of `pca$sdev`, which stands for “standard deviation”, to calculate how much variation in the original data each PC accounts for

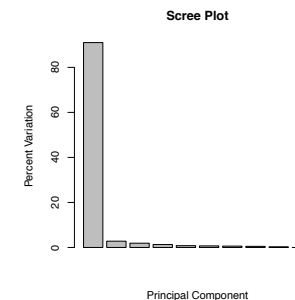
```
pca.var <- pca$sdev^2
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)

barplot(pca.var.per, main="Scree Plot",
        xlab="Principal Component", ylab="Percent Variation")
```

- From the “**scree plot**” it is clear that **PC1** accounts for almost all of the variation in the data!

```
pca.var <- pca$sdev^2
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)

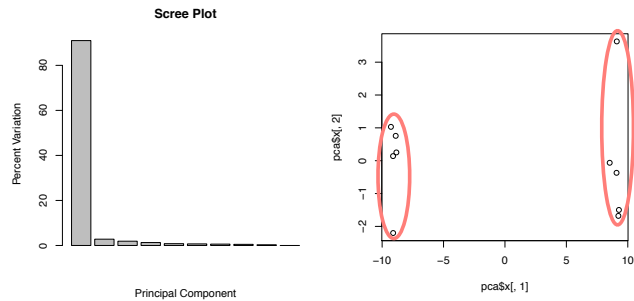
barplot(pca.var.per, main="Scree Plot",
        xlab="Principal Component", ylab="Percent Variation")
```



- Which means there are big differences between these two groups that are separated along the PC1 axis...

```
pca.var <- pca$sdev^2
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)
```

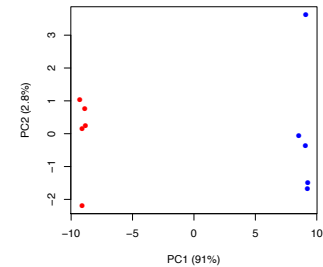
```
barplot(pca.var.per, main="Scree Plot",
        xlab="Principal Component", ylab="Percent Variation")
```



- Lets make our plot a bit more useful...

```
## A vector of colors for wt and ko samples
colvec <- colnames(mydata)
colvec[grep("wt", colvec)] <- "red"
colvec[grep("ko", colvec)] <- "blue"

plot(pca$x[,1], pca$x[,2], col=colvec, pch=16,
     xlab=paste0("PC1 (", pca.var.per[1], "%)"),
     ylab=paste0("PC2 (", pca.var.per[2], "%)"))
```

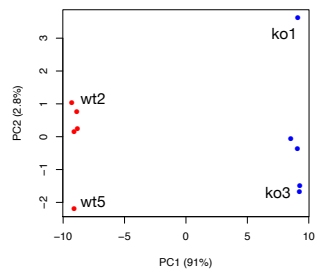


- And add some labels...

```
plot(pca$x[,1], pca$x[,2], col=colvec, pch=16,
     xlab=paste0("PC1 (", pca.var.per[1], "%)"),
     ylab=paste0("PC2 (", pca.var.per[2], "%)"))
```

```
## Click to identify which sample is which
identify(pca$x[,1], pca$x[,2], labels=colnames(mydata))
```

```
## Press ESC to exit...
```



- Finally, lets look at how to use the loading scores to determine which genes have the largest effect on where samples are plotted in the PCA plot

- The `prcomp()` function calls loading scores `$rotation`

```
## Lets focus on PC1 as it accounts for > 90% of variance
loading_scores <- pca$rotation[,1]
```

- Finally, lets look at how to use the **loading scores** to determine which genes have the largest effect on where samples are plotted in the PCA plot
 - The `prcomp()` function calls loading scores `$rotation`

```
## Lets focus on PC1 as it accounts for > 90% of variance
loading_scores <- pca$rotation[,1]

summary(loading_scores)
  Min.   1st Qu.   Median     Mean   3rd Qu.    Max.
-0.104763 -0.104276 -0.068784 -0.005656  0.103926  0.104797

## We are interested in the magnitudes of both plus
## and minus contributing genes
gene_scores <- abs(loading_scores)
```

- Finally, lets look at how to use the **loading scores** to determine which genes have the largest effect on where samples are plotted in the PCA plot
 - The `prcomp()` function calls loading scores `$rotation`

```
loading_scores <- pca$rotation[,1]
gene_scores <- abs(loading_scores)

## Sort by magnitudes from high to low
gene_score_ranked <- sort(gene_scores, decreasing=TRUE)
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top_5_genes <- names(gene_score_ranked[1:5])
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## Find the names of the top 5 genes
top_5_genes <- names(gene_score_ranked[1:5])

## Show the scores (with +/- sign)
pca$rotation[top_5_genes,1]
```

- Here we see genes with the largest positive **loading scores** that effectively ‘push’ the “ko” samples to the right positive side of the plot.
- And the genes with high negative scores that push “wt” samples to the left side of the plot.

```
loading_scores <- pca$rotation[,1]
gene_scores <- abs(loading_scores)

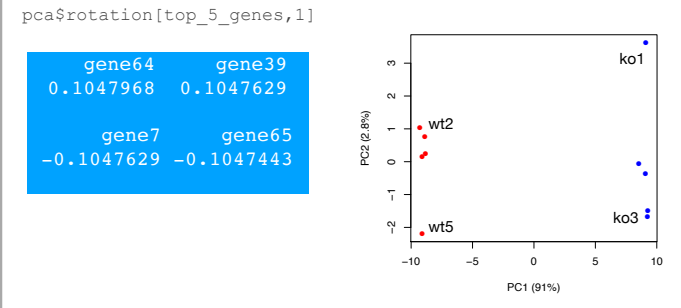
## Sort by magnitudes from high to low
gene_score_ranked <- sort(gene_scores, decreasing=TRUE)

## Find the names of the top 5 genes
top_5_genes <- names(gene_score_ranked[1:5])

## Show the scores (with +/- sign)
pca$rotation[top_5_genes,1]
```

gene64	gene39	gene7	gene60	gene65
0.1047968	0.1047629	-0.1047629	0.1047601	-0.1047443

- Here we see genes with the largest positive **loading scores** that effectively ‘push’ the “ko” samples to the right positive side of the plot.
- And the genes with high negative scores that push “wt” samples to the left side of the plot.



PCA Recap

- PCA is classic “**multivariate statistical technique**” used to reduce the dimensionality of a complex data set to a more manageable number (typically 2D or 3D)
- For a matrix of m genes \times n samples, we mean center (i.e. subtract the sample mean from each sample column), optionally rescale the values for each sample column, then calculate a new **covariance matrix** of size $n \times n$
- We finally diagonalize the covariance matrix to yield our n **Eigenvectors** (called principal components or PCs) and n **Eigenvalues**.
- The top PCs (with largest Eigenvalues) retain the essential features of the original data and represent a useful subspace for further analysis (e.g. visualization, clustering, feature extraction, outlier detection etc...)

PCA objectives in a nutshell

- to reduce dimensionality
- to visualize multidimensional data
- to choose the most useful variables (features)
- to identify groupings of objects (e.g. genes/samples)
- to identify outliers

Do it Yourself!

Your turn!

Perform a PCA on the UK foods dataset

https://bioboot.github.io/bimm143_F18/class-material/UK_food_pca/

Input: read, View/head,
PCA: prcomp,
Plots: PCA plot
scree plot,
loadings plot.

[[Muddy Point Feedback Link](#)]

The End!

[[Muddy Point Feedback Link](#)]

Practical issues with PCA

- Scaling the data
- Missing values:

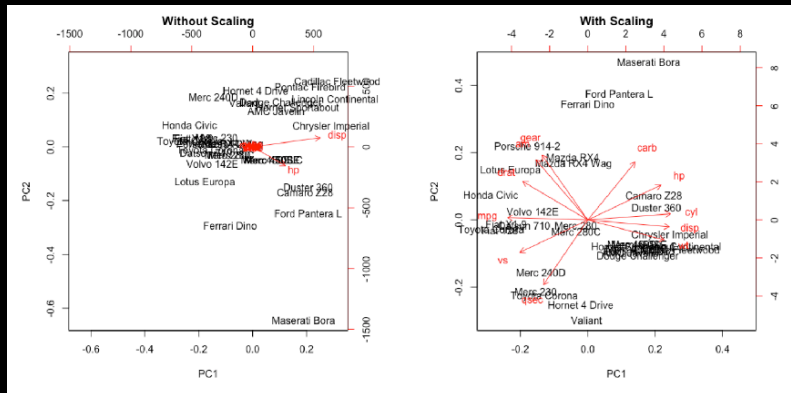
Scaling

```
> data(mtcars)
> head(mtcars)
      mpg  cyl  disp  hp  drat   wt  qsec vs  am  gear  carb
Mazda RX4    21.0   6  160  110  3.90 2.620 16.46 0  1   4   4
Mazda RX4 Wag 21.0   6  160  110  3.90 2.875 17.02 0  1   4   4
Datsun 710    22.8   4  108   93  3.85 2.320 18.61 1  1   4   1
Hornet 4 Drive 21.4   6  258  110  3.08 3.215 19.44 1  0   3   1
Hornet Sportabout 18.7  8  360  175  3.15 3.440 17.02 0  0   3   2
Valiant      18.1   6  225  105  2.76 3.460 20.22 1  0   3   1

# Means and standard deviations vary a lot
> round(colMeans(mtcars), 2)
  mpg   cyl  disp  hp  drat   wt  qsec   vs   am  gear  carb
20.09  6.19 230.72 146.69  3.60  3.22 17.85  0.44  0.41  3.69  2.81
> round(apply(mtcars, 2, sd), 2)
  mpg   cyl  disp  hp  drat   wt  qsec   vs   am  gear  carb
 6.03  1.79 123.94  68.56  0.53  0.98  1.79  0.50  0.50  0.74  1.62
```

Scaling

```
prcomp(x, center=TRUE, scale=FALSE)  
prcomp(x, center=TRUE, scale=TRUE)
```



Practical issues with PCA

- Scaling the data
- Missing values:
 - Drop observations with missing values
 - Impute / estimate missing values