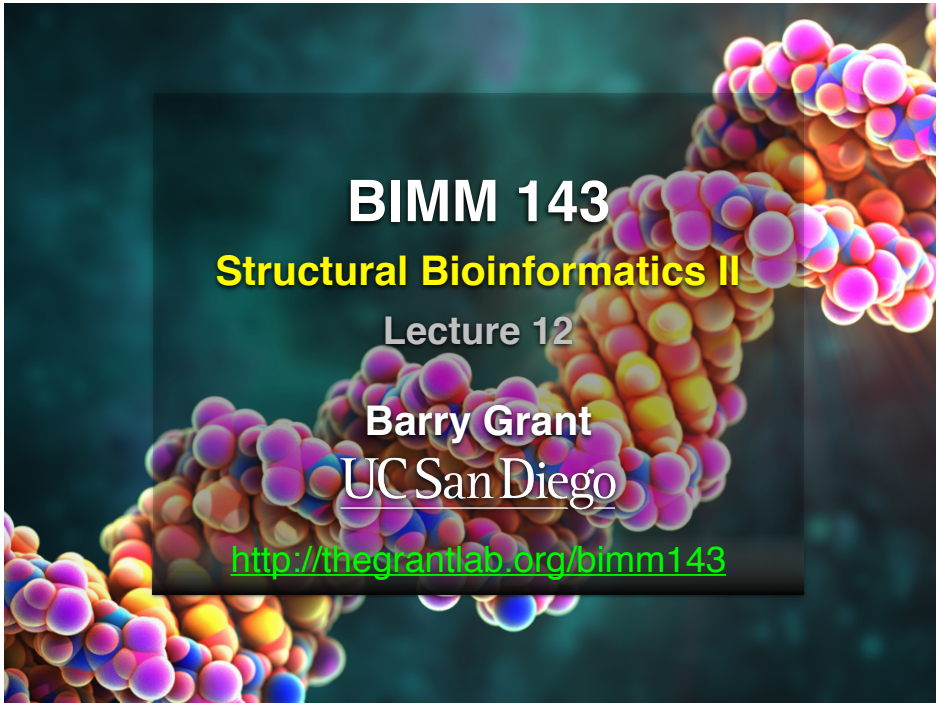
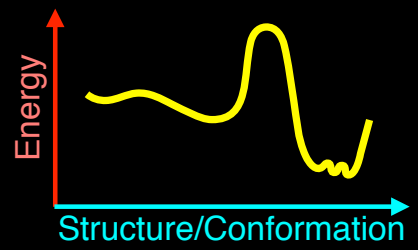


Next Up:

- **Overview of structural bioinformatics**
  - Motivations, goals and challenges
- **Fundamentals of protein structure**
  - Structure composition, form and forces
- **Representing, interpreting & modeling protein structure**
  - Visualizing and interpreting protein structures
  - Analyzing protein structures
  - Modeling energy as a function of structure
  - Drug discovery & Predicting functional dynamics

Key concept:

Potential functions describe a systems **energy** as a function of its **structure**



Two main approaches:

(1). Physics-Based

(2). Knowledge-Based

Two main approaches:

(1). Physics-Based

(2). Knowledge-Based

For physics based potentials
energy terms come from physical theory

$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

$$V(R) = E_{bonded} + E_{non.bonded}$$

Sum of bonded and non-bonded
atom-type and position based terms

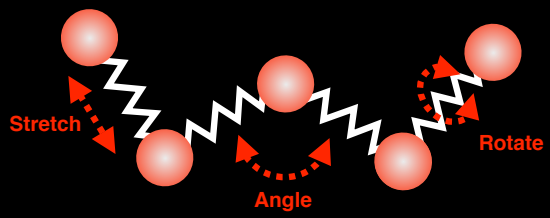
$$V(R) = E_{bonded} + E_{non.bonded}$$

E_{bonded} is itself a sum of three terms:

$$V(R) = E_{bonded} + E_{non.bonded}$$

E_{bonded} is itself a sum of three terms:

$$E_{bond.stretch} + E_{bond.angle} + E_{bond.rotate}$$



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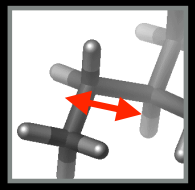
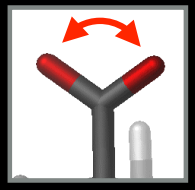


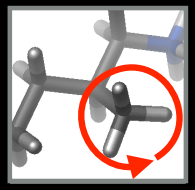
Bond Stretch

$E_{bond.stretch}$



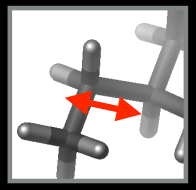
Bond Angle

$E_{bond.angle}$



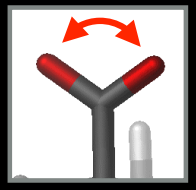
Bond Rotate

$E_{bond.rotate}$



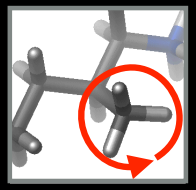
Bond Stretch

$$\sum_{bonds} K_i^{bs}(b_i - b_o)$$



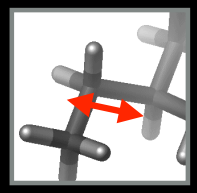
Bond Angle

$$\sum_{angles} K_i^{ba}(\theta_i - \theta_o)$$



Bond Rotate

$$\sum_{dihedrals} K_i^{br}[1 - cos(n_i\phi_i - \phi_o)$$



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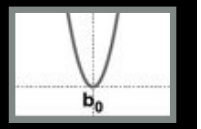

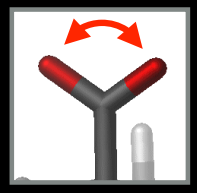

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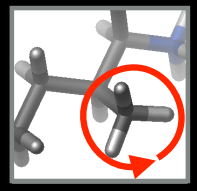

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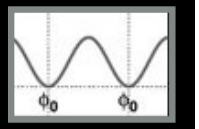

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$E_{non.bonded}$ is a sum of two terms:

$$V(R) = E_{bonded} + E_{non.bonded}$$

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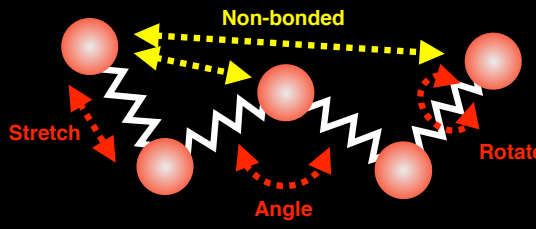
$$E_{van.der.Waals} + E_{electrostatic}$$



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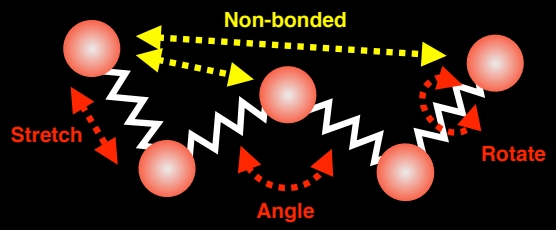
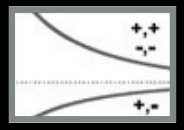
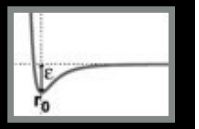


$$E_{electrostatic} = \sum_{pairs.i.j} \frac{q_i q j}{\epsilon r_{ij}}$$



$$E_{van.der.Waals} = \sum_{pairs.i.j} \left[\epsilon_{ij} \left(\frac{r_{o.ij}}{r_{ij}} \right)^{12} - 2\epsilon_{ij} \left(\frac{r_{o.ij}}{r_{ij}} \right)^{6} \right]$$



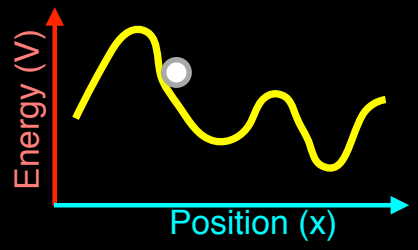
Total potential energy

The potential energy can be given as a sum of terms for: Bond stretching, Bond angles, Bond rotations, van der Walls and Electrostatic interactions between atom pairs

$$V(R) = \underbrace{E_{bond.stretch} + E_{bond.angle} + E_{bond.rotate}}_{E_{bonded}} + \underbrace{E_{van.der.Waals} + E_{electrostatic}}_{E_{non.bonded}}$$

Potential energy surface

Now we can calculate the potential energy surface that fully describes the energy of a molecular system as a function of its geometry



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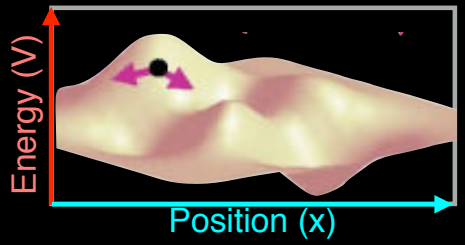
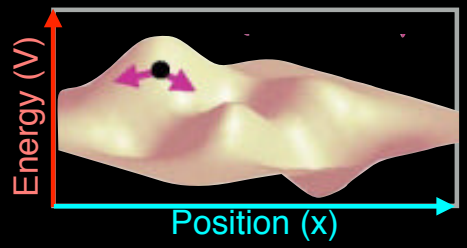


Key concept:

Now we can calculate the potential energy surface that fully describes the energy of a molecular system as a function of its geometry



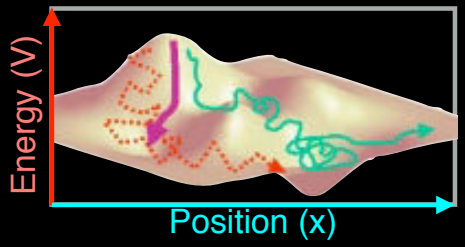
- The **forces** are the gradients of the energy

$$F(x) = -dV/dx$$

Moving Over The Energy Surface

- **Energy Minimization** drops into local minimum
- **Molecular Dynamics** uses thermal energy to move smoothly over surface
- **Monte Carlo Moves** are random. Accept with probability:

$$exp(-\Delta V/dx)$$



PHYSICS-ORIENTED APPROACHES

Weaknesses

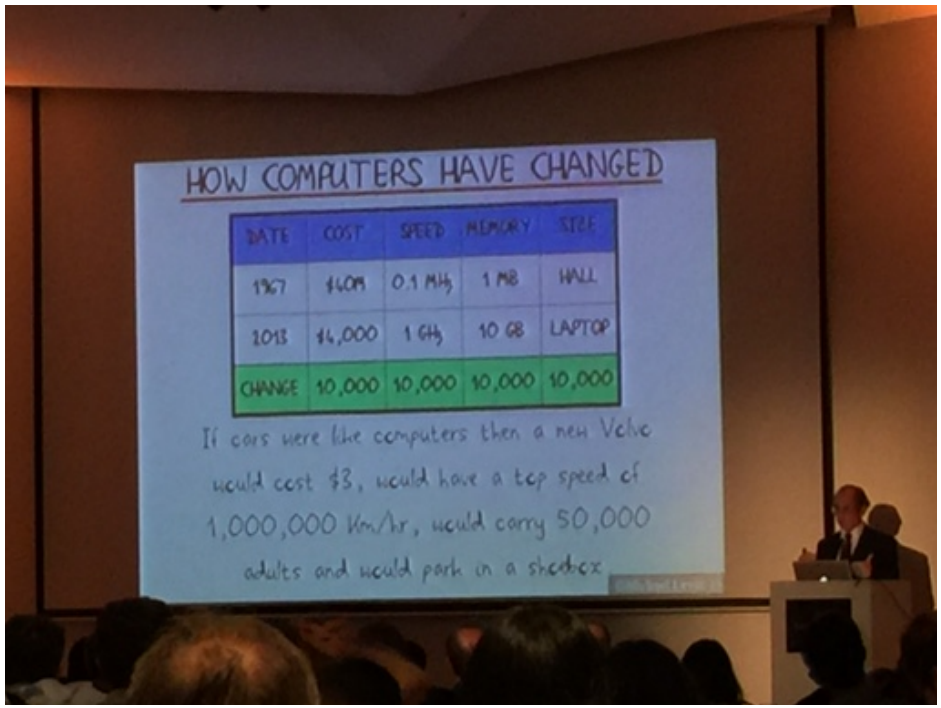
- Fully physical detail becomes computationally intractable
- Approximations are unavoidable
  - (Quantum effects approximated classically, water may be treated crudely)
- Parameterization still required

Strengths

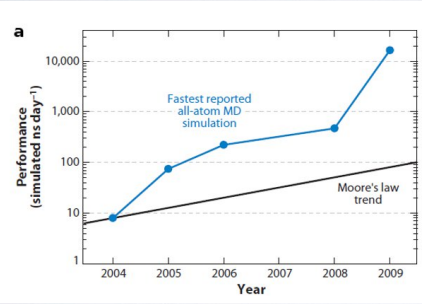
- Interpretable, provides guides to design
- Broadly applicable, in principle at least
- Clear pathways to improving accuracy

Status

- Useful, widely adopted but far from perfect
- Multiple groups working on fewer, better approxs
  - Force fields, quantum
  - entropy, water effects
- Moore's law: hardware improving



SIDE-NOTE: GPUS AND ANTON SUPERCOMPUTER



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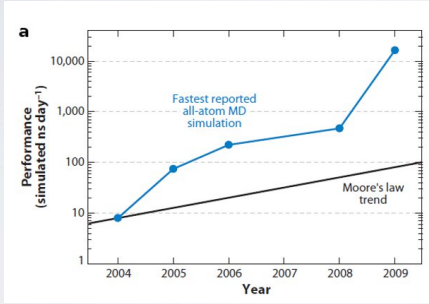


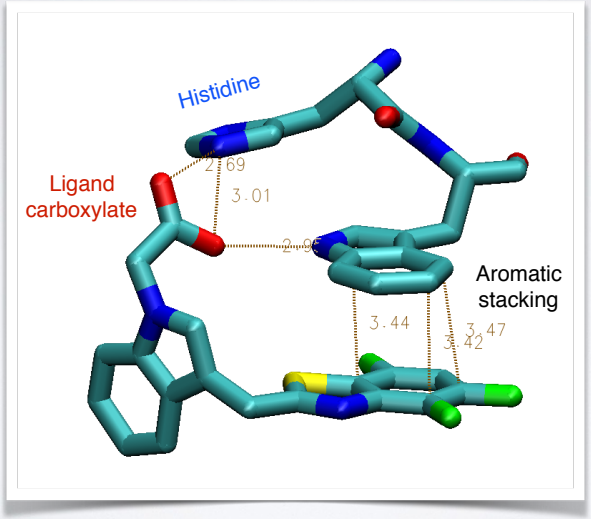
POTENTIAL FUNCTIONS DESCRIBE A SYSTEMS **ENERGY** AS A FUNCTION OF ITS **STRUCTURE**

Two main approaches:

(1). Physics-Based

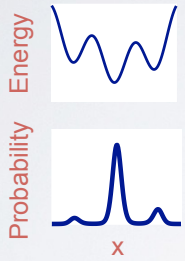
(2). Knowledge-Based

KNOWLEDGE-BASED DOCKING POTENTIALS



ENERGY DETERMINES **PROBABILITY** (STABILITY)

Basic idea: Use probability as a proxy for energy



Boltzmann:

$$p(r) \propto e^{-E(r)/RT}$$

Inverse Boltzmann:

$$E(r) = -RT \ln\left[p(r)\right]$$

Example: ligand carboxylate O to protein histidine N

Find all protein-ligand structures in the PDB with a ligand carboxylate O

1. For each structure, histogram the distances from O to every histidine N
2. Sum the histograms over all structures to obtain $p(r_{O\text{-}N})$
3. Compute $E(r_{O\text{-}N})$ from $p(r_{O\text{-}N})$

KNOWLEDGE-BASED POTENTIALS

Weaknesses

Accuracy limited by availability of data

Strengths

Relatively easy to implement
Computationally fast

Status

Useful, far from perfect
May be at point of diminishing returns
(not always clear how to make improvements)

Computer Aided Drug Discovery

Next Up:

- **Overview of structural bioinformatics**
  - Motivations, goals and challenges
- **Fundamentals of protein structure**
  - Structure composition, form and forces
- **Representing, interpreting & modeling protein structure**
  - Visualizing and interpreting protein structures
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  - Modeling energy as a function of structure
  - Drug discovery & Predicting functional dynamics

THE TRADITIONAL EMPIRICAL PATH TO DRUG DISCOVERY

Compound library
(commercial, in-house, synthetic, natural)
→ High throughput screening
(HTS)
→ Hit confirmation
→ Lead compounds
(e.g., μM K_d)
→ Lead optimization
(Medicinal chemistry)
→ Potent drug candidates
(nM K_d)
→ Animal and clinical evaluation

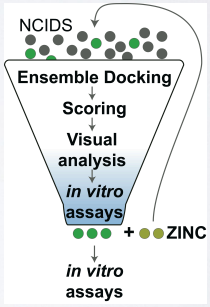
COMPUTER-AIDED LIGAND DESIGN

Aims to reduce number of compounds synthesized and assayed

Lower costs

Reduce chemical waste

Facilitate faster progress



Two main approaches:

(1). Receptor/Target-Based

(2). Ligand/Drug-Based

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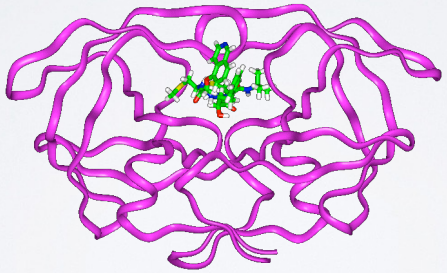
(1). Receptor/Target-Based

(2). Ligand/Drug-Based

SCENARIO 1: RECEPTOR-BASED DRUG DISCOVERY

Structure of Targeted Protein Known: Structure-Based Drug Discovery

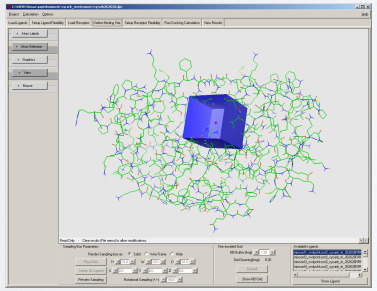


HIV Protease/KNI-272 complex

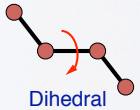
PROTEIN-LIGAND DOCKING

Structure-Based Ligand Design

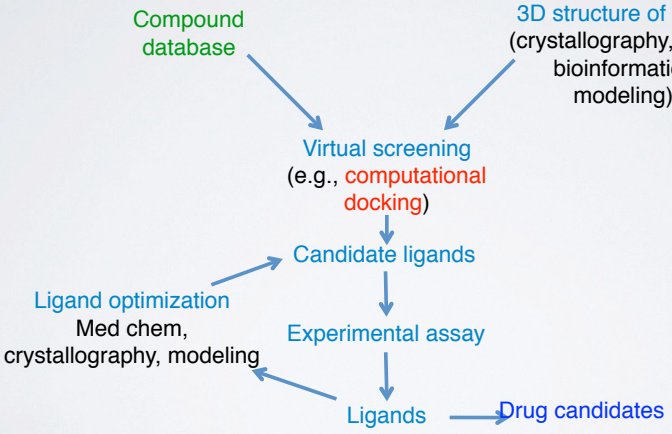
Docking software
Search for structure of lowest energy



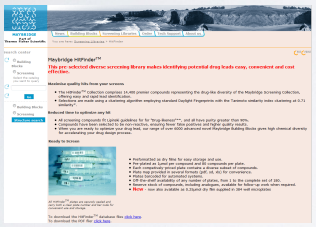
Potential function
Energy as function of structure



STRUCTURE-BASED VIRTUAL SCREENING

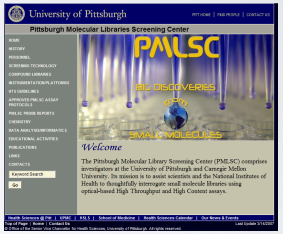


COMPOUND LIBRARIES



Commercial (in-house pharma)

Government (NIH)



Academia

COMMON SIMPLIFICATIONS USED IN PHYSICS-BASED DOCKING

Quantum effects approximated classically

Protein often held rigid

Configurational entropy neglected

Influence of water treated crudely

Do it Yourself!

Hand-on time!

https://bioboot.github.io/bimm143_S19/lectures/#13

You can use the classroom computers or your own laptops. If you are using your laptops then you will need to install **MGLTools**

Two main approaches:

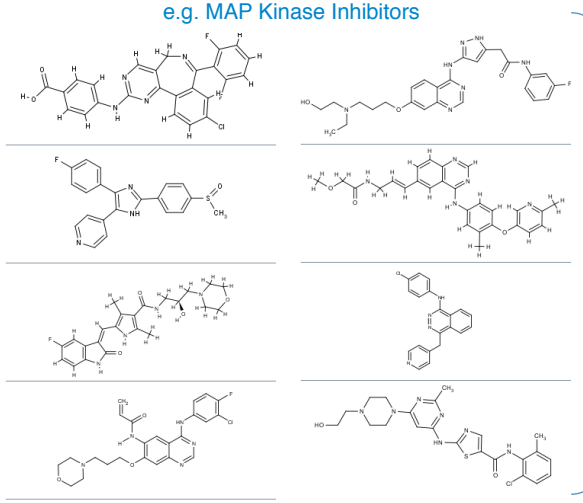
(1). Receptor/Target-Based

(2). Ligand/Drug-Based

Scenario 2

Structure of Targeted Protein Unknown: Ligand-Based Drug Discovery

e.g. MAP Kinase Inhibitors



Using knowledge of existing inhibitors to discover more

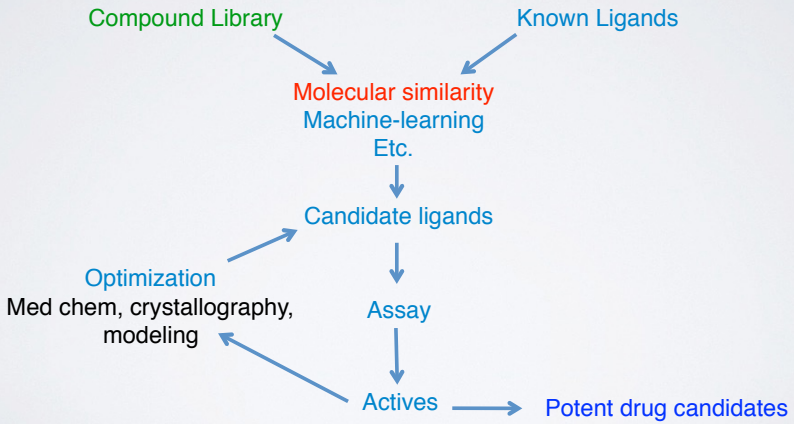
Why Look for Another Ligand if You Already Have Some?

Experimental screening generated some ligands, but they don't bind tightly enough

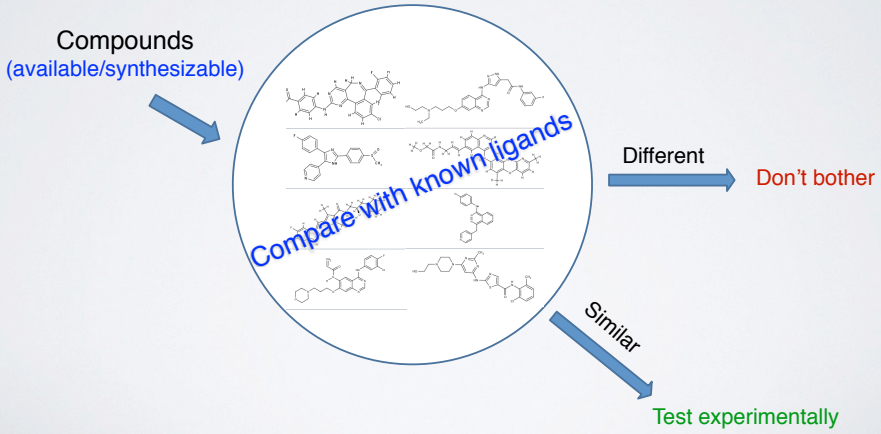
A company wants to work around another company's chemical patents

An high-affinity ligand is toxic, is not well-absorbed, difficult to synthesize etc.

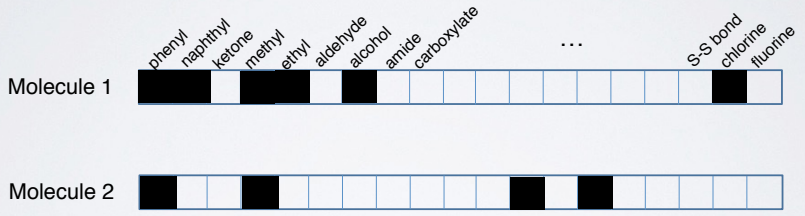
LIGAND-BASED VIRTUAL SCREENING



CHEMICAL SIMILARITY LIGAND-BASED DRUG-DISCOVERY



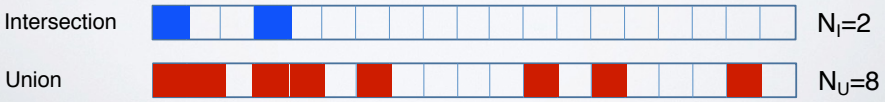
CHEMICAL FINGERPRINTS
BINARY STRUCTURE KEYS



CHEMICAL SIMILARITY FROM FINGERPRINTS

Tanimoto Similarity (or Jaccard Index), T

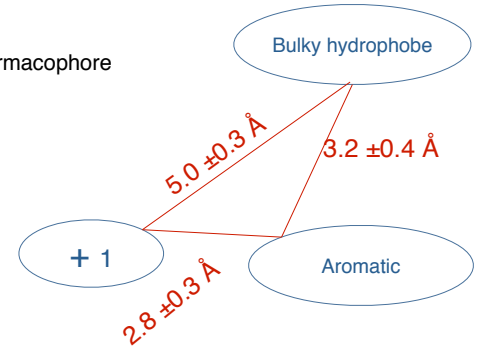
$$T \equiv \frac{N_I}{N_U} = 0.25$$



Pharmacophore Models

Φάρμακο (drug) + Φορά (carry)

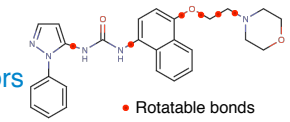
A 3-point pharmacophore



Molecular Descriptors

More abstract than chemical fingerprints

Physical descriptors
- molecular weight
- charge
- dipole moment
- number of H-bond donors/acceptors
- number of rotatable bonds
- hydrophobicity (log P and clogP)



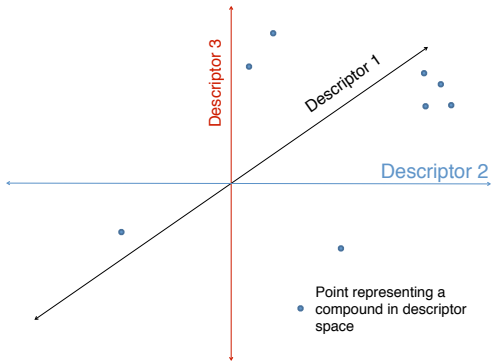
Topological
- branching index
- measures of linearity vs interconnectedness

Etc. etc.

A High-Dimensional "Chemical Space"

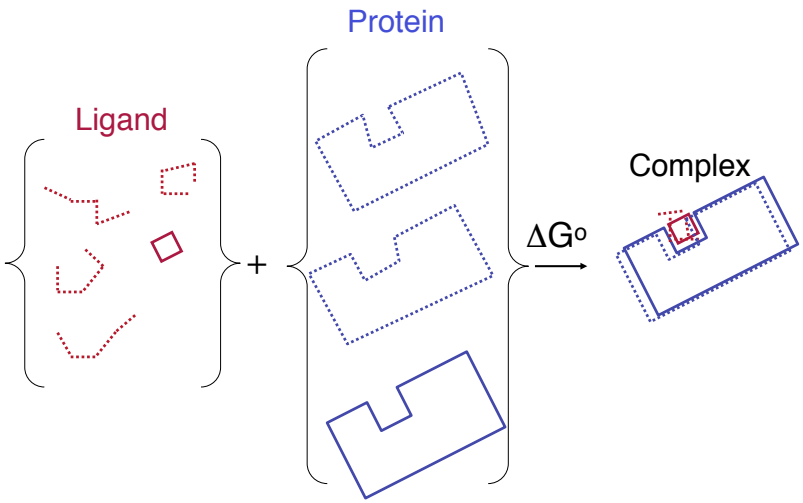
Each compound is a point in an n-dimensional space

Compounds with similar properties are near each other



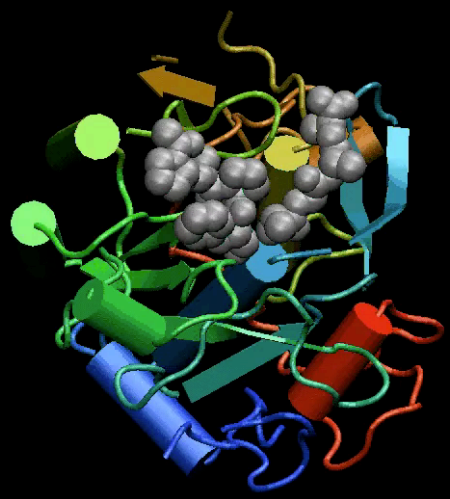
Apply multivariate statistics and machine learning for descriptor-selection. (e.g. partial least squares, PCA, support vector machines, random forest, deep learning etc.)

Proteins and Ligand are Flexible



NMA (Normal Mode Analysis) is a bioinformatics method to predict the intrinsic dynamics of biomolecules

Do it Yourself!



https://bioboot.github.io/bimm143_S19/lectures/#12

NMA in Bio3D

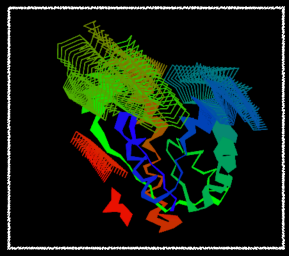
- Normal Mode Analysis (NMA) is a bioinformatics method that can predict the major motions of biomolecules.

```
pdb <- read.pdb("1hel")
modes <- nma( pdb )
m7 <- mktrj(modes, mode=7, file="mode_7.pdb")
```

Then you can open the resulting mode_7.pdb file in VMD
> Use "TUBE" representation and hit the play button...

```
view(m7, col=vec2color(rmsf(m7)))
```

Bio3D view()



- If you want the 3D viewer in your R markdown you can install the development version of bio3d.view
- In your R console:

```
> install.packages("devtools")
> devtools::install_bitbucket("Grantlab/bio3d-view")
> install.packages("rgl")
```

- To use in your R session:

```
> library("bio3d.view")
> pdb <- read.pdb("5p21")
> view(pdb)
> view(pdb, "overview", col="sse")
> view(m7)
```

SideNote: view()

- If you want the interactive 3D viewer in **Rmd** rendered to output: html_output document:

```
```{r}
library(bio3d.view)
library(rgl)
```
```

```
```{r}
modes <- nma( read.pdb("1hel") )
m7 <- mktrj(modes, mode=7, file="mode_7.pdb")

view(m7, col=vec2color(rmsf(m7)))
rglwidget(width=500, height=500)
```
```

Do it Yourself!

Hand-on time!

https://bioboot.github.io/bimm143_S19/lectures/#13

Focus on **section 3** & **4** exploring **NMA** and **PCA apps**

Reference Slides

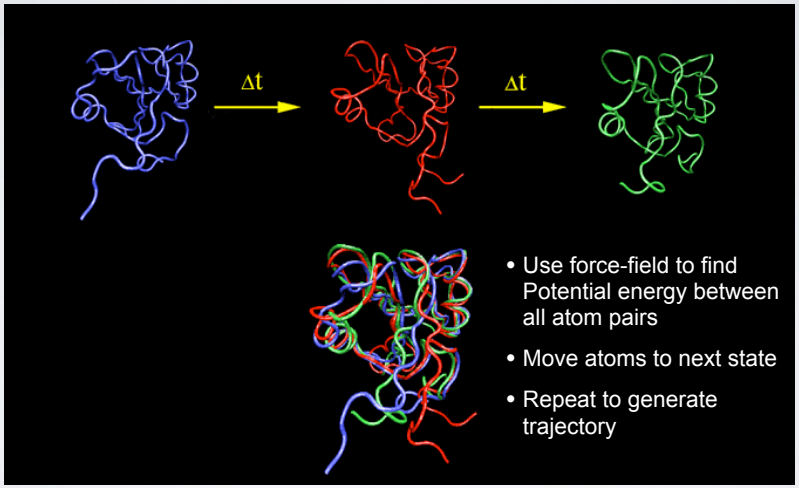
Molecular Dynamics (MD) and Normal Mode Analysis (NMA) Background and Cautionary Notes

[Muddy Point Assessment]

PREDICTING FUNCTIONAL DYNAMICS

- Proteins are intrinsically flexible molecules with internal motions that are often intimately coupled to their biochemical function
  - E.g. ligand and substrate binding, conformational activation, allosteric regulation, etc.
- Thus knowledge of dynamics can provide a deeper understanding of the mapping of structure to function
  - Molecular dynamics (MD) and normal mode analysis (NMA) are two major methods for predicting and characterizing molecular motions and their properties

MOLECULAR DYNAMICS SIMULATION



McCammon, Gelin & Karplus, *Nature* (1977)
[See: https://www.youtube.com/watch?v=ui1ZysMFcKk]

- Divide time into discrete (~1fs) time steps (Δt)
  (for integrating equations of motion, see below)

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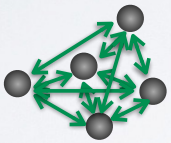
  (for integrating equations of motion, see below)

- At each time step calculate pair-wise atomic forces (F(t))
  (by evaluating force-field gradient)



Nucleic motion described classically

$$m_i \frac{d^2}{dt^2}\vec{R}_i = -\vec{\nabla}_i E(\vec{R})$$

Empirical force field

$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_i(\vec{R})$$

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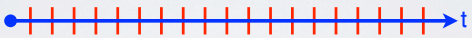

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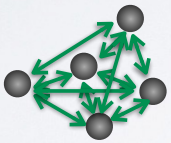

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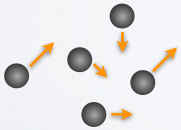
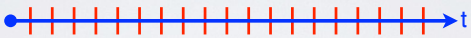
- Use the forces to calculate velocities and move atoms to new positions
  (by integrating numerically via the "leapfrog" scheme)



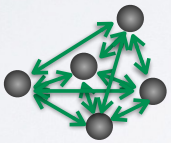
$$\begin{aligned} v(t+\frac{\Delta t}{2}) &= v(t-\frac{\Delta t}{2}) + \frac{F(t)}{m}\Delta t \\ r(t+\Delta t) &= r(t) + v(t+\frac{\Delta t}{2})\Delta t \end{aligned}$$

BASIC ANATOMY OF A MD SIMULATION

- Divide time into discrete (~1fs) time steps (Δt)
  (for integrating equations of motion, see below)



- At each time step calculate pair-wise atomic forces (F(t))
  (by evaluating force-field gradient)



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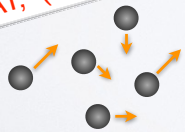
$$m_i \frac{d^2}{dt^2}\vec{R}_i = -\vec{\nabla}_i E(\vec{R})$$

Empirical force field

$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_i(\vec{R})$$

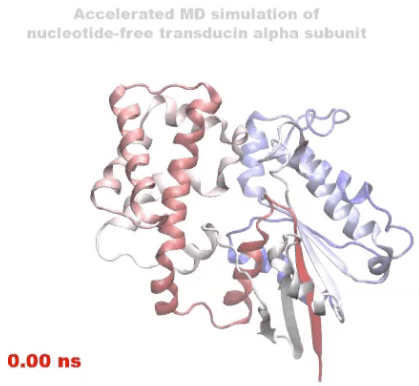
- Use the forces to calculate velocities and move atoms to new positions
  (by integrating numerically via the "leapfrog" scheme)

REPEAT, (iterate many, many times… 1ms = 10^{12} time steps)

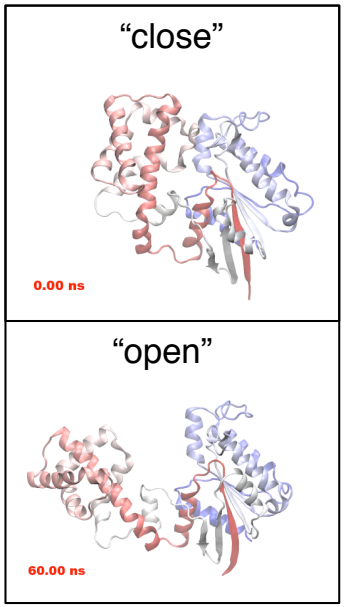


$$\begin{aligned} v(t+\frac{\Delta t}{2}) &= v(t-\frac{\Delta t}{2}) + \frac{F(t)}{m}\Delta t \\ r(t+\Delta t) &= r(t) + v(t+\frac{\Delta t}{2})\Delta t \end{aligned}$$

MD Prediction of Functional Motions

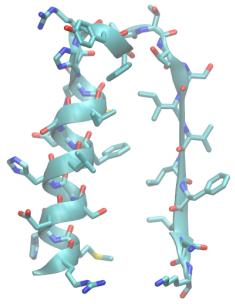


Yao and Grant, Biophys J. (2013)



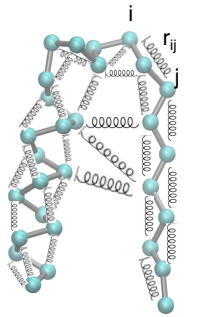
COARSE GRAINING: **NORMAL MODE ANALYSIS** (NMA)

- MD is still time-consuming for large systems
- Elastic network model NMA (ENM-NMA) is an example of a lower resolution approach that finishes in seconds even for large systems.



Atomistic

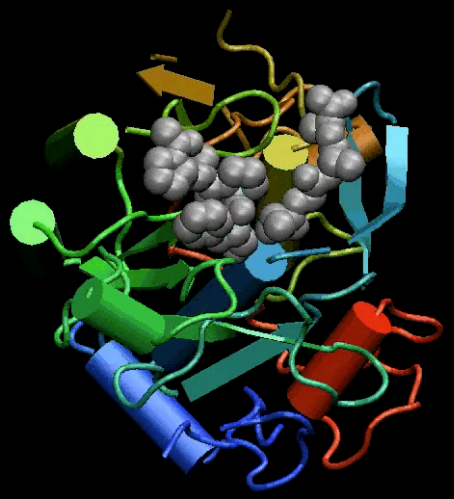
C. G.



Coarse Grained

- 1 bead / 1 amino acid
- Connected by springs

NMA models the protein as a network of elastic strings



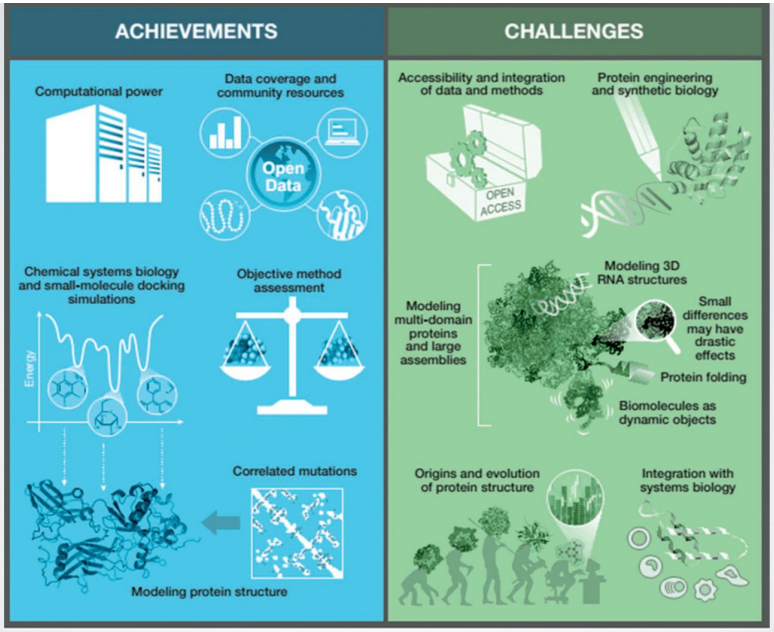
Proteinase K

Do it Yourself!

Hand-on time!

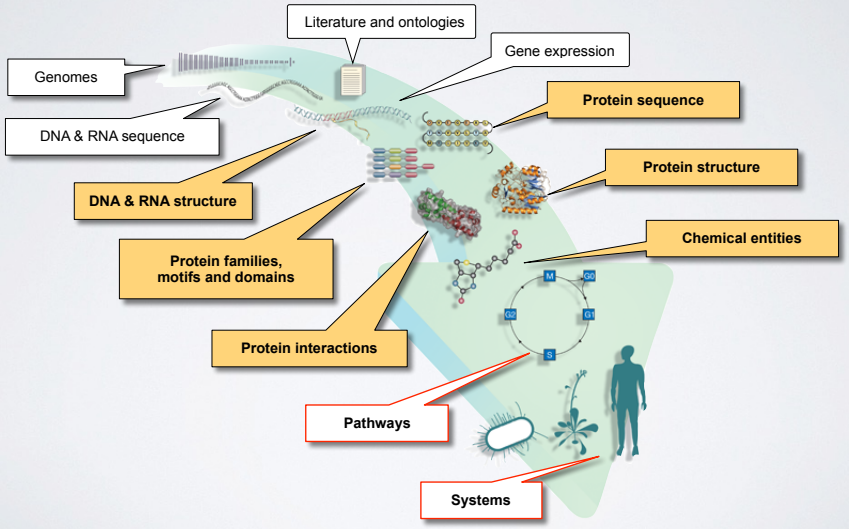
https://bioboot.github.io/bimm143_S19/lectures/#13

Focus on **section 3** & **4** exploring **NMA** and **PCA apps**



Ilan Samish et al. Bioinformatics 2015;31:146-150

INFORMING SYSTEMS BIOLOGY?



SUMMARY

- Structural bioinformatics is computer aided structural biology
- Described major motivations, goals and challenges of structural bioinformatics
- Reviewed the fundamentals of protein structure
- Explored how to visualize protein structure with VMD and use R to perform more advanced structural bioinformatics analysis!
- Introduced both physics and knowledge based modeling approaches for describing the structure, energetics and dynamics of proteins computationally
- Introduced both structure and ligand based bioinformatics approaches for drug discovery and design

[Muddy Point Assessment]

CAUTIONARY NOTES

- A model is never perfect

  A model that is not quantitatively accurate in every respect does not preclude one from establishing results relevant to our understanding of biomolecules as long as the biophysics of the model are properly understood and explored.

- Calibration of parameters is an ongoing imperfect process

  Questions and hypotheses should always be designed such that they do not depend crucially on the precise numbers used for the various parameters.

- A computational model is rarely universally right or wrong

  A model may be accurate in some regards, inaccurate in others. These subtleties can only be uncovered by comparing to all available experimental data.