



BIMM 143
Hands-on Lab Session
Class 03

Barry Grant
UC San Diego

<http://thegrantlab.org/bimm143>

Class 3: Hands-on section

<http://thegrantlab.org/bimm143/>

Week 2

The screenshot shows the bioboot.github.io website. The left sidebar contains links: Overview, **Schedule** (highlighted with a red box and an arrow), Computer Setup, and Learning Goals. The main content area displays a table of course sessions. The table has columns for session number, date, and description. The sessions are:

Session	Date	Description
2	09/30/21	Needleman-Wunsch, Smith-Waterman and BLAST heuristic approaches, Hands on with dot plots, Needleman-Wunsch and BLAST algorithms highlighting their utility and limitations.
3	Tue 10/05/21	Project: Find a gene project assignment (Part 1) Principles of database searching, due in 2 weeks. (Part 2) Sequence analysis, structure analysis and general data analysis with R due at the end of the quarter.
*	Tue 10/05/21	Optional: Advanced sequence alignment and database searching Detecting remote sequence similarity, Database searching beyond BLAST, Substitution matrices, Using PSI-BLAST, Profiles and HMMs, Protein structure comparisons as a gold standard.
4	Thu 10/07/21	Bioinformatics data analysis with R Why do we use R for bioinformatics? R language basics and the RStudio IDE, Major R data structures and functions, Using R interactively from the RStudio console. Introducing Rmarkdown documents.
		Data exploration and visualization in R

Find-a-Gene Project Assignment

- A total of 20% of the course grade will be assigned based on the [“find-a-gene project assignment”](#)

Find-a-Gene Project Assignment

- A total of 20% of the course grade will be assigned based on the [“find-a-gene project assignment”](#)
- The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.

Find-a-Gene Project Assignment

- A total of 20% of the course grade will be assigned based on the [“find-a-gene project assignment”](#)
- The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.
- You may wish to consult the scoring rubric (in the linked project description) and the [example report](#) for format and content guidance.

Find-a-Gene Project Assignment

- A total of 20% of the course grade will be assigned based on the [“find-a-gene project assignment”](#)
- The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.
- You may wish to consult the scoring rubric (in the linked project description) and the [example report](#) for format and content guidance.
- ➔ Your responses to questions **Q1-Q4** are due 12pm San Diego time on Tuesday **Oct 19th** (10/19/21).
- ➔ The complete assignment, including responses to **all questions**, is due 12pm San Diego time on **Dec 2nd** (12/02/21).

Find-a-Gene Project Assignment

- A total of 20% of the course grade will be assigned based on the [“find-a-gene project assignment”](#)
- The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.
- You may wish to consult the scoring rubric (in the linked project description) and the [example report](#) for format and content guidance.

- ➔ Your responses to questions **Q1-Q4** are due 12pm San Diego time on Tuesday **Oct 19th** (10/19/21).
- ➔ The complete assignment, including responses to **all questions**, is due 12pm San Diego time on **Dec 2nd** (12/02/21).

Questions:

[Q1] Tell me the name of a protein you are interested in. Include the species and the accession number. This can be a human protein or a protein from any other species as long as it's function is known.

If you do not have a favorite protein, select human RBP4 or KIF11. Do not use beta globin as this is in the worked example report that I provide you with online.

[Q2] Perform a BLAST search against a DNA database, such as a database consisting of genomic DNA or ESTs. The BLAST server can be at NCBI or elsewhere. Include details of the BLAST method used, database searched and any limits applied (e.g. Organism).

Also include the output of that BLAST search in your document. If appropriate, change the font to Courier size 10 so that the results are displayed neatly. You can also screen capture a BLAST output (e.g. alt print screen on a PC or on a MAC press ⌘-shift-4. The pointer becomes a bulls eye. Select the area you wish to capture and release. The image is saved as a file called Screen Shot [].png in your Desktop directory). It is **not** necessary to print out all of the blast results if there are many pages.

On the BLAST results, clearly indicate a match that represents a protein sequence, encoded from some DNA sequence, that is homologous to your query protein. I need to be able to inspect the pairwise alignment you have selected, including the E value and score. It should be labeled a "genomic clone" or "mRNA sequence", etc. - but include no functional annotation.

In general, [Q2] is the most difficult for students because it requires you to have a “feel” for how to interpret BLAST results. You need to distinguish between a perfect match to your query (i.e. a sequence that is not “novel”), a near match (something that might be “novel”, depending on the results of [Q4]), and a non-homologous result.

If you are having trouble finding a novel gene try restricting your search to an organism that is poorly annotated.

[Q3] Gather information about this “novel” **protein**. At a minimum, show me the protein sequence of the “novel” protein as displayed in your BLAST results from [Q2] as FASTA format (you can copy and paste the aligned sequence subject lines from your BLAST result page if necessary) or translate your novel DNA sequence using a tool called EMBOSS Transeq at the EBI. Don't forget to translate all six reading frames; the ORF (open reading frame) is likely to be the longest sequence without a stop codon. It may not start with a methionine if you don't have the complete coding region. Make sure the sequence you provide includes a header/subject line and is in traditional FASTA format.

Here, tell me the name of the novel protein, and the species from which it derives. It is very unlikely (but still definitely possible) that you will find a novel gene from an organism such as *S. cerevisiae*, human or mouse, because those genomes have already been thoroughly annotated. It is more likely that you will discover a new gene in a genome that is currently being sequenced, such as bacteria or plants or protozoa.

[Q4] Prove that this gene, and its corresponding protein, are novel. For the purposes of this project, “novel” is defined as follows. Take the protein sequence (your answer to [Q3]), and use it as a query in a blastp search of the nr database at NCBI.

- If there is a match with 100% amino acid identity to a protein in the database, from the same species, then your protein is NOT novel (even if the match is to a protein with a name such as “unknown”). Someone has already found and annotated this sequence, and assigned it an accession number.
- If the top match reported has less than 100% identity, then it is likely that your protein is novel, and you have succeeded.
- If there is a match with 100% identity, but to a different species than the one you started with, then you have likely succeeded in finding a novel gene.
- If there are no database matches to the original query from [Q1], this indicates that you have partially succeeded: yes, you may have found a new gene, but no, it is not actually homologous to the original query. You should probably start over.

[Q5] Generate a multiple sequence alignment with your novel protein, your original query protein, and a group of other members of this family from different species. A typical number of proteins to use in a multiple sequence alignment for this assignment purpose is a minimum of 5 and a maximum of 20 - although the exact number is up to you. Include the multiple sequence alignment in your report. Use Courier font with a size appropriate to fit page width.

Side-note: Indicate your sequence in the alignment by choosing an appropriate name for each sequence in the input unaligned sequence file (i.e. edit the sequence file so that the species, or short common, names (rather than accession numbers) display in the output alignment and in the subsequent answers below). The goal in this step is to create an interesting an alignment for building a phylogenetic tree that illustrates species divergence.

BIMM 143

Overview

Schedule

Computer Setup

Learning Goals

Assignments & Grading

Ethics Code

The [find-a-gene project](#)  is a required assignment for BIMM-143. The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.

You may wish to consult the scoring rubric at the end of the above linked project description and the **example report** for format and content guidance.

- Your responses to questions Q1-Q4 are due **Tuesday Oct 19th** (10/19/21) at 12pm San Diego time.
- The complete assignment, including responses to all questions, is due **Thursday Dec 2nd** (12/02/21) at 12pm San Diego time.
- In both instances your PDF format report should be submitted to GradeScope. Late responses will not be accepted under any circumstances.

Videos:

- 3.1 - [Project introduction](#)  Please note: due dates may differ from those in video.

The screenshot shows a web browser window with the address bar displaying 'bioboot.github.io'. The browser's tab bar shows a tab titled 'Schedule · BIMM 143'. The website's navigation bar includes links for Home, Gmail, Gcal, GitHub, BIMM143, BGGN213, GDrive, Atmosphere, CloudLaunch, BIMM194, Blink, News, and a plus sign. On the left side, there is a blue banner with the text 'UC San Diego' and 'BIMM 143'. The main content area features the heading '3: (Project) Find a Gene Assignment Part 1' and a paragraph stating: 'The [find-a-gene project](#) is a required assignment for BIMM-143. The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered to date in class.'

- Your responses to questions **Q1-Q4** are due 12pm San Diego time on Tuesday **Oct 19th** (11/19/21).
- The complete assignment, including responses to **all questions**, is due 12pm San Diego time on Friday **Dec 2nd** (12/02/21).

Class 3: Hands-on section

Class 03

<http://thegrantlab.org/bimm143/>

The screenshot shows the bioboot.github.io website. The left sidebar contains a navigation menu with links: Overview, **Schedule** (highlighted with a red box and an arrow), Computer Setup, and Learning Goals. The main content area displays the course schedule for BIMM 143. The schedule is organized into four sessions, with the third session (Optional: Advanced sequence alignment and database searching) highlighted with a red box and an arrow.

Session	Date	Topic
2	09/30/21	Needleman-Wunsch, Smith-Waterman and BLAST heuristic approaches, Hands on with dot plots, Needleman-Wunsch and BLAST algorithms highlighting their utility and limitations.
3	Tue 10/05/21	Project: Find a gene project assignment (Part 1) Principles of database searching, due in 2 weeks. (Part 2) Sequence analysis, structure analysis and general data analysis with R due at the end of the quarter.
*	Tue 10/05/21	Optional: Advanced sequence alignment and database searching Detecting remote sequence similarity, Database searching beyond BLAST, Substitution matrices, Using PSI-BLAST, Profiles and HMMs, Protein structure comparisons as a gold standard.
4	Thu 10/07/21	Bioinformatics data analysis with R Why do we use R for bioinformatics? R language basics and the RStudio IDE, Major R data structures and functions, Using R interactively from the RStudio console. Introducing Rmarkdown documents.

Below the schedule table, the following topics are listed:

- Data exploration and visualization in R**

► Details:

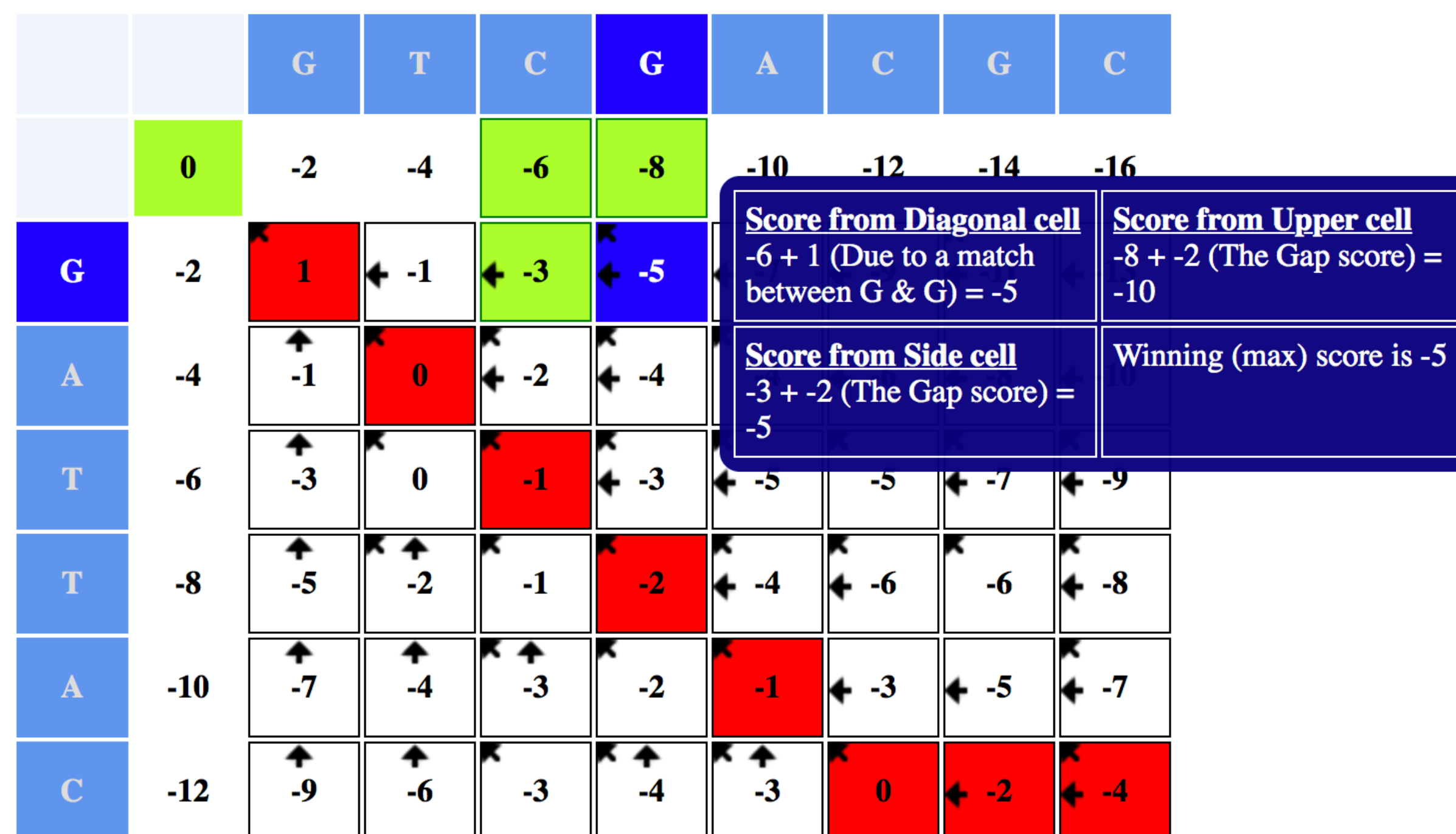
Sequence 1 Sequence 2

Match Score Mismatch Score Gap Score

1 -1 -2

G	T	C	G	A	C	G	C
G	A	T	T	A	C	-	-

Score = -4



▼ Reference:

See the lecture and hands-on session for class 2 for a full discussion of Global, Local, and various Heuristic approaches to biomolecular sequence alignment.

[Barry J Grant](#).

[NW App Link](#)

YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

1. Limits of using BLAST [~10 mins]
2. Using PSI-BLAST [~30 mins]
3. Examining conservation patterns [~20 mins]
— BREAK [15 mins]—
4. [Optional] Using HMMER [~10 mins]
5. Divergence of protein sequence and structure [~25 mins]

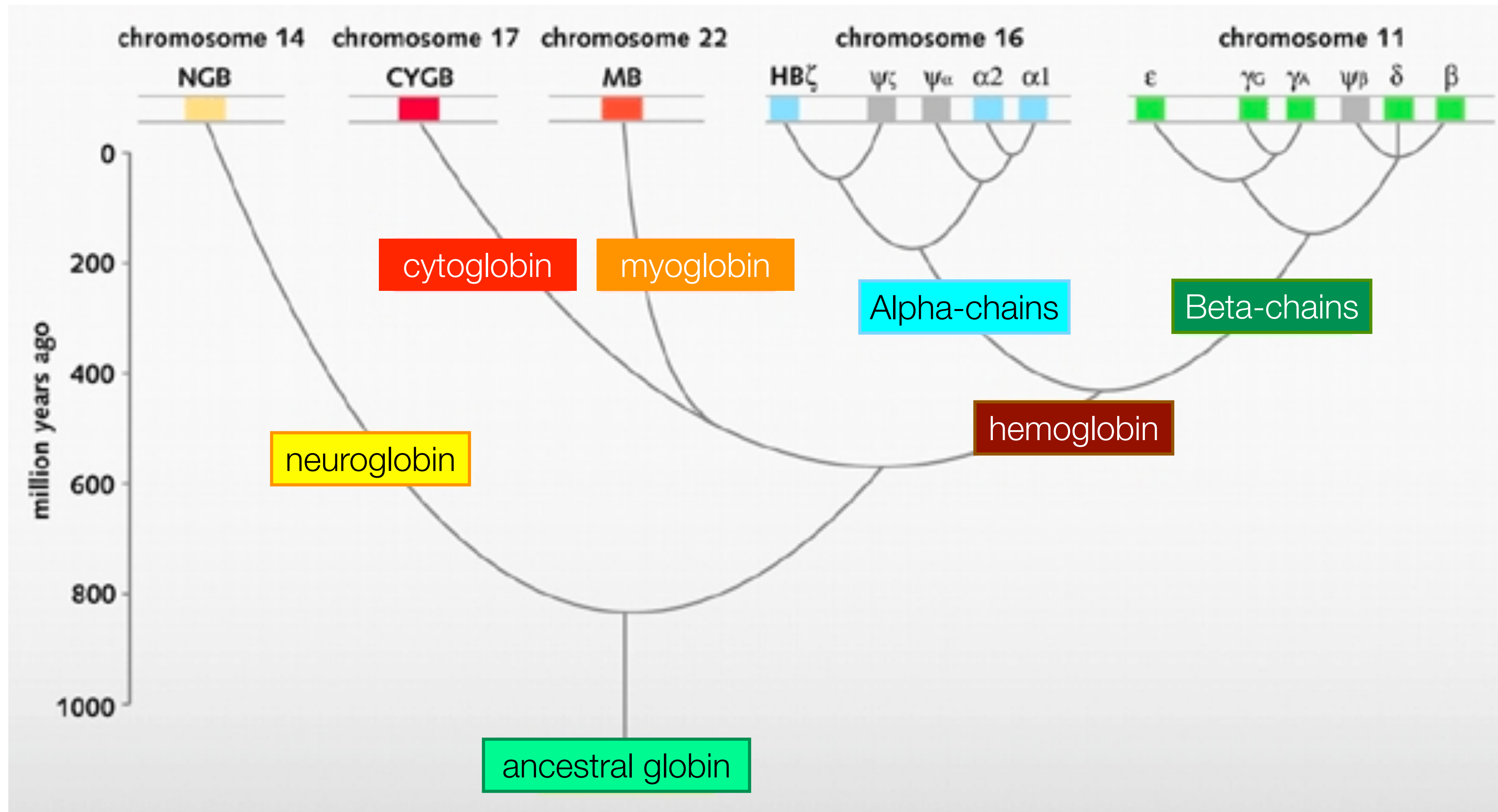
- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!

YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

- | | |
|---|------------|
| 1. Limits of using BLAST | [~10 mins] |
| 2. Using PSI-BLAST | [~30 mins] |
| 3. Examining conservation patterns | [~20 mins] |
| — BREAK [15 mins]— | |
| 4. [Optional] Using HMMER | [~10 mins] |
| 5. Divergence of protein sequence and structure | [~25 mins] |

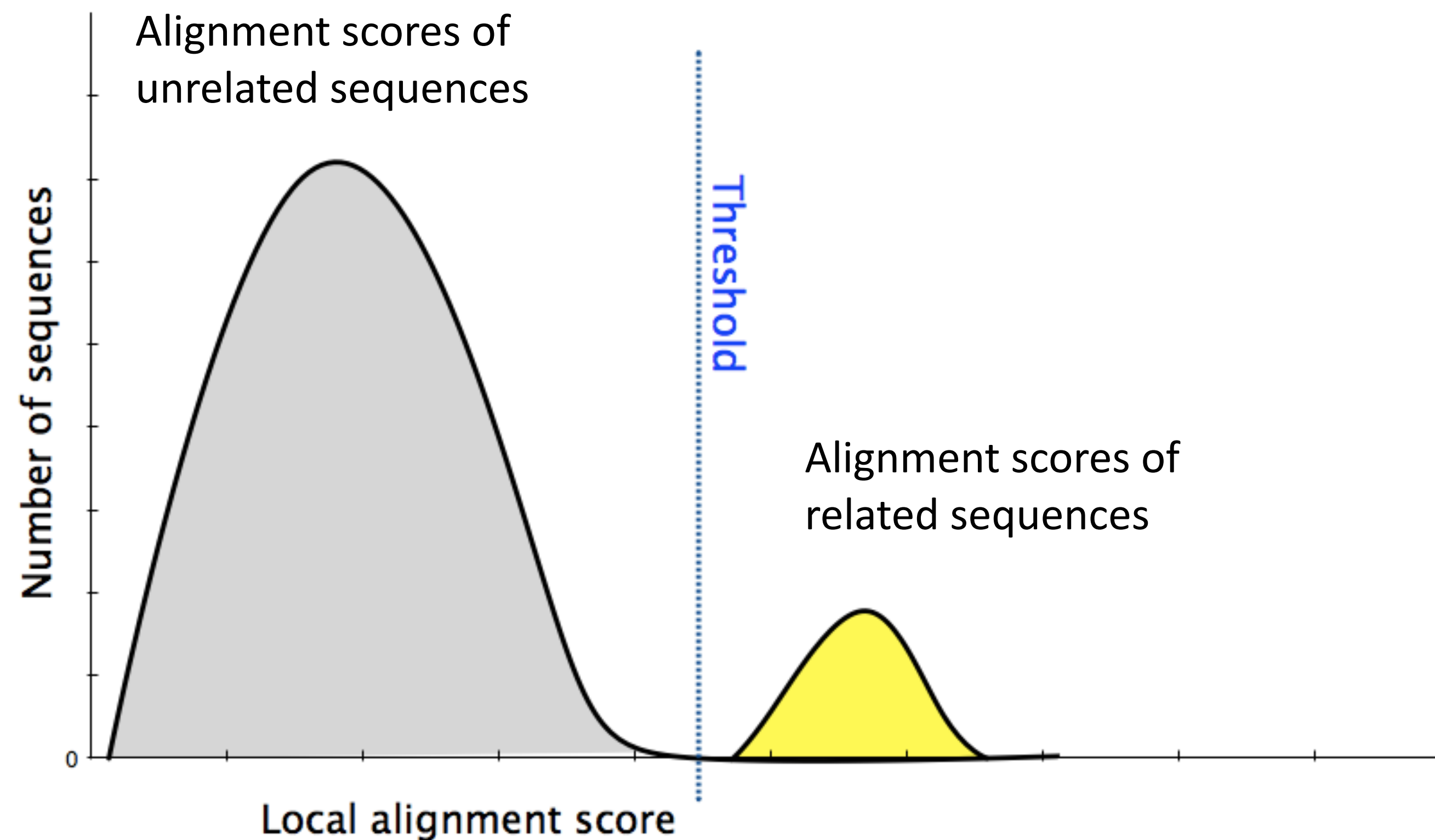
- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!



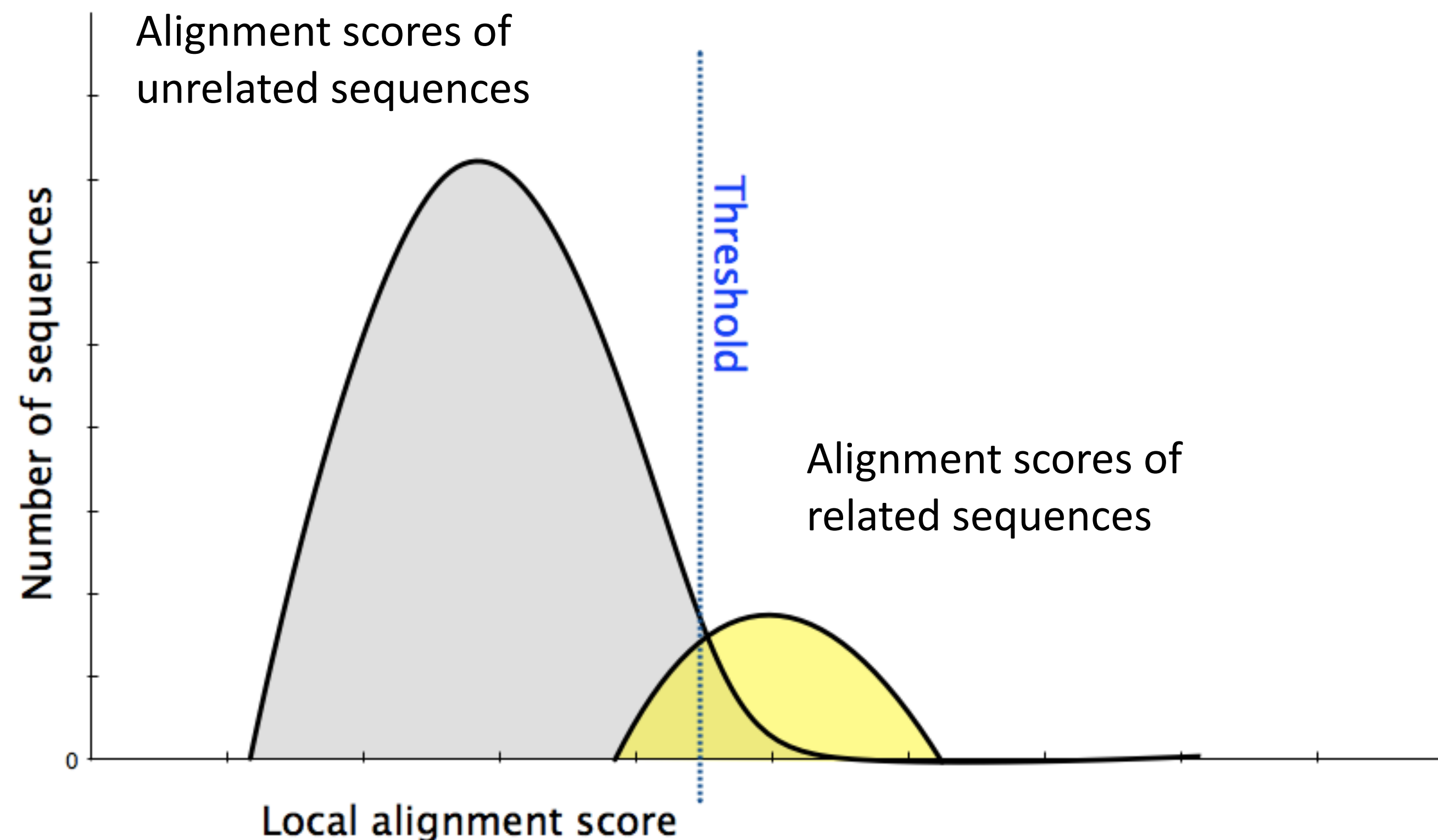
An evolutionary model of human globins.

The different locations of globin genes in human chromosomes are reported at the top of the figure, distinguishing between the functional genes (in color) and the pseudogenes (in grey).

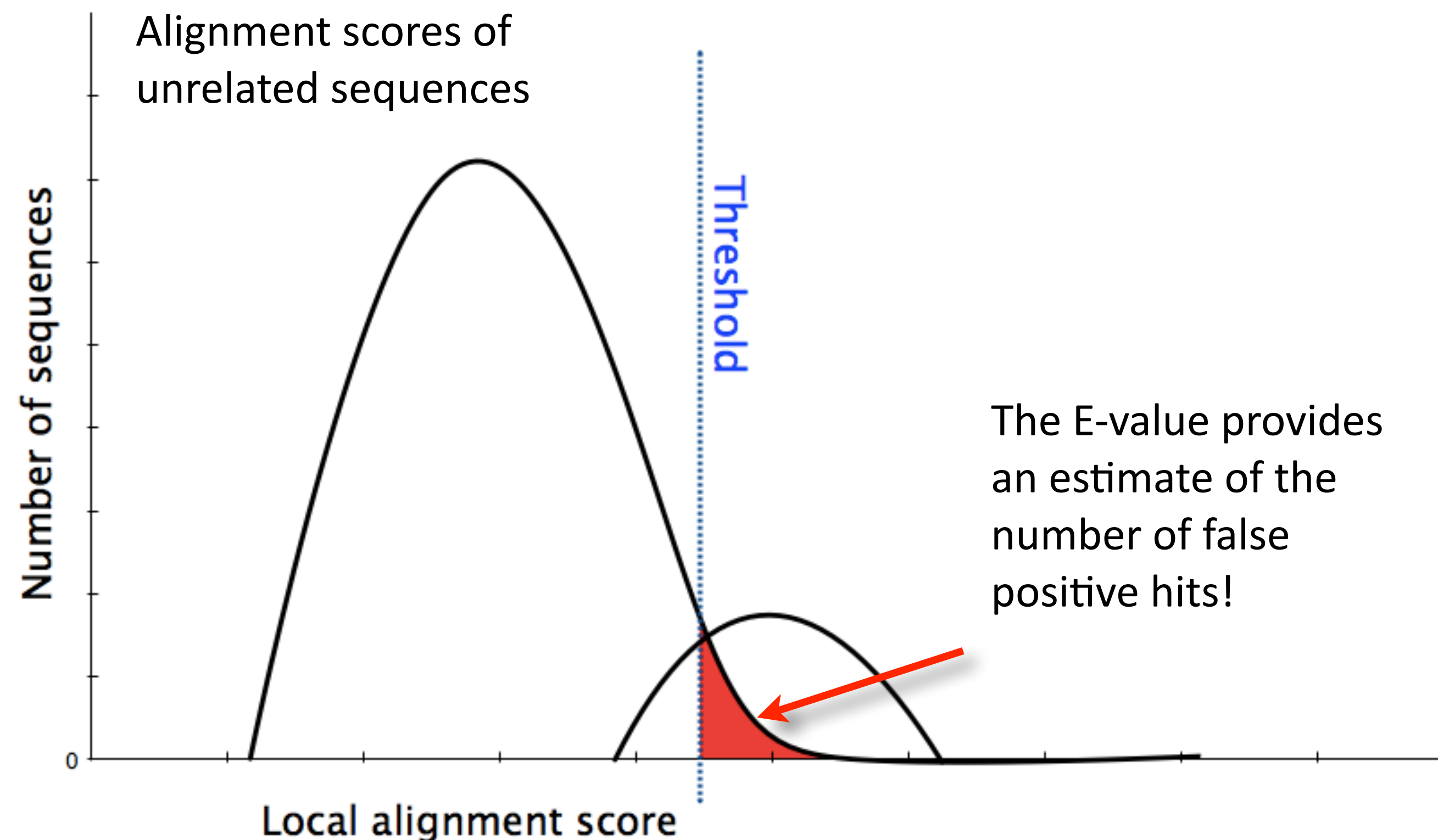
- Ideally, a threshold separates all query related sequences (yellow) from all unrelated sequences (gray)



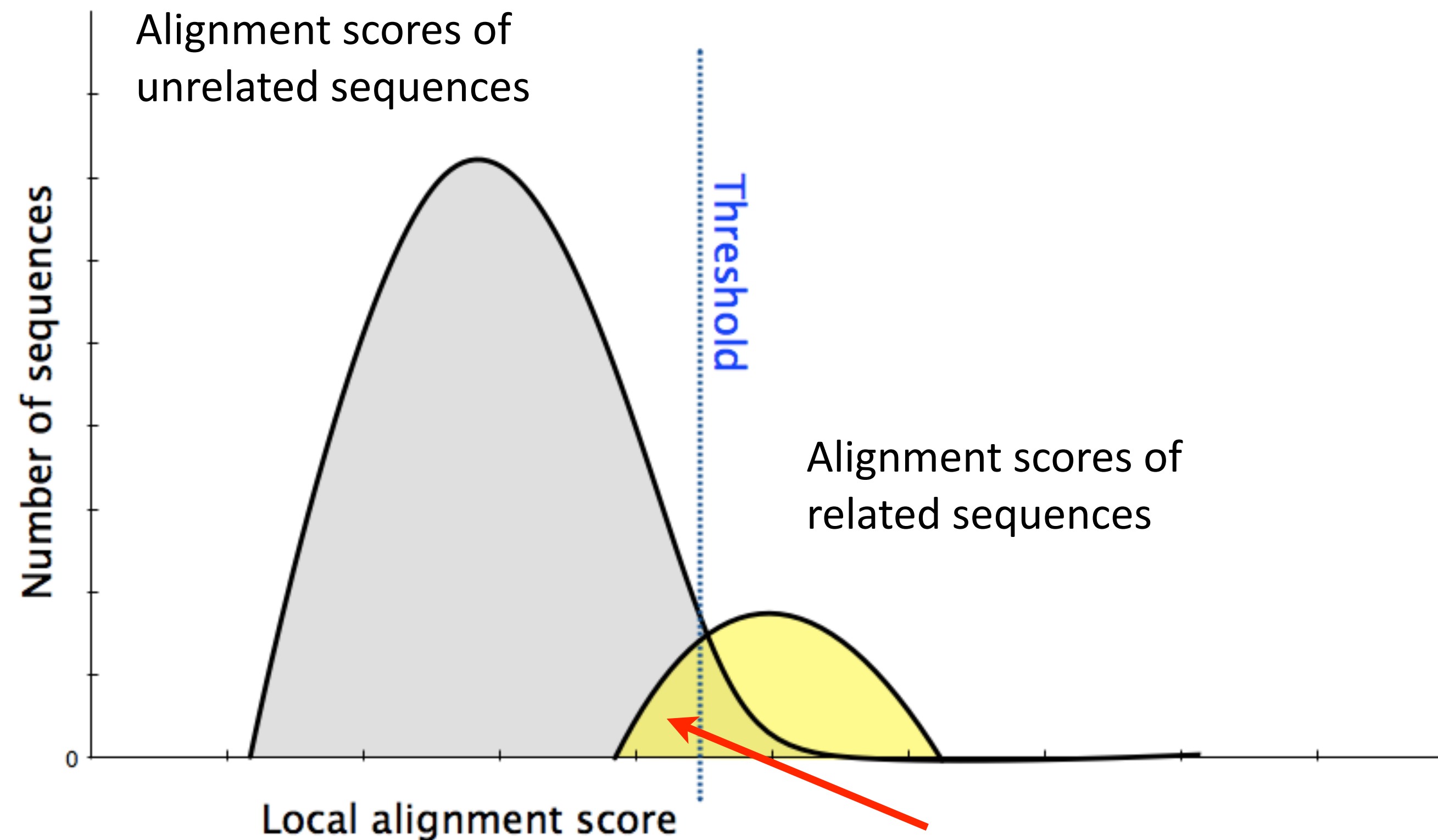
- Unfortunately, often both score distributions overlap
 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



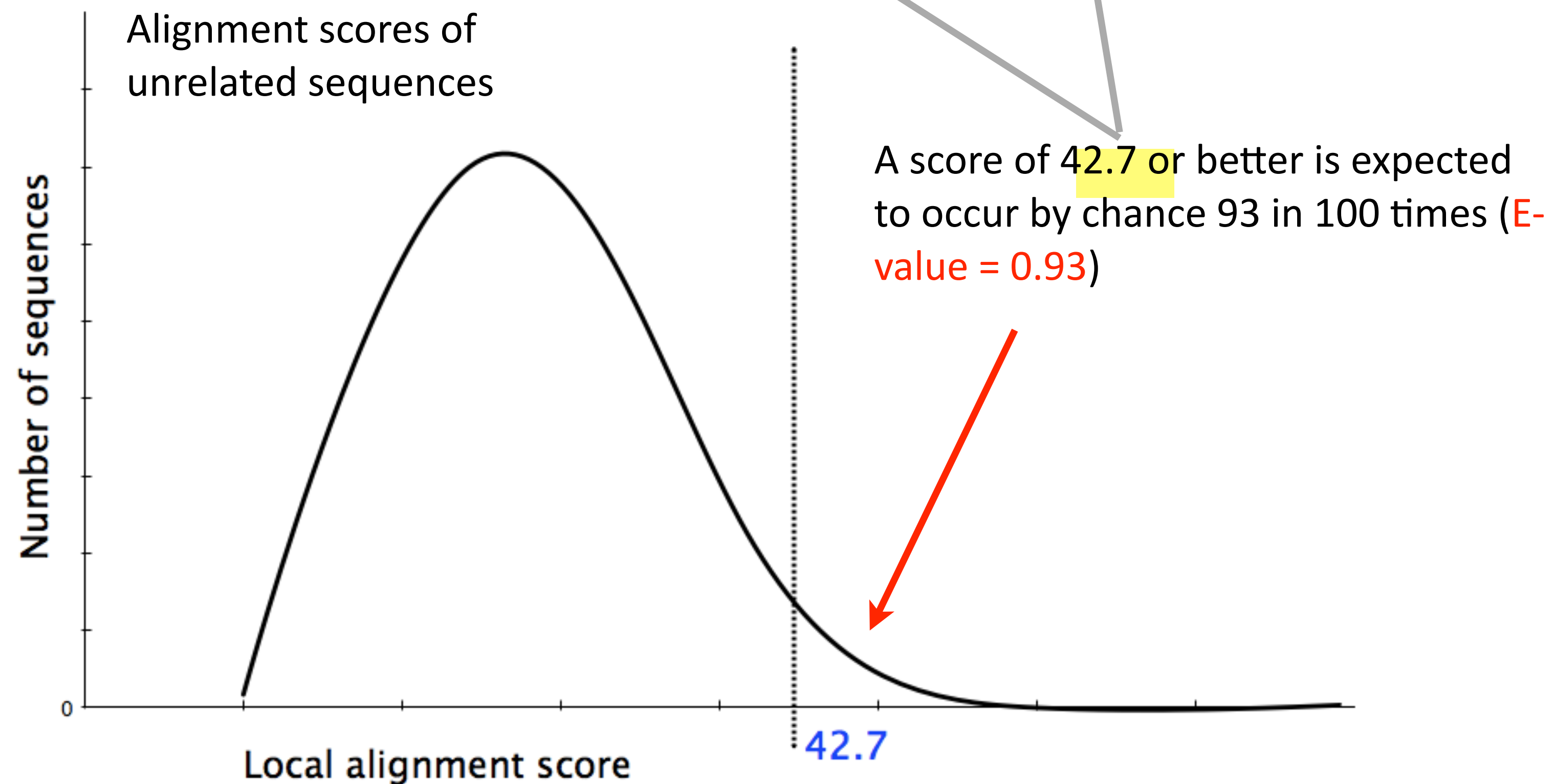
- Unfortunately, often both score distributions overlap
 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



- Maybe myoglobin, cytoglobin, neuroglobin etc. are found but not reported because of our E-value cutoff?
 - Lets change the cutoff and see...



Description	Max score	Query cover	E value	Max ident	Accession
hemoglobin subunit beta	284	100%	0	100%	NP_000510.1
hemoglobin subunit delta	240	100%	0	75.5%	NP_005321.1
hemoglobin subunit alpha	114	97%	0	43.45%	NP_000508.1
probable ATP-dependent RNA helicase	42.7	10%	0.93	32%	XP_011530405.1



E value: The number of alignments with a part

YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

1. Limits of using BLAST [~10 mins]

2. **Using PSI-BLAST** [~30 mins]

3. Examining conservation patterns [~20 mins]

— BREAK [15 mins]—

4. [Optional] Using HMMER [~10 mins]

5. Divergence of protein sequence and structure [~25 mins]

- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!

Recall: BLOUSM62 does not take the local context of a particular position into account

(*i.e.* all like substitutions are scored the same regardless of their location in the molecules).

▼ Algorithm parameters

Protein BLAST (BLASTp)

General Parameters

Max target sequences

100

Select the maximum number of aligned sequences to display ?

Short queries

☒ Automatically adjust parameters for short input sequences ?

Expect threshold

10 ?

Word size

3 ?

Max matches in a query range

0 ?

Scoring Parameters

Matrix

BLOSUM62 ?

Gap Costs

Existence: 11 Extension: 1 ?

Compositional adjustments

Conditional compositional score matrix adjustment ?

Filters and Masking

Filter

☐ Low complexity regions ?

Mask

☐ Mask for lookup table only ?

☐ Mask lower case letters ?

BLAST

Search **database Non-redundant protein sequences (nr)** using **Blastp**

☐ Show results in a new window

Scoring matrix
For match & mis-match scores

By default BLASTp match scores come from the BLOSUM62 matrix

C

S

T

P

A

G

N

D

E

Q

H

R

K

M

I

L

V

F

Y

W

9

-1

-1

-3

0

-3

-3

-3

-4

-3

-3

-3

-1

-1

-1

-2

-2

-2

-3

-2

4

1

-1

1

0

0

-1

-1

0

-1

-1

1

1

0

-2

-3

-3

0

-1

5

7

0

-2

-1

6

6

2

0

0

-1

0

-1

-2

3

-1

-1

-1

2

4

-1

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

-2

6

6

8

5

5

5

5

5

4

4

4

6

7

11

C

S

T

P

A

G

N

D

E

Q

H

R

K

M

I

L

V

F

Y

W

9

4

1

7

4

6

6

1

6

2

5

8

5

5

4

4

4

6

7

11

Blocks

Substitution

Matrix.

Scores obtained

observed frequencies of substitutions in block

sequences with no more than 62% identity.

Blocks **S**ubstitution **M**atrix. Scores obtained from observed frequencies of substitutions in blocks of aligned sequences with no more than 62% identity.

By default BLASTp match scores come from the BLOSUM62 matrix

C	9																			
S	-1	4																		
T	-1	1	5																	
P	-3	-1	-1	7																
A	0	1	0	-1	4															
G	-3	0	-2	-2	0	6														
N	-3	1	0	-2	-2	0	6													
D	-3	0	-1	-1	-2	-1	1	6												
E	-4	0	-1	-1	-1	-2	0	2	5											
Q	-3	0	-1	-1	-1	-2	0	0	2	5										
H	-3	-1	-2	-2	-2	-2	1	-1	0	0	8									
R	-3	-1	-1	-2	-1	-2	0	-2	0	1	0	5								
K	-3	0	-1	-1	-1	-2	0	-1	1	1	-1	2	5							
M	-1	-1	-1	-2	-1	-3	-2	-3	-2	0	-2	-1	-1	5						
I	-1	-2	-1	-3	-1	-4	-3	-3	-3	-3	-3	-3	-3	1	4					
L	-1	-2	-1	-3	-1	-4	-3	-4	-3	-2	-3	-2	-2	2	2	4				
V	-1	-2	0	-2	0	-3	-3	-3	-2	-2	-3	-3	-2	1	3	1	4			
F	-2	-2	-2	-4	-2	-3	-3	-3	-3	-3	-1	-3	-3	0	0	0	-1	6		
Y	-2	-2	-2	-3	-2	-3	-2	-3	-2	-1	2	-2	-2	-1	-1	-1	-1	3	7	
W	-2	-3	-2	-4	-3	-2	-4	-4	-3	-2	-2	-3	-3	-1	-3	-2	-3	1	2	11
	C	S	T	P	A	G	N	D	E	Q	H	R	K	M	I	L	V	F	Y	W

Note. All matches of Alanine score +4 regardless of their context in the molecule.

Note. All matches of Alanine for Alanine score +4 regardless of their position or context in the molecule.

PSI-BLAST: Position specific iterated BLAST

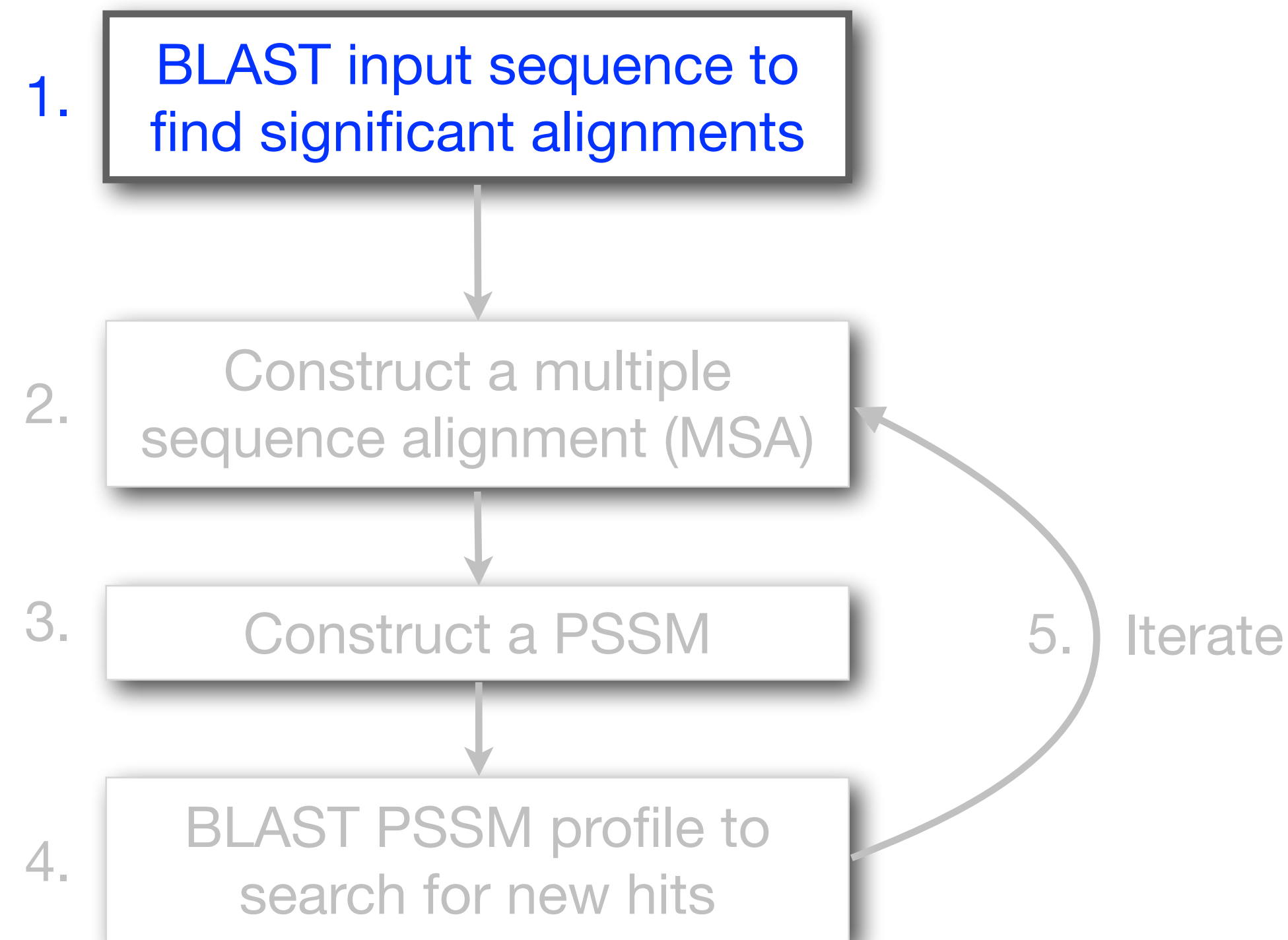
- The purpose of PSI-BLAST is to look deeper into the database for matches to your query protein sequence by employing a scoring matrix that is customized to your query

PSI-BLAST: Position specific iterated BLAST

- The purpose of PSI-BLAST is to look deeper into the database for matches to your query protein sequence by employing a scoring matrix that is customized to your query
 - PSI-BLAST constructs a multiple sequence alignment from the results of a first round BLAST search and then creates a “profile” or specialized **position-specific scoring matrix (PSSM)** for subsequent search rounds

PSI-BLAST: Position-Specific Iterated BLAST

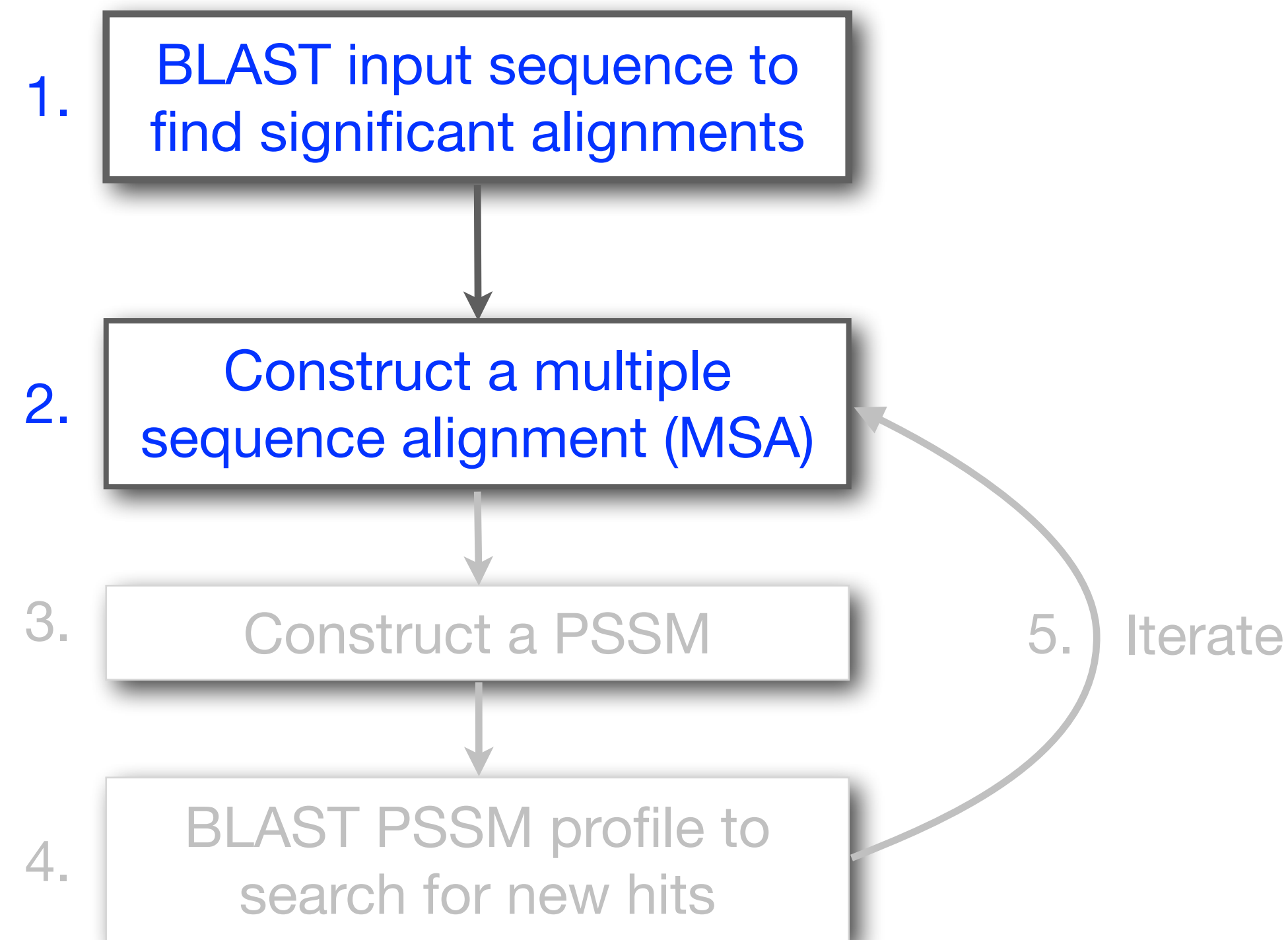
Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

PSI-BLAST: Position-Specific Iterated BLAST

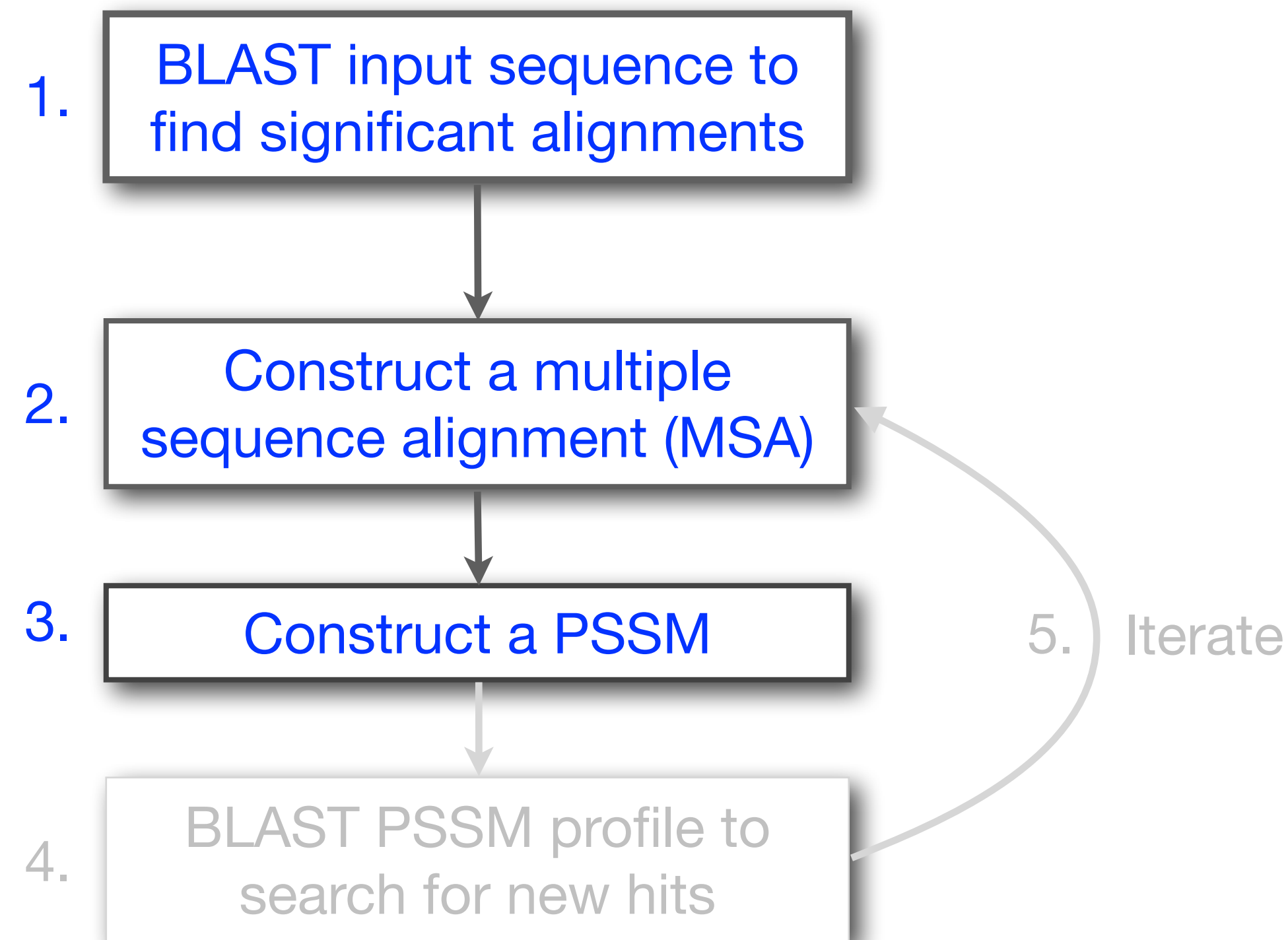
Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

PSI-BLAST: Position-Specific Iterated BLAST

Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

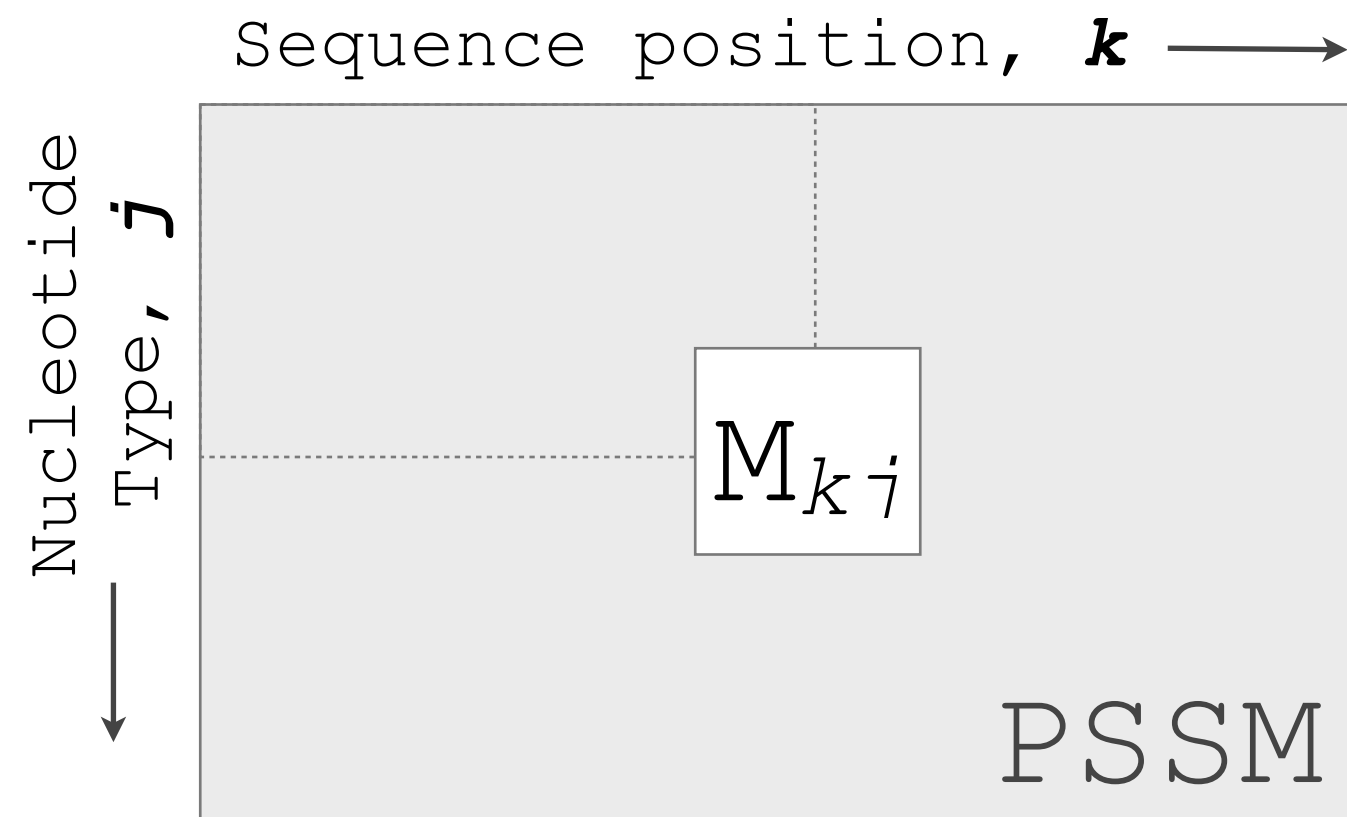
What is a **PSSM**?

What are PSSM sequence profiles?

A sequence profile is a **position-specific scoring matrix** (or **PSSM**, often pronounced 'possum') that gives a *quantitative* description of a set of aligned sequences.

PSSMs assign a score to a query sequence and are widely used for database searching.

A simple PSSM has as many columns as there are positions in the alignment, and either 4 rows (one for each DNA nucleotide) or 20 rows (one for each amino acid).



$$M_{kj} = \log \left(\frac{p_{kj}}{p_j} \right)$$

M_{kj} score for the j th nucleotide at position k

p_{kj} probability of nucleotide j at position k

p_j "background" probability of nucleotide j

Example: Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTTAGTATA
CCAAATTAGGAAA
CCAAATTGGCAAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Here we have **10 aligned** transcription factor binding site nucleotide sequences

That span **13 positions** (i.e. columns of nucleotides).

We will build a 13 x 4 **PSSM** ($k=13$, $j=4$).

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTTCGAAAA
CCTATTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

First we will build an alignment **Counts matrix**

[illegible]

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTAGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:													
C:													
G:													
T:													

Position k = 1

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0												
C:	9												
G:	0												
T:	1												

Position k = 1

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCATTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0												
C:	9												
G:	0												
T:	1												
Consensus	C												

Position k = 1

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCATTTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0											
C:	9	10											
G:	0	0											
T:	1	0											
Consensus	C	C											

Position k = 2

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	6										
C:	9	10	1										
G:	0	0	0										
T:	1	0	3										
Consensus	C	C	[AT]										

Position k = 3

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTAGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	6	10	5	0	1	5	0	3	10	8	10
C:	9	10	1	0	0	0	0	2	1	1	0	0	0
G:	0	0	0	0	0	0	0	1	9	5	0	0	0
T:	1	0	3	0	5	10	9	2	0	1	0	2	0
Consensus	C	C	[AT]	A	[AT]	T	T	[ACT]	G	[GA]	A	[AT]	A

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
CCTATTAAGAAAA
CCAAATTAGGAAA
CCAAATTCGGATA
CCCATTTCGAAAA
CCTATTTAGGTATA
CCAAATTAGGAAA
CCAAATTGGCAAA
TCTATTTTGGAAA
CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	6	10	5	0	1	5	0	3	10	8	10
C:	9	10	1	0	0	0	0	2	1	1	0	0	0
G:	0	0	0	0	0	0	0	1	9	5	0	0	0
T:	1	0	3	0	5	10	9	2	0	1	0	2	0
Consensus	C	C	[AT]	A	[AT]	T	T	[ACT]	G	[GA]	A	[AT]	A

Often we will not communicate with the count matrix but rather the derived **average profile** (a.k.a. frequency matrix).

Average Profile (Frequency) matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	0.6	1	0.5	0	0.1	0.5	0	0.3	1	0.8	1
C:	0.9	1	0.1	0	0	0	0	0.2	0.1	0.1	0	0	0
G:	0	0	0	0	0	0	0	0.1	0.9	0.5	0	0	0
T:	0.1	0	0.3	0	0.5	1	0.9	0.2	0	0.1	0	0.2	0
Consensus	C	C	[AT]	A	[AT]	T	T	[ACT]	G	[GA]	A	[AT]	A

Computing a transcription factor bind site PSSM

CCAAATTAGGAAA
 CCTATTAAGAAAA
 CCAAATTAGGAAA
 CCAAATTCGGATA
 CCCATTTTCGAAAA
 CCTATTTAGGTATA
 CCAAATTAGGAAA
 CCAAATTGGCAAAA
 TCTATTTTGGAAA
 CCAATTTTCAAAA

Alignment Counts matrix:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	6	10	5	0	1	5	0	3	10	8	10
C:	9	10	1	0	0	0	0	2	1	1	0	0	0
G:	0	0	0	0	0	0	0	1	9	5	0	0	0
T:	1	0	3	0	5	10	9	2	0	1	0	2	0
Consensus	C	C	[AT]	A	[AT]	T	T	[ACT]	G	[GA]	A	[AT]	A

Or the "score (M_{kj}) matrix" = PSSM

C_{kj} Number of j th type nucleotide at position k

Z Total number of aligned sequences

p_j "background" probability of nucleotide j

p_{kj} probability of nucleotide j at position k

$$M_{kj} = \log \left(\frac{p_{kj}}{p_j} \right) \quad p_{kj} = \frac{C_{kj} + p_j}{Z + 1}$$

$$M_{kj} = \log \left(\frac{C_{kj} + p_j / Z + 1}{p_j} \right)$$

Computing a transcription factor bind site PSSM...

Alignment Matrix: C_{kj}

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	0	0	6	10	5	0	1	5	0	3	10	8	10
C:	9	10	1	0	0	0	0	2	1	1	0	0	0
G:	0	0	0	0	0	0	0	1	9	5	0	0	0
T:	1	0	3	0	5	10	9	2	0	1	0	2	0

$$k=1, j=A: M_{kj} = \log\left(\frac{C_{kj} + p_j / Z + 1}{p_j}\right) = \log\left(\frac{0 + 0.25 / 10 + 1}{0.25}\right) = -2.4$$

$$k=1, j=C: M_{kj} = \log\left(\frac{C_{kj} + p_j / Z + 1}{p_j}\right) = \log\left(\frac{9 + 0.25 / 10 + 1}{0.25}\right) = 1.2$$

$$k=1, j=T: M_{kj} = \log\left(\frac{C_{kj} + p_j / Z + 1}{p_j}\right) = \log\left(\frac{1 + 0.25 / 10 + 1}{0.25}\right) = -0.8$$

PSSM: M_{kj}

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	-2.4	-2.4	0.8	1.3	0.6	-2.4	-0.8	0.6	-2.4	0.2	1.3	1.1	1.3
C:	1.2	1.3	-0.8	-2.4	-2.4	-2.4	-2.4	-0.2	-0.8	-0.8	-2.4	-2.4	-2.4
G:	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-0.8	1.2	0.6	-2.4	-2.4	-2.4
T:	-0.8	-2.4	0.2	-2.4	0.6	1.3	1.2	-0.2	-2.4	-0.8	-2.4	-0.2	-2.4

Scoring a test sequence

Query Sequence

CCTATTAGGATA

PSSM:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	-2.4	-2.4	0.8	1.3	0.6	-2.4	-0.8	0.6	-2.4	0.2	1.3	1.1	1.3
C:	1.2	1.3	-0.8	-2.4	-2.4	-2.4	-2.4	-0.2	-0.8	-0.8	-2.4	-2.4	-2.4
G:	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-0.8	1.2	0.6	-2.4	-2.4	-2.4
T:	-0.8	-2.4	0.2	-2.4	0.6	1.3	1.2	-0.2	-2.4	-0.8	-2.4	-0.2	-2.4
Test seq:	C	C	T	A	T	T	T	A	G	G	A	T	A

$$\begin{aligned}\text{Query Score} &= 1.2 + 1.3 + 0.2 + 1.3 + 0.6 + 1.3 + 1.2 \\ &\quad + 0.6 + 1.2 + 0.6 + 1.3 + -0.2 + 1.3 \\ &= 11.9\end{aligned}$$

Scoring a test sequence

Query Sequence

CCTATTAGGATA

PSSM:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	-2.4	-2.4	0.8	1.3	0.6	-2.4	-0.8	0.6	-2.4	0.2	1.3	1.1	1.3
C:	1.2	1.3	-0.8	-2.4	-2.4	-2.4	-2.4	-0.2	-0.8	-0.8	-2.4	-2.4	-2.4
G:	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-0.8	1.2	0.6	-2.4	-2.4	-2.4
T:	-0.8	-2.4	0.2	-2.4	0.6	1.3	1.2	-0.2	-2.4	-0.8	-2.4	-0.2	-2.4
Test seq:	C	C	T	A	T	T	T	A	G	G	A	T	A

$$\begin{aligned}\text{Query Score} &= 1.2 + 1.3 + 0.2 + 1.3 + 0.6 + 1.3 + 1.2 \\ &\quad + 0.6 + 1.2 + 0.6 + 1.3 + -0.2 + 1.3 \\ &= 11.9\end{aligned}$$

Q. Does the query sequence match the DNA sequence profile?

Scoring a test sequence...

Query Sequence

CCTATTAGGATA

Best Possible Sequence

CCAATTAGGAAA

PSSM:

Position k =	1	2	3	4	5	6	7	8	9	10	11	12	13
A:	-2.4	-2.4	0.8	1.3	0.6	-2.4	-0.8	0.6	-2.4	0.2	1.3	1.1	1.3
C:	1.2	1.3	-0.8	-2.4	-2.4	-2.4	-2.4	-0.2	-0.8	-0.8	-2.4	-2.4	-2.4
G:	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-2.4	-0.8	1.2	0.6	-2.4	-2.4	-2.4
T:	-0.8	-2.4	0.2	-2.4	0.6	1.3	1.2	-0.2	-2.4	-0.8	-2.4	-0.2	-2.4
Max Score:	C	C	A	A	T	T	T	A	G	G	A	A	A

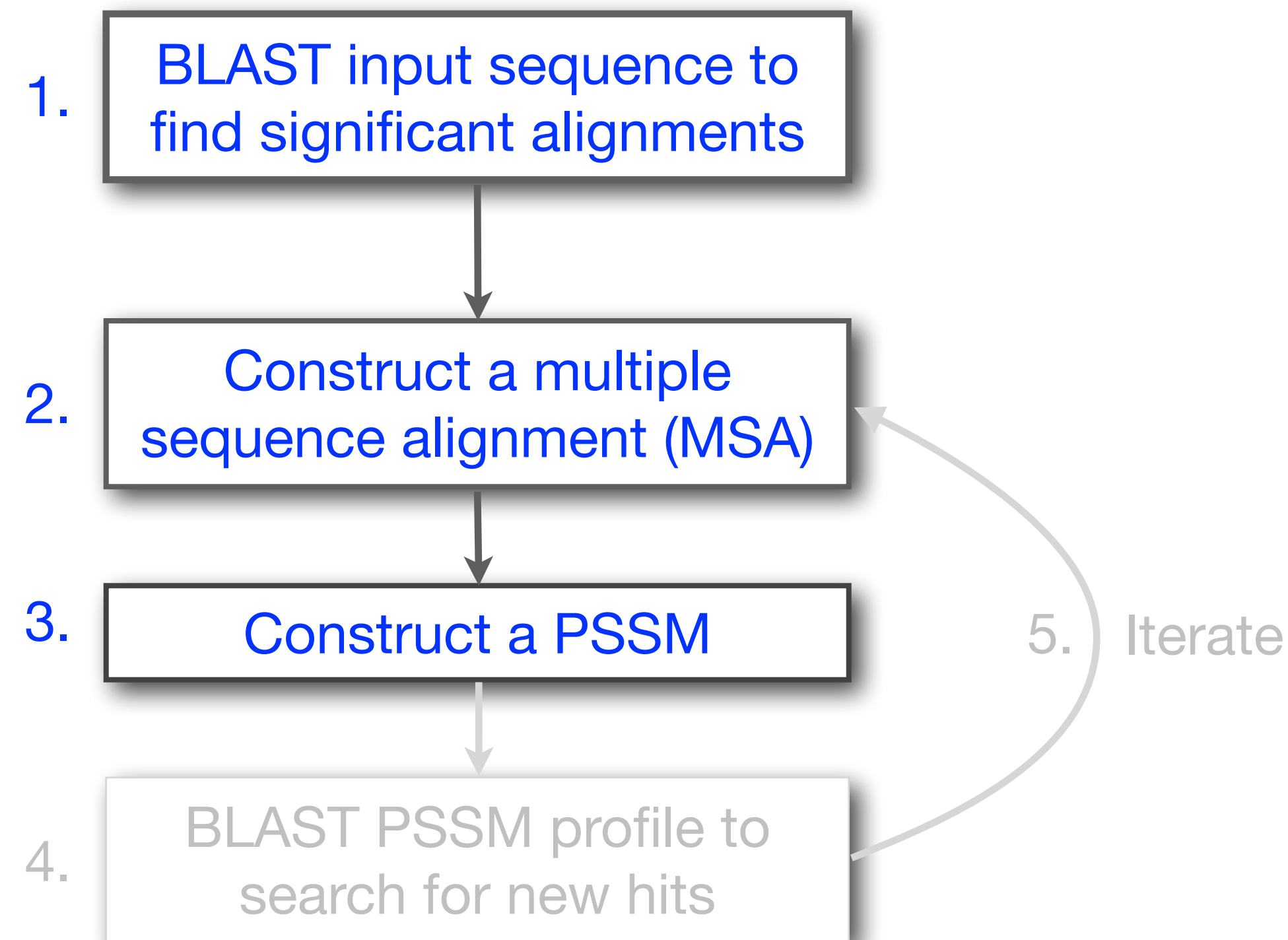
$$\begin{aligned}
 \text{Max Score} &= 1.2 + 1.3 + 0.8 + 1.3 + 0.6 + 1.3 + 1.2 \\
 &\quad + 0.6 + 1.2 + 0.6 + 1.3 + 1.1 + 1.3 \\
 &= 13.8
 \end{aligned}$$

A. Following method in Harbison *et al.* (2004) Nature 431:99-104

Heuristic threshold for match = 60% x Max Score = (0.6 x 13.8 = 8.28);
 11.9 > 8.28; Therefore our query is a potential TFBS!

PSI-BLAST: Position-Specific Iterated BLAST

Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

Inspect the blastp output to identify empirical “rules” regarding amino acids tolerated at each position

730496	66	FTVDENGQMSATAKGRVRLFNNWDVCADMIGSFTDTE	PAKFKMKYWG	VASFLQKG	NDDH	125		
200679	63	FSVDEKGHMSATAKGRVRLLSNWEVCADMVGTF	TDTE	PAKFKMKYWG	VASFLQ	RG	NDDH	122
206589	34	FSVDEKGHMSATAKGRVRLLSNWEVCADMVGTF	TDTE	PAKFKMKYWG	VASFLQ	RG	NDDH	93
2136812	2	MSATAKGRVRLLSNWDVCADMVGTF	TDTE	PAKFKMKYWG	VASFLQ	KG	NDDH	53
132408	65	FKIEDNGKTTATAKGRVRILDKLELCANMVGTF	FIETND	PAKYRMKYHG	ALAILER	GL	DDH	124
267584	44	FSVDESGKVTATAHGRVILNNWEMCANMFGTF	FEDTPD	PAKFKMRYWG	AASYLQ	TG	NDDH	103
267585	44	FSVDGSGKVTATAQGRVILNNWEMCANMFGTF	FEDTPD	PAKFKMRYWG	AAAYLQ	SG	NDDH	103
8777608	63	FTIHEDGAMTATAKGRVILNNWEMCADMMATF	ETTPD	PAKFRMRYWG	AASYLQ	TG	NDDH	122
6687453	60	FKVEEDGTMTATAIGRVILNNWEMCANMFGTF	FEDTE	PAKFKMKYWG	AAAYLQ	TG	YDDH	119
10697027	81	FKVQEDGTMTATATGRVILNNWEMCANMFGTF	FEDTE	EPARFKMKYWG	AAAYLQ	TG	YDDH	140
13645517	1	MVGTF	TDTE	PAKFKMKYWG	VASFLQ	KG	NDDH	32
13925316	38	FSVDGSGKMTATAQGRVILNNWEMCANMFGTF	FEDTPD	PAKFKMRYWG	AAAYLQ	SG	NDDH	97
131649	65	YTVEEDGTMTASSKGRVKLFGFVVICADMAA	QYTDPT	PAKMYMTYQ	GLASYL	SS	GGDNY	126

↑
M

↑
N,M,L,Y,G

		A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	M	-1	-2	-2	-3	-2	-1	-2	-3	-2	1	2	-2	6	0	-3	-2	-1	-2	-1	1
2	K	-1	1	-2	-1	-1	-2	-1	-2	-2	3	3	3	2	-1	1	0	1	3	2	3
3	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-3	-3	-2	1	-4	-3	-3	12	2	-3
4	V	0	-3	-3	-4	-1	-3	-3	-4	-3	-3	-3	-3	-2	-1	-4	-2	0	-3	-1	4
5	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-3	-3	-2	1	-4	-3	-3	12	2	-3
6	A	5	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0
7	L	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
8	L	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
9	L	-1	-3	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	2
10	L	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
11	A	5	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0
12	A	5	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0
13	W	-2	-2	-3	-3	-1	-2	-2	-2	-2	-1	-1	-1	3	2	1	-3	-3	-2	7	0
14	A	3	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1	-2	-3	-1	1	-1	-3	-3
15	A	2	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	0	-2	-3	-1	3	0	-3	-2
16	A	4	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1	-1	-3	-1	1	0	-3	-2
...																					
37	S	2	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	0	-2	-3	-1	4	1	-3	-2
38	G	0	-3	-1	-2	-3	-2	-2	6	-2	-4	-4	-2	-3	-4	-2	0	-2	-3	-3	-4
39	T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5	-3	-2	0
40	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-2	-3	-2	1	-4	-3	-3	9	2	-3
41	Y	-2	-2	-2	-3	-3	-2	-2	-3	2	-2	-1	-2	-1	3	-3	-2	-2	2	7	-1
42	A	4	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0

		A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	M	-1	-2	-2	-3	-2	-1	-2	-3	-2	1	2	-2	6	0	-3	-2	-1	-2	-1	1
2	K	-1	1	0	1	-4	2	4	-2	0	-3	-3	3	-2	-4	-1	0	-1	-3	-2	-3
3	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-2	-3	-2	1	-4	-3	-3	12	2	-3
4	V	0	-3	-3	-4	-1	-3	-3	-4	-4	3	1	-3	1	-1	-3	-2	0	-3	-1	4
5	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-2	-3	-2	1	-4	-3	-3	12	2	-3
6	A	5	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0
7	L	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
8	L	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
9	L	-1	-3	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	2
10	L	-2	-2	-4	-4	-	-	-	-	-	-	-	-	-	-	-	-3	-1	-2	-1	1
11	A	5	-2	-2	-2	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	0
12	A	5	-2	-2	-2	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	0
13	W	-2	-3	-4	-4	-	-	-	-	-	-	-	-	-	-	-	-3	-2	7	0	0
14	A	3	-2	-1	-2	-	-	-	-	-	-	-	-	-	-	-	1	-1	-3	-3	-1
15	A	2	-1	0	-1	-	-	-	-	-	-	-	-	-	-	-	3	0	-3	-2	-2
16	A	4	-2	-1	-	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	-1
...																					
37	S	2	-1	0	-	-	-	-	-	-	-	-	-	-	-	-	4	1	-3	-2	-2
38	G	0	-3	-1	-2	-	-	-	-	-	-	-	-	-	-	-	0	-2	-3	-3	-4
39	T	0	-1	0	-1	-	-	-	-	-	-	-	-	-	-	-	1	5	-3	-2	0
40	W	-3	-3	-4	-5	-	-	-	-	-	-	-	-	-	-	-	-3	-3	9	2	-3
41	Y	-2	-2	-2	-3	-	-	-	-	-	-	-	-	-	-	-	-2	-2	2	7	-1
42	A	4	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0

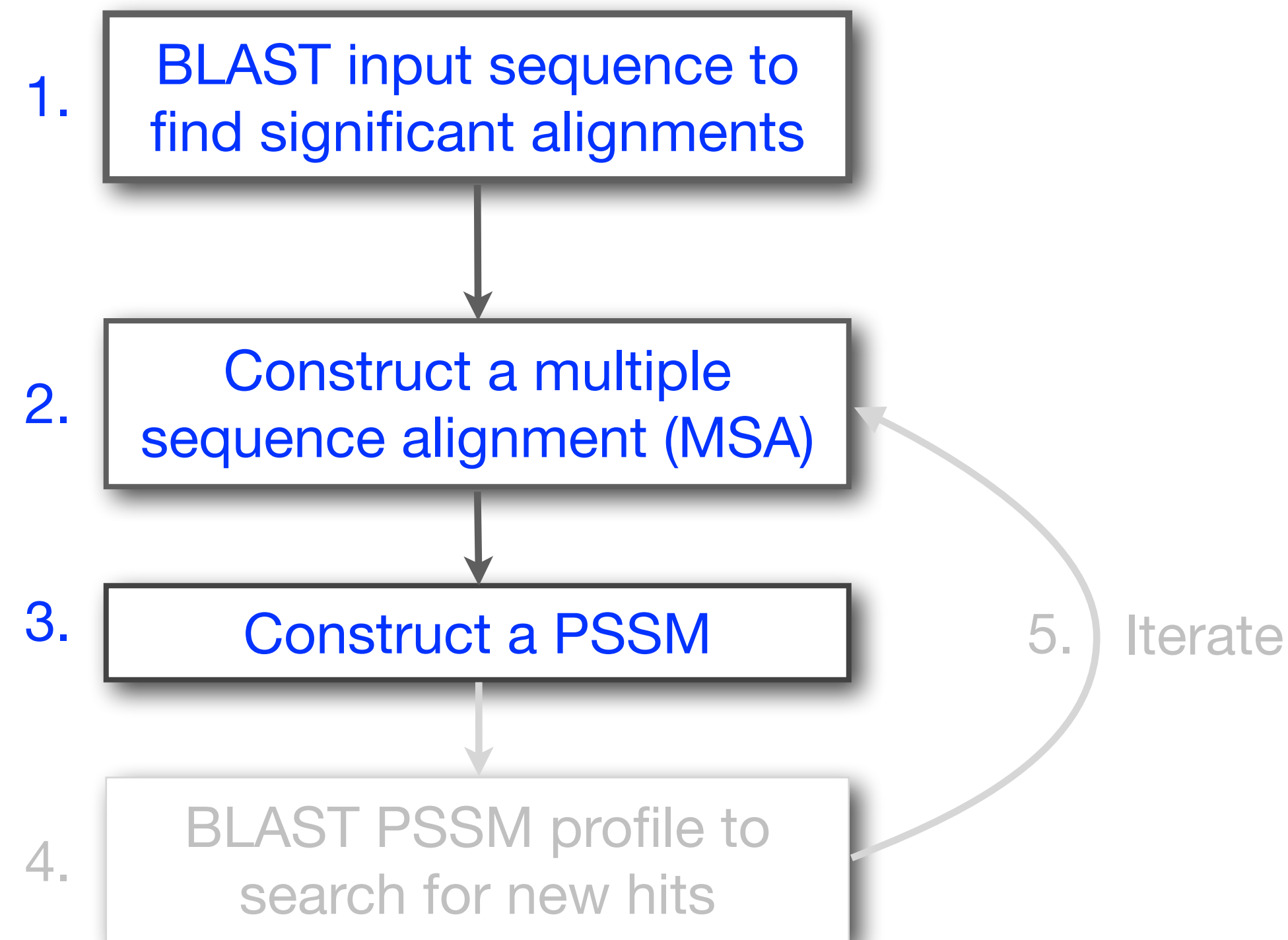
Note: A given amino acid (such as alanine) in your query protein can receive different scores for matching alanine depending on the position in the protein (BLOSUM $S_{AA} = +4$)

		A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1	M	<p>The PSI-BLAST PSSM is essentially a query customized scoring matrix that is more sensitive than BLOSUM.</p>																			
2	K																				
3	W																				
4	V																				
5	W	-3	-3	-4	-5	-3	-2	-3	-3	-3	-3	-2	-3	-2	1	-4	-3	-3	12	2	-3
6	A	5	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0
7	L	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
8	L	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
9	L	-1	-3	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	2
10	L	-2	-2	-4	-4	-	-	-	-	-	-	-	-	-	-	-	-3	-1	-2	-1	1
11	A	5	-2	-2	-2	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	0
12	A	5	-2	-2	-2	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	0
13	W	-2	-3	-4	-4	-	-	-	-	-	-	-	-	-	-	-	-3	-2	7	0	0
14	A	3	-2	-1	-2	-	-	-	-	-	-	-	-	-	-	-	1	-1	-3	-3	-1
15	A	2	-1	0	-1	-	-	-	-	-	-	-	-	-	-	-	3	0	-3	-2	-2
16	A	4	-2	-1	-	-	-	-	-	-	-	-	-	-	-	-	1	0	-3	-2	-1
...																					
37	S	2	-1	0	-	-	-	-	-	-	-	-	-	-	-	-	4	1	-3	-2	-2
38	G	0	-3	-1	-2	-	-	-	-	-	-	-	-	-	-	-	0	-2	-3	-3	-4
39	T	0	-1	0	-1	-	-	-	-	-	-	-	-	-	-	-	1	5	-3	-2	0
40	W	-3	-3	-4	-5	-	-	-	-	-	-	-	-	-	-	-	-3	-3	9	2	-3
41	Y	-2	-2	-2	-3	-	-	-	-	-	-	-	-	-	-	-	-2	-2	2	7	-1
42	A	4	-2	-2	-2	-1	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0

Note: A given amino acid (such as alanine) in your query protein can receive different scores for matching alanine depending on the position in the protein (BLOSUM $S_{AA} = +4$)

PSI-BLAST: Position-Specific Iterated BLAST

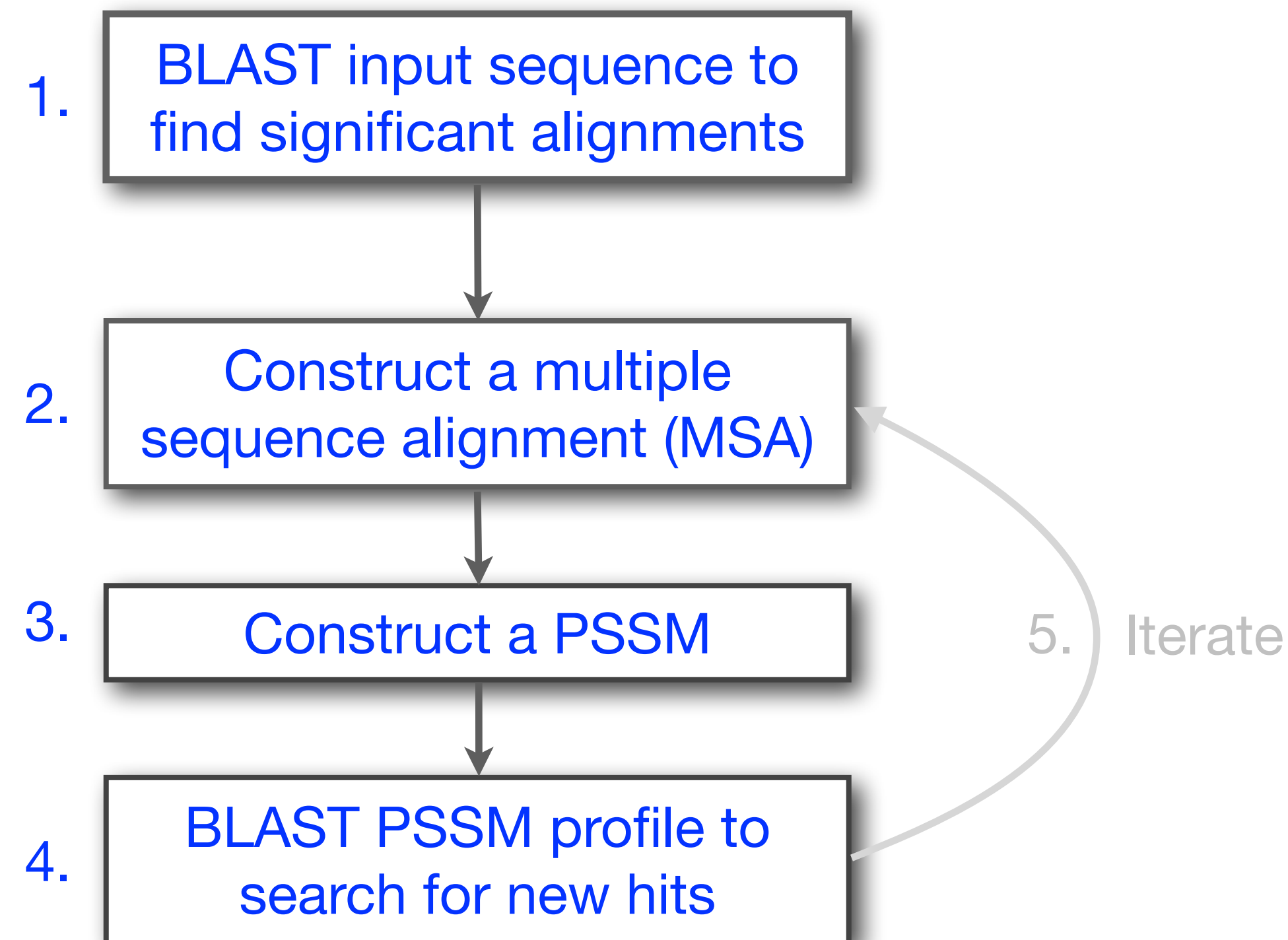
Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

PSI-BLAST: Position-Specific Iterated BLAST

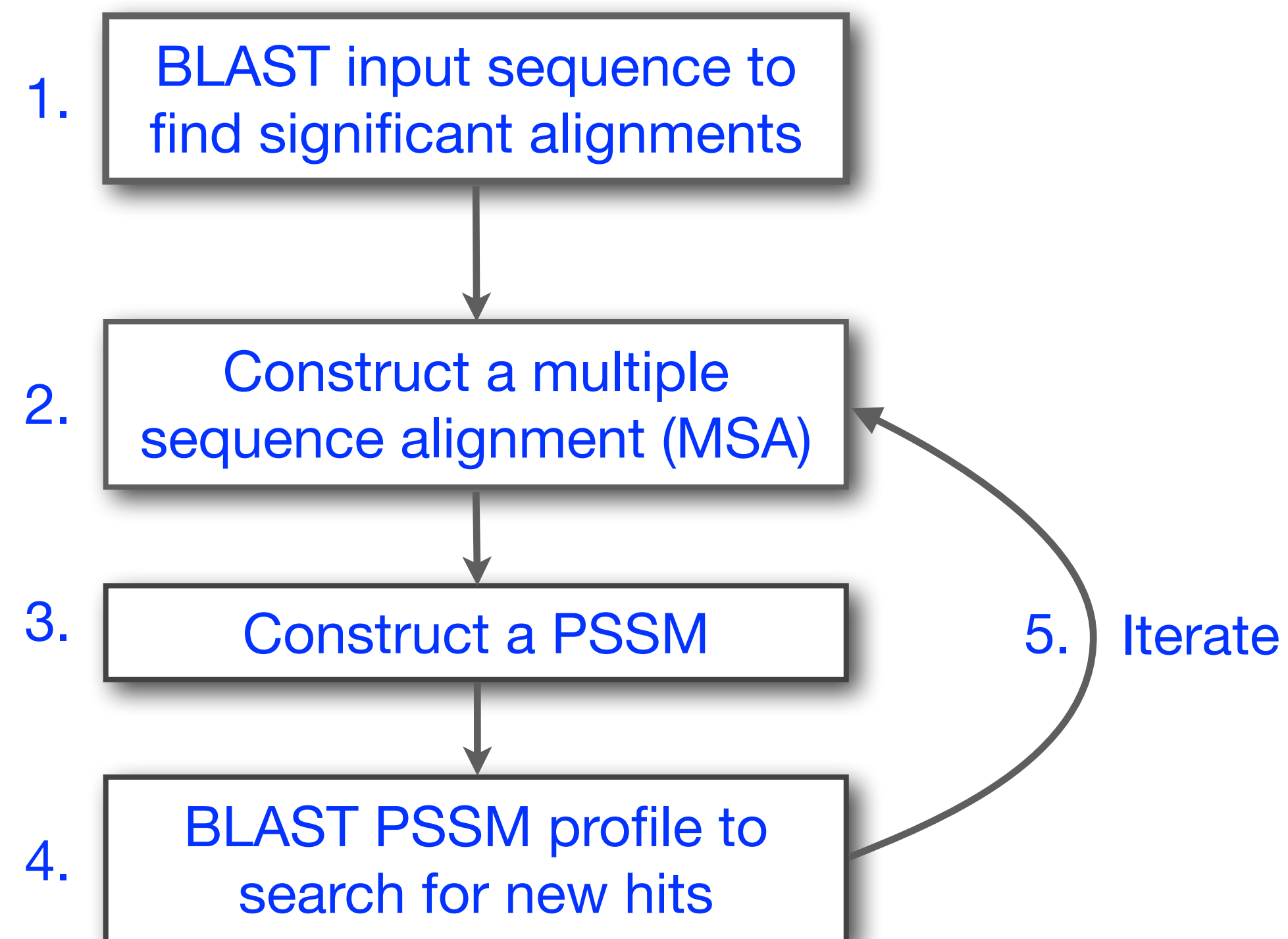
Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



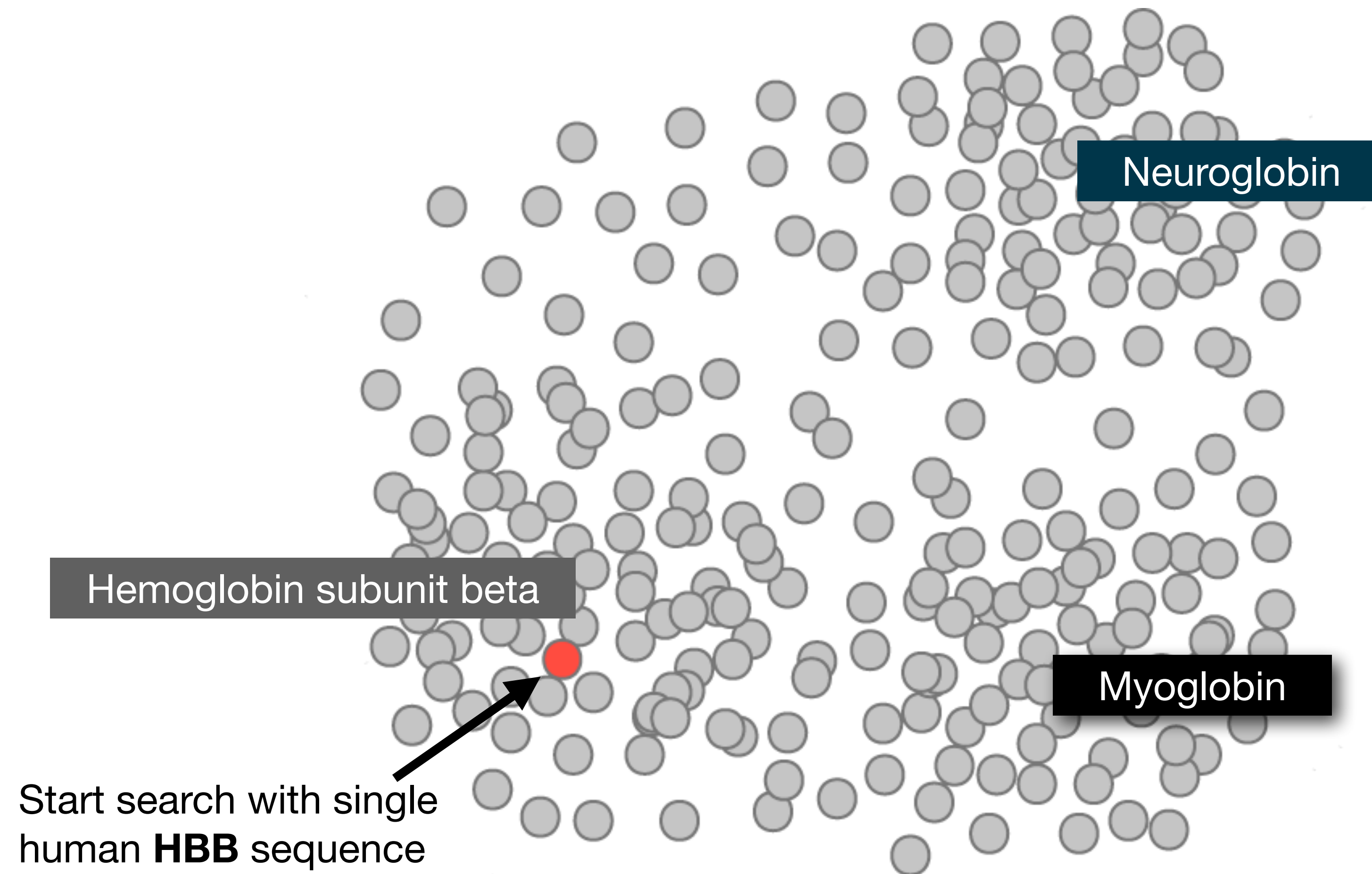
(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)

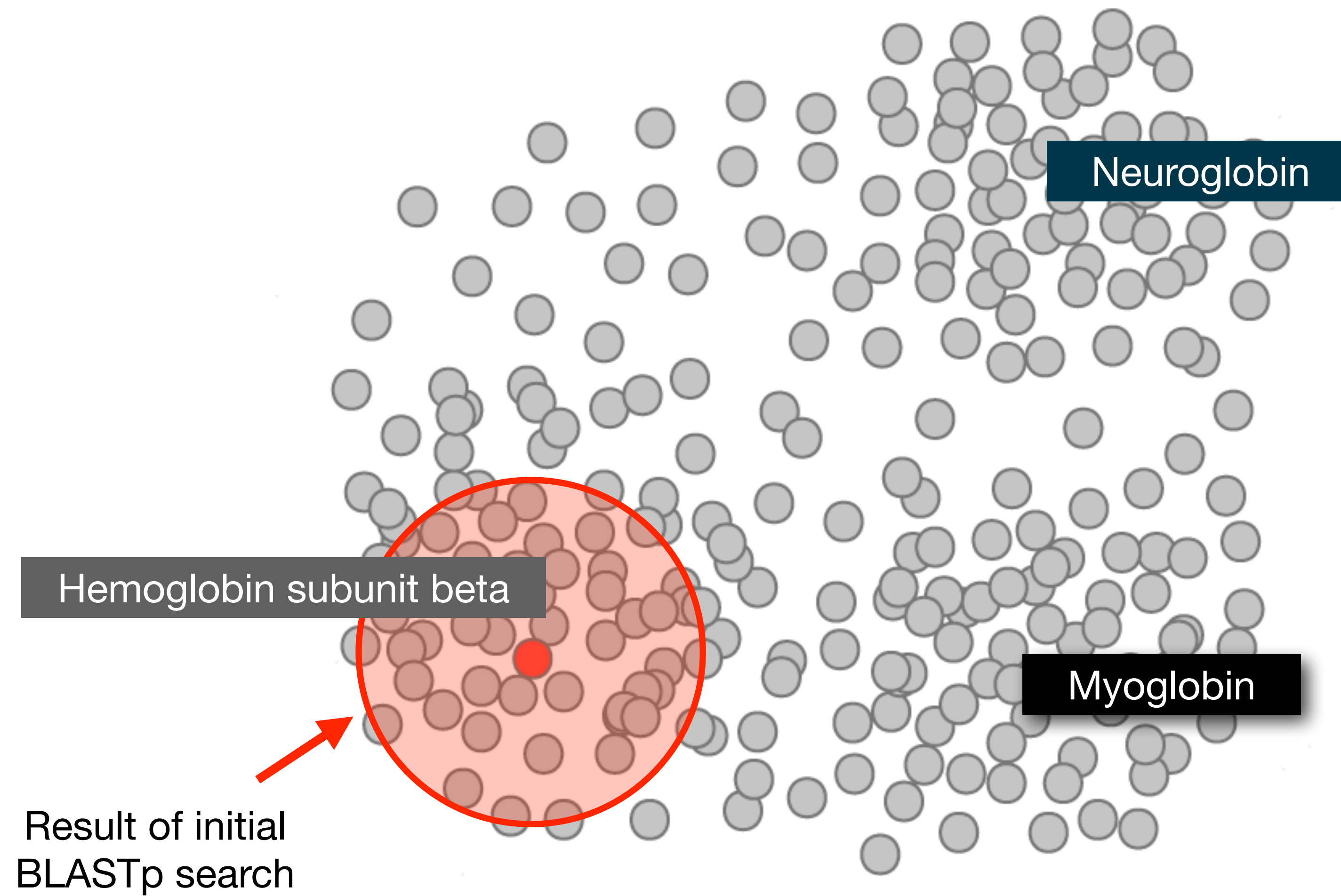
PSI-BLAST: Position-Specific Iterated BLAST

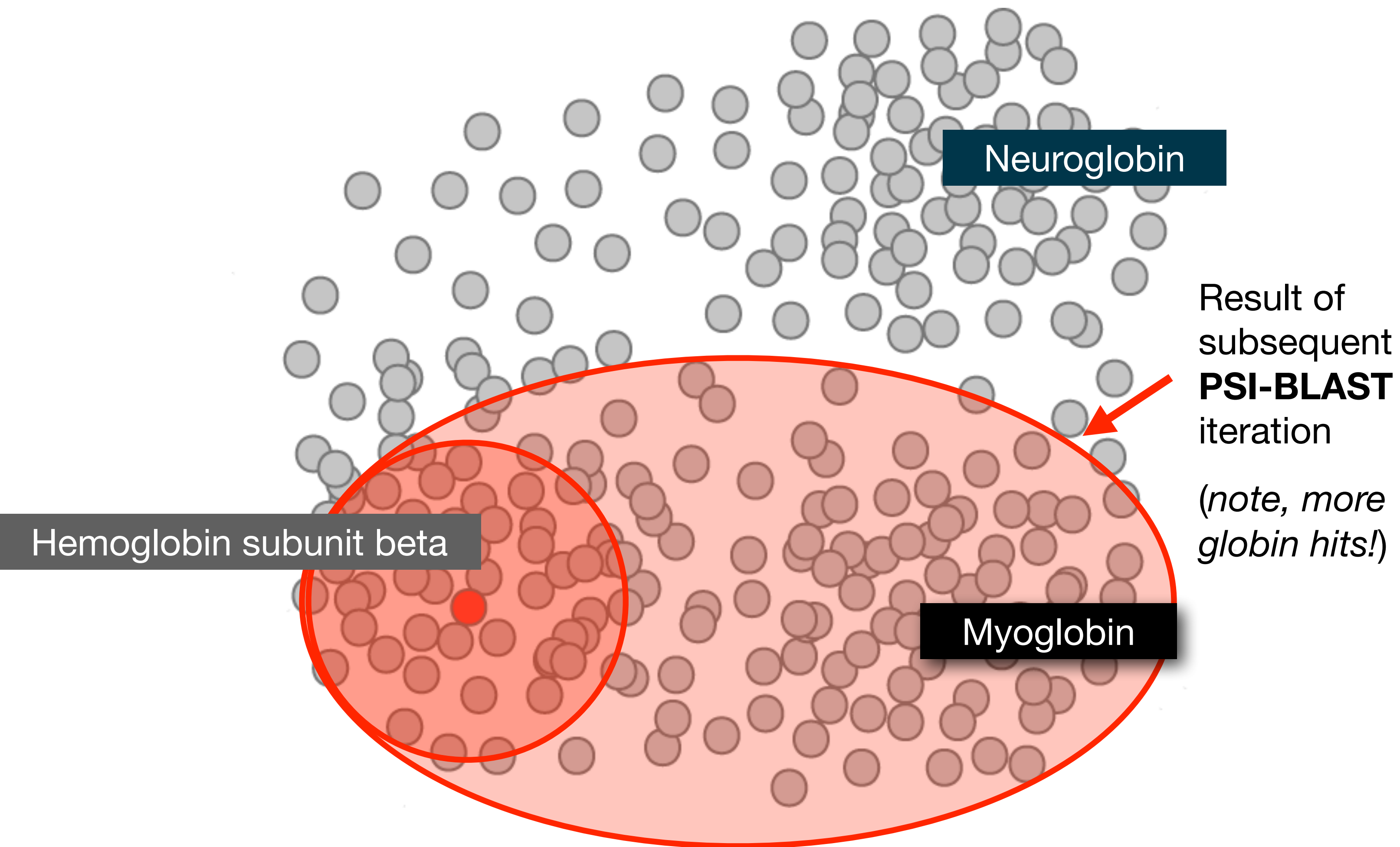
Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST

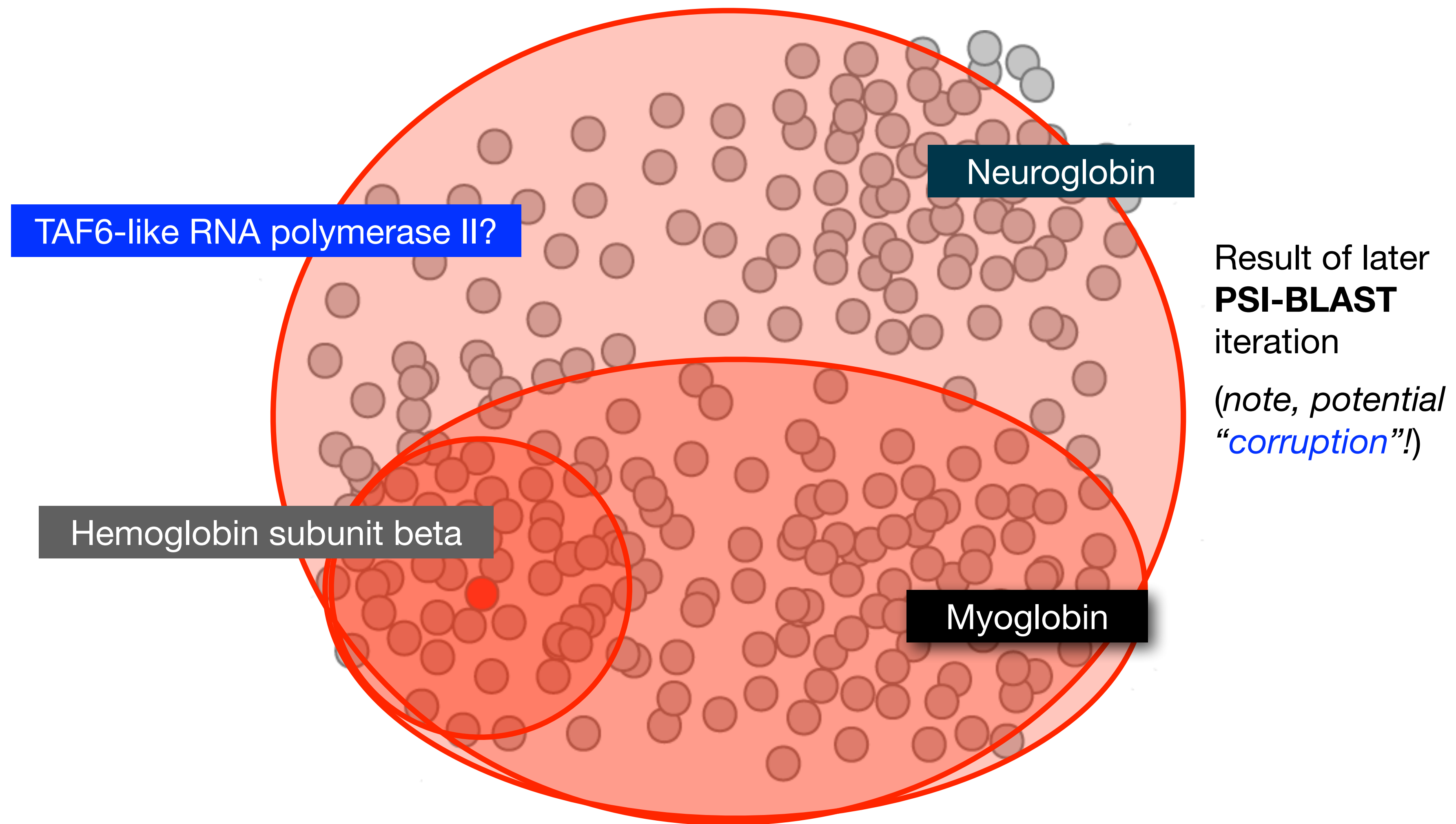


(see Altschul *et al.*, Nuc. Acids Res. (1997) 25:3389-3402)









Description	Max score	Total score	Query cover	E value	Ident	Accession
hemoglobin subunit beta [Homo sapiens]	301	301	100%	2e-106	100%	NP_000509.1
hemoglobin subunit delta [Homo sapiens]	284	284	100%	7e-100	93%	NP_000510.1
hemoglobin subunit epsilon [Homo sapiens]	240	240	100%	2e-82	76%	NP_005321.1
hemoglobin subunit gamma-2 [Homo sapiens]	235	235	100%	2e-80	73%	NP_000175.1
hemoglobin subunit gamma-1 [Homo sapiens]	232	232	100%	3e-79	73%	NP_000550.2
hemoglobin subunit alpha [Homo sapiens]	114	114	97%	7e-33	43%	NP_000508.1
hemoglobin subunit zeta [Homo sapiens]	100	100	97%	3e-27	36%	NP_005323.1

Description	Max score	Total score	Query cover	E value	Ident	Accession
hemoglobin subunit beta [Homo sapiens]	301	301	100%	2e-106	100%	NP_000509.1
hemoglobin subunit delta [Homo sapiens]	284	284	100%	7e-100	93%	NP_000510.1
hemoglobin subunit epsilon [Homo sapiens]	240	240	100%	2e-82	76%	NP_005321.1
hemoglobin subunit gamma-2 [Homo sapiens]	235	235	100%	2e-80	73%	NP_000175.1
hemoglobin subunit gamma-1 [Homo sapiens]	232	232	100%	3e-79	73%	NP_000550.2
hemoglobin subunit alpha [Homo sapiens]	114	114	97%	7e-33	43%	NP_000508.1
hemoglobin subunit zeta [Homo sapiens]	100	100	97%	3e-27	36%	NP_005323.1
myoglobin [Homo sapiens]	80.5	80.5	97%	2e-19	26%	NP_005359.1
neuroglobin [Homo sapiens]	54.7	54.7	92%	2e-09	23%	NP_067080.1

1

2

New relevant globins found only by PSI-BLAST

Description	Max score	Total score	Query cover	E value	Ident	Accession
hemoglobin subunit beta [Homo sapiens]	301	301	100%	2e-106	100%	NP_000509.1
hemoglobin subunit delta [Homo sapiens]	284	284	100%	7e-100	93%	NP_000510.1
hemoglobin subunit epsilon [Homo sapiens]	240	240	100%	2e-82	76%	NP_005321.1
hemoglobin subunit gamma-2 [Homo sapiens]	235	235	100%	2e-80	73%	NP_000175.1
hemoglobin subunit gamma-1 [Homo sapiens]	232	232	100%	3e-79	73%	NP_000550.2
hemoglobin subunit alpha [Homo sapiens]	114	114	97%	7e-33	43%	NP_000508.1
hemoglobin subunit zeta [Homo sapiens]	100	100	97%	3e-27	36%	NP_005323.1

1

myoglobin [Homo sapiens]	80.5	80.5	97%	2e-19	26%	NP_005359.1
neuroglobin [Homo sapiens]	54.7	54.7	92%	2e-09	23%	NP_067080.1

2

myoglobin [Homo sapiens]	159	159	97%	3e-50	26%	NP_005359.1
hemoglobin subunit alpha [Homo sapiens]	151	151	97%	3e-47	42%	NP_000508.1
hemoglobin subunit mu [Homo sapiens]	147	147	97%	6e-46	35%	NP_001003938.1
hemoglobin subunit theta-1 [Homo sapiens]	147	147	97%	2e-45	37%	NP_005322.1
neuroglobin [Homo sapiens]	134	134	92%	3e-40	23%	NP_067080.1
PREDICTED: cytoglobin isoform X2 [Homo sapiens]	115	115	66%	3e-33	25%	XP_016879605.1

3

PREDICTED: microtubule cross-linking factor 1 isoform X1 [Homo sapie	46.3	46.3	27%	7e-06	39%	XP_011523942.1
PREDICTED: microtubule cross-linking factor 1 isoform X4 [Homo sapie	46.3	46.3	27%	7e-06	39%	XP_005258156.1

?

Inclusion of irrelevant hits can lead to PSSM corruption

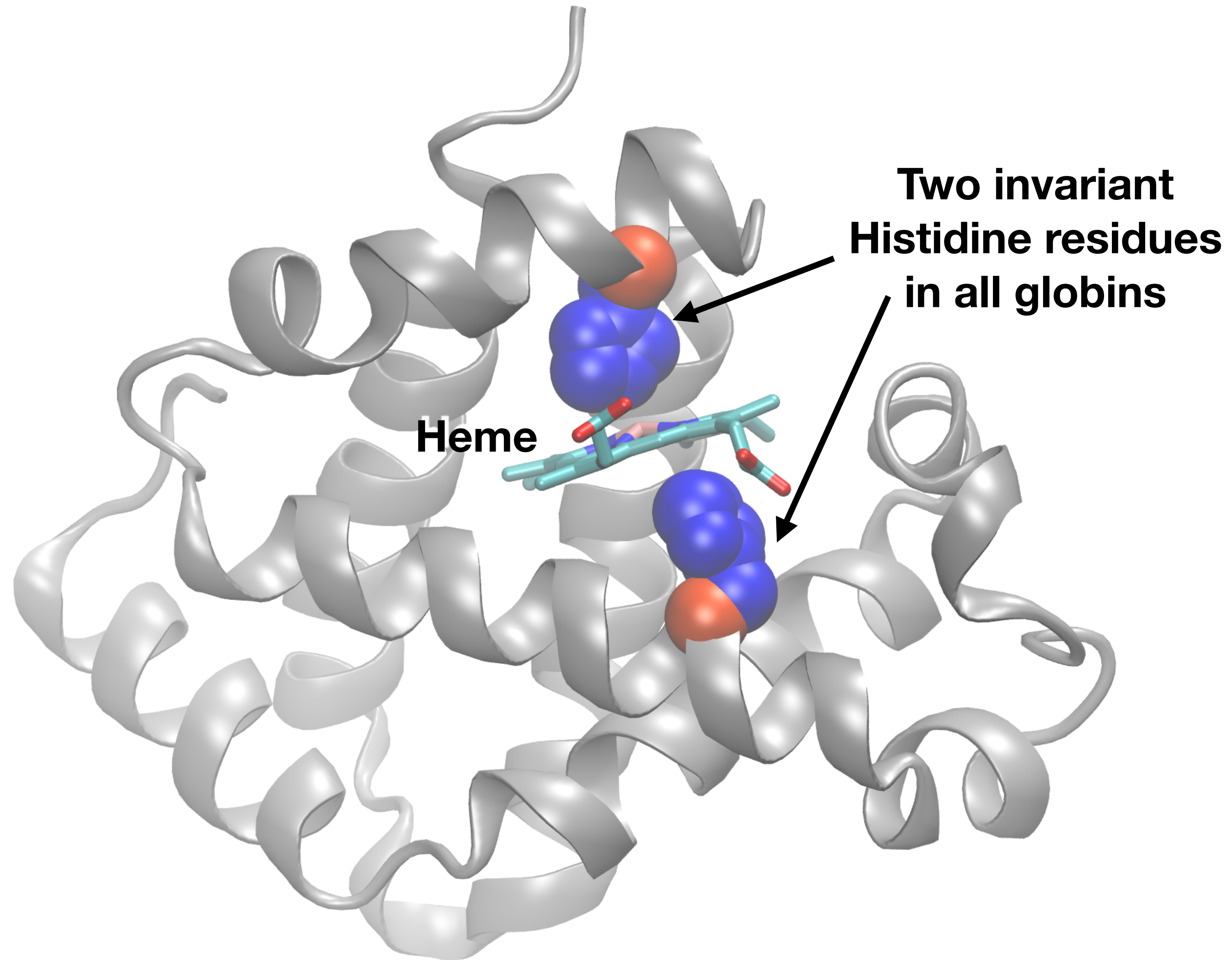
YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

1. Limits of using BLAST [~10 mins]
2. Using PSI-BLAST [~30 mins]
3. **Examining conservation patterns** [~20 mins]
— BREAK [15 mins]—
4. [Optional] Using HMMER [~10 mins]
5. Divergence of protein sequence and structure [~25 mins]

- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!

✓Query_73613	1	MVHLTPEEKSAVTALWGKV--NVDEVGGEALGRLLVVYPWTQRFFE-SFGDLSTPDAVM-GNPKVKAHGKKVLGAF	72
✓NP_000510.1	1	MVHLTPEEKTAVNALWGKV--NVDAVGGEALGRLLVVYPWTQRFFE-SFGDLSSPDAVM-GNPKVKAHGKKVLGAF	72
✓NP_000175.1	1	MGHFTTEEDKATITSLWGKV--NVEDAGGETLGRLLVVYPWTQRFFD-SFGNLSSASAIM-GNPKVKAHGKKVLTSL	72
✓NP_000509.1	1	MVHLTPEEKSAVTALWGKV--NVDEVGGEALGRLLVVYPWTQRFFE-SFGDLSTPDAVM-GNPKVKAHGKKVLGAF	72
✓NP_005321.1	1	MVHFTAEEKA AVTSLWSKM--NVEEAGGEALGRLLVVYPWTQRFFD-SFGNLSSPSAIL-GNPKVKAHGKKVLTSL	72
✓NP_000550.2	1	MGHFTTEEDKATITSLWGKV--NVEDAGGETLGRLLVVYPWTQRFFD-SFGNLSSASAIM-GNPKVKAHGKKVLTSL	72
✓NP_005323.1	1	-MSLTKTERTIIIVSMWAKISTQADTIGTETLERLFLSHPQTKTYFP-HF-----DLHpGSAQLRAHGSKVVA AV	67
✓NP_000508.1	1	-MVLSPADKTNVKA AWGKVG AHAGEYGA EALERMFLSFPTTKTYFP-HF-----DLShGSAQVKGHGKKVADAL	67
✓XP_005257062.1	1	[15] SEELSEAERKAVQAMWARLYANCEDVG VAILVRFFVNFPSAKQYFS-QFKHMEDPLEME-RSPQLRK HACRVMGAL	89
✓NP_001003938.1	1	--MLSAQERAQIAQVWDLIAGHEAQFGAELLRLFTVYPSTKVYFP-HL-----SACQ-DATQLLSHGQRM LA AV	66
✓NP_005322.1	1	-MALSAEDRALVRALWKKLGSNVGVYTTEALERTFLAFPATKTYFS-H-----LDLSpGSSQVRAHGQKVADAL	67
✓NP_599030.1	1	[15] SEELSEAERKAVQAMWARLYANCEDVG VAILVRFFVNFPSAKQYFS-QFKHMEDPLEME-RSPQLRK HACRVMGAL	89
✓XP_016879605.1	1	-----MEDPLEME-RSPQLRK HACRVMGAL	24
✓NP_001349775.1	1	-MGLSDGEWQLVLNVWGKVEADIPGHGQEV LIRLFKGHPETLEKFD-KFKHLKSEDEMK-ASEDLKKHGATVLTAL	73
✓NP_067080.1	1	---MERPEPELIRQSWRAVSRSPLEHGT VLFARLFALEPDLLPLFQyNCRQFSSPEDCL-SSPEFLDHIRKVMLVI	72
✓NP_001369741.1	1	-----MK-ASEDLKKHGATVLTAL	18
✓Query_73613	73	SDGLAHL DNLKGT---FATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH	147
✓NP_000510.1	73	SDGLAHL DNLKGT---FSQLSELHCDKLHVDPENFRLLGNVLVCVLAHNFNGKEFTPQMQAAYQKVVAGVANALAHKYH	147
✓NP_000175.1	73	GDAIKHLDDLKGT---FAQLSELHCDKLHVDPENFKLLGNVLVTVLAIHFGKEFTPEVQASWQKMVTGVASALSSRYH	147
✓NP_000509.1	73	SDGLAHL DNLKGT---FATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH	147
✓NP_005321.1	73	GDAIKNMDNLKPA---FAKLSELHCDKLHVDPENFKLLGNVMV IILATHFGKEFTPEVQA AWQKLVS AVAIALAHKYH	147
✓NP_000550.2	73	GDA TKHLDDLKGT---FAQLSELHCDKLHVDPENFKLLGNVLVTVLAIHFGKEFTPEVQASWQKMVTAVASALSSRYH	147
✓NP_005323.1	68	GDA VKSIDDIGGA---LSKLSELHAYILRVDPVNFKLLSHCLLVTLAARFPADFTA EAHAAWDKFLSVVSSVLTEKYR	142
✓NP_000508.1	68	TNAVAHVDDMPNA---LSALSDLHA HKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLT SKYR	142
✓XP_005257062.1	90	NTVVENLHDPDKVssvLALVGKAHALKHKVEPVYFKILSGVILEVVAEEFASDFPPETQRAWAKLRGLIYSHVTAAYK [35]	202
✓NP_001003938.1	67	GAAVQHVDNLRAA---LSPLADLHALVLRVDPANFP LLIQCFHVVLASHLQDEFTVQMQA AWDKFLTGVAVVLTEKYR	141
✓NP_005322.1	68	SLAVERLDDLPHA---LSALSHLHACQLRVDPASFQLLGHCLLVTLARHYPGDFSPALQASLDKFLSHVISALVSEYR	142
✓NP_599030.1	90	NTVVENLHDPDKVssvLALVGKAHALKHKVEPVYFKILSGVILEVVAEEFASDFPPETQRAWAKLRGLIYSHVTAAYK [23]	190
✓XP_016879605.1	25	NTVVENLHDPDKVssvLALVGKAHALKHKVEPVYFKILSGVILEVVAEEFASDFPPETQRAWAKLRGLIYSHVTAAYK [35]	137
✓NP_001349775.1	74	GGILKKKGHHEAE---IKPLAQSHATKHKIPVKYLEFISECIIQVLQSKHPGDFGADAQGAMNKALELFRKDMASNYK [6]	154
✓NP_067080.1	73	DAAVTNVEDLSSLeeyLASLGRKHRA-VGVKLSSFSTVGESLLYMLEKCLGPAFTPATRAAWSQLYGAVVQAMSRGWD [2]	151
✓NP_001369741.1	19	GGILKKKGHHEAE---IKPLAQSHATKHKIPVKYLEFISECIIQVLQSKHPGDFGADAQGAMNKALELFRKDMASNYK [6]	99



YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

1. Limits of using BLAST [~10 mins]

2. Using PSI-BLAST [~30 mins]

3. Examining conservation patterns [~20 mins]

— BREAK [15 mins]—

4. **[Optional] Using HMMER** [~10 mins]

5. Divergence of protein sequence and structure [~25 mins]

- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!

Problems with PSSMs: Positional dependencies

Do not capture positional dependencies

WEIRD
WEIRD
WEIQH
WEIRD
WEIQH

D					0.6
E		I			
H					0.4
I			I		
Q				0.4	
R				0.6	
W	I				

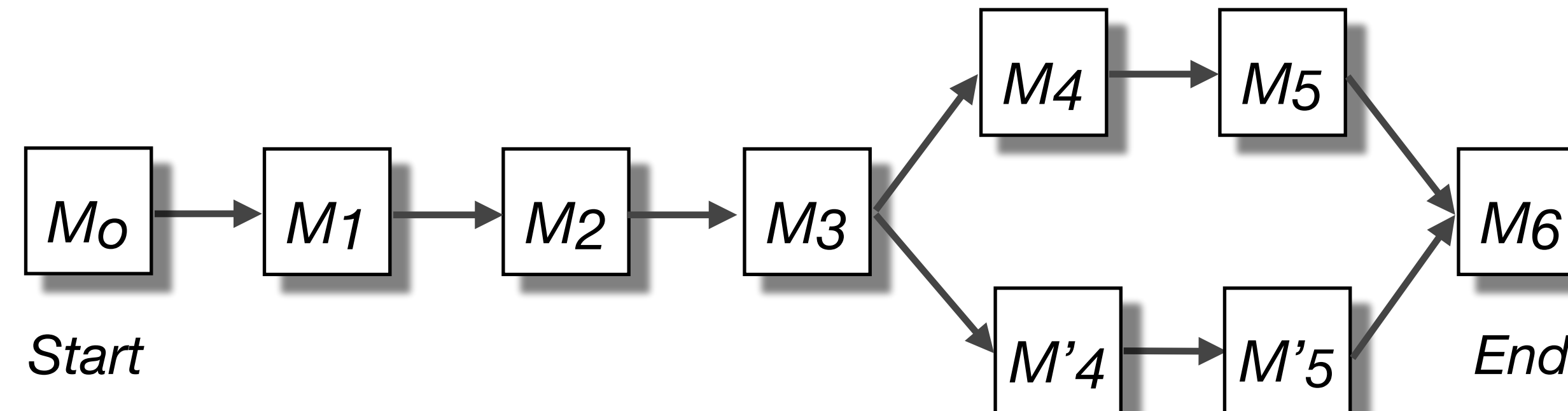
Note: We never see **QD** or **RH**, we only see **RD** and **QH**.
However, $P(RH)=0.24$, $P(QD)=0.24$, while $P(QH)=0.16$

Markov chains: Positional dependencies



The connectivity or **topology** of a Markov chain can easily be designed to capture dependencies and variable length motifs.

WEIRD
WEIRD
WEIQH
WEIRD
WEIQH

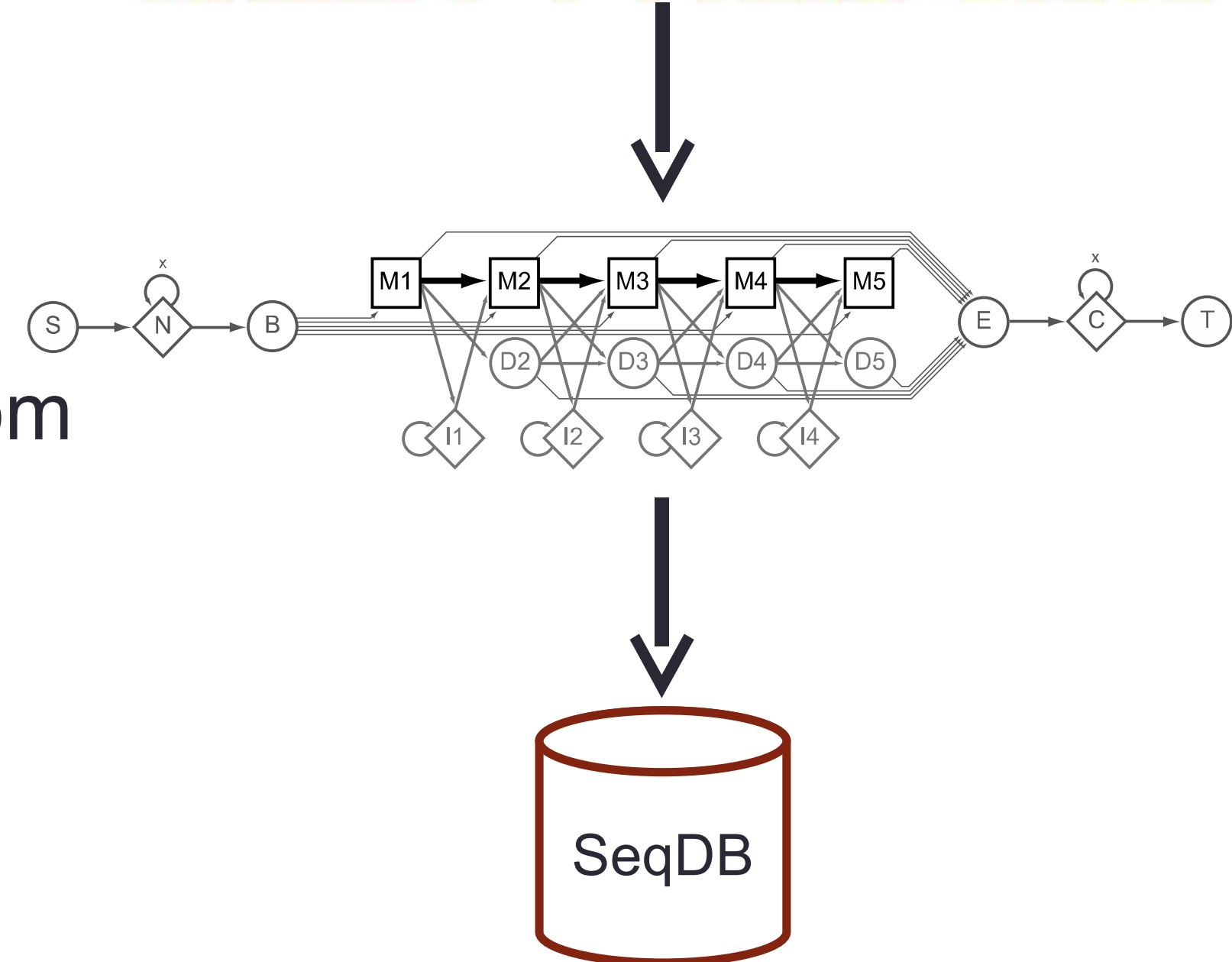


Recall that a PSSM for this motif would give the sequences **WEIRD** and **WEIRH** equally good scores even though the **RH** and **QR** combinations were not observed

Use of HMMER

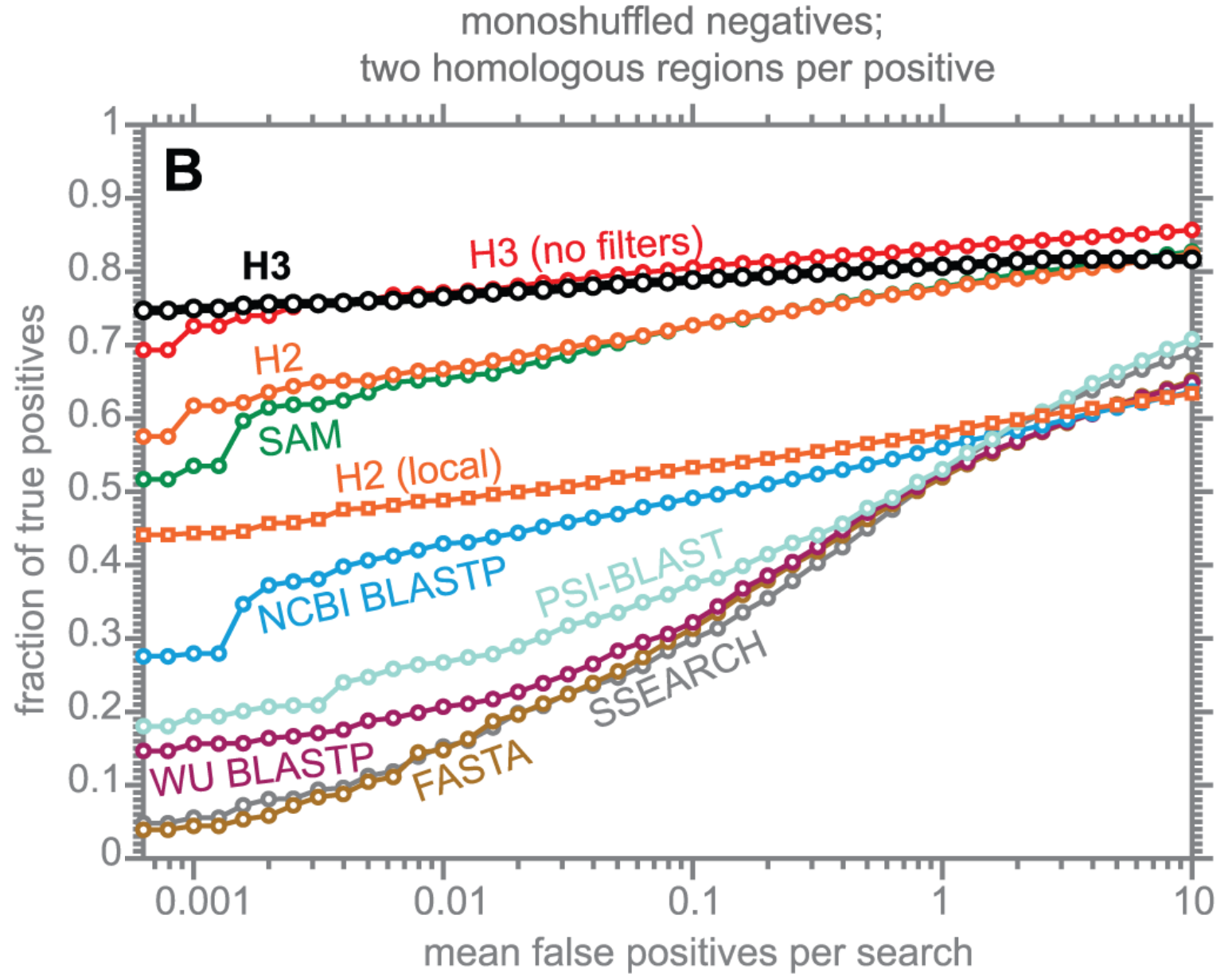
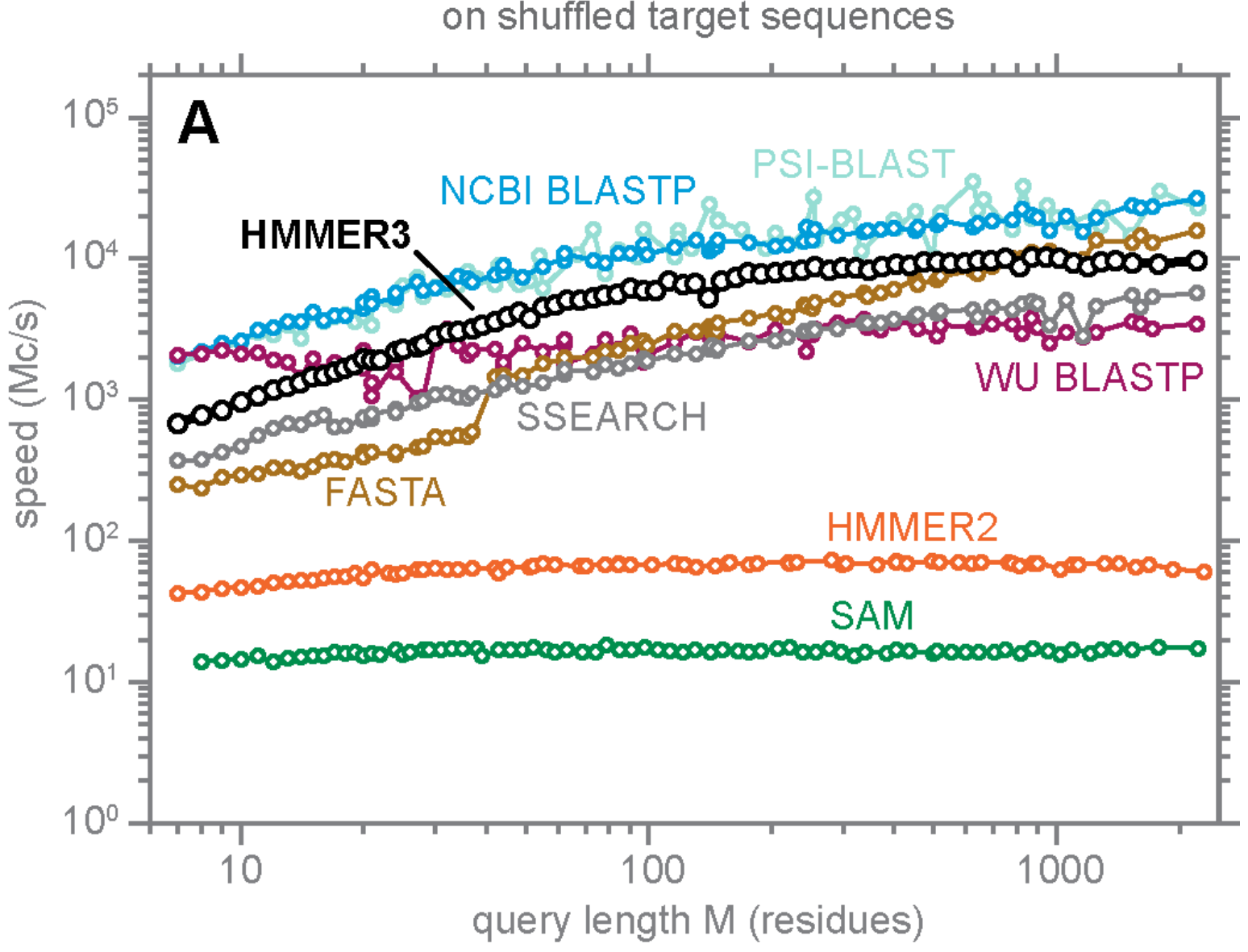
- Widely used by protein family databases
 - Use 'seed' alignments
- Until 2010
 - Computationally expensive
 - Restricted to HMMs constructed from multiple sequence alignments
- Command line application

```
KIIITGEPGVGKTTLVKKIVERL---GKRAIGFWTEEVDPETKKRTGFRIITTE  
KILITGRPGVGKTTLIKLSRL-----QNAGGFYTEEMR--EGEKRIQFKIITLD  
RFFVSGMPGVGKTTLAKRIADEVRREGFKVGGIITEEIR--EGGKRTGFRVIALD  
RIFITGMPGVGKTTLALKIAEKLKELGYKVGGIITKEIR--DGGKRVGFKIITLD  
RFFVSGMPGVGKTTLAKRIADEIKREGFKVGGIITQEIR--SGARRSGFRVIALD  
HVFLTGPFGVGKTTLIQKAIEVLQSSGLPVDGFYTQEVN--QEGKRIQFDVVTLS  
HVFLTGVPGVGKTTLVKKVCDAL--SGLSVSGFYTEEVN--EHGRRVGFDVVTVS  
HVFLTGSPGVGKTTLIQKAITVLQSSGLPVDGFYTQEVN--QGGKRIQFDVVTLS  
HVFLTGPFGVGKTTLIHKASEVLKSSGVPVDGFYTEEVN--QGGRRIGFDVVTLS
```



HMMER vs BLAST

	HMMER	BLAST
Program	<i>PHMMER</i>	<i>BLASTP</i>
Query	Single sequence	
Target Database	Sequence database	
Program	<i>HMMSCAN</i>	<i>RPSBLAST</i>
Query	Single sequence	
Target Database	Profile HMM database, e.g. Pfam	PSSM database, e.g. CDD
Program	<i>HMMSEARCH</i>	<i>PSI-BLAST</i>
Query	Profile HMM	PSSM
Target Database	Sequence database	
Program	<i>JACKHMMER</i>	<i>PSI-BLAST</i>
Query	Single sequence	
Target Database	Sequence database	



Modified from: S. R. Eddy
PLoS Comp. Biol., 7:e1002195, 2011.



Fast Web Searches

- Parallelized searches across compute farm
 - Average query returns ~1 sec
- Range of sequence databases
 - Large Comprehensive
 - Curated / Structure
 - Metagenomics
 - Representative Proteomes
- Family Annotations
 - Pfam
- Batch and RESTful API
 - Automatic and Human interface





HMMER

Biosequence analysis using profile hidden Markov Models

[Home](#)[Search](#)[Results](#)[Software](#)[Help](#)[About](#)[Contact](#)[phmmer](#)[hmmsearch](#)[hmmsearch](#)[jackhmmmer](#)

protein sequence vs protein sequence database

[Paste a Sequence](#) | [Upload a File](#) | [Accession Search](#)

Paste in your sequence or use the [example](#) ?

```
>NP_000509.1 hemoglobin subunit beta [Homo sapiens]  
MVHLTPEEKSAVTALWGKVNVDVEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPKVKAHGKKVLG  
AFSDGLAHLNLIKGTATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVAN  
ALAHKYH
```

[Submit](#)[Reset](#)

▼ Sequence Database ?

Frequently used databases:

[Reference Proteomes](#)[UniProtKB](#)[SwissProt](#)[PDB](#)[Ensembl](#)

Current database selection:














































































































SwissProt ▼

▼ Restrict by Taxonomy ?

☒ Taxon search

☐ Pre-defined representatives

Organism:

Significant Query Matches (12) in <i>swissprot</i> (v.2018_11)						Customise	Customise
	Target	Description	Species	 Cross-references	E-value		
>	HBB_HUMAN 	Hemoglobin subunit beta	Homo sapiens 	      	6.8e-99		
>	HBD_HUMAN 	Hemoglobin subunit delta	Homo sapiens 	      	1.6e-91		
>	HBE_HUMAN 	Hemoglobin subunit epsilon	Homo sapiens 	      	1.5e-74		
>	HBG2_HUMAN 	Hemoglobin subunit gamma-2	Homo sapiens 	      	8.8e-73		
>	HBG1_HUMAN 	Hemoglobin subunit gamma-1	Homo sapiens 	      	6.2e-72		
>	HBA_HUMAN 	Hemoglobin subunit alpha	Homo sapiens 	      	3.8e-29		
>	HBAZ_HUMAN 	Hemoglobin subunit zeta	Homo sapiens 	      	4.5e-23		
>	HBAT_HUMAN 	Hemoglobin subunit theta-1	Homo sapiens 	      	5.2e-22		
>	HBM_HUMAN 	Hemoglobin subunit mu	Homo sapiens 	      	3.4e-19		
>	CYGB_HUMAN 	Cytoglobin	Homo sapiens 	      	3.1e-14		
>	MYG_HUMAN 	Myoglobin	Homo sapiens 	      	2.3e-06		
>	NGB_HUMAN 	Neuroglobin	Homo sapiens 	      	0.0017		
(show all) alignments						Your search took: 0.06 secs	
						showing rows 1 - 12 of 12	

PFAM: Protein Family Database of Profile HMMs

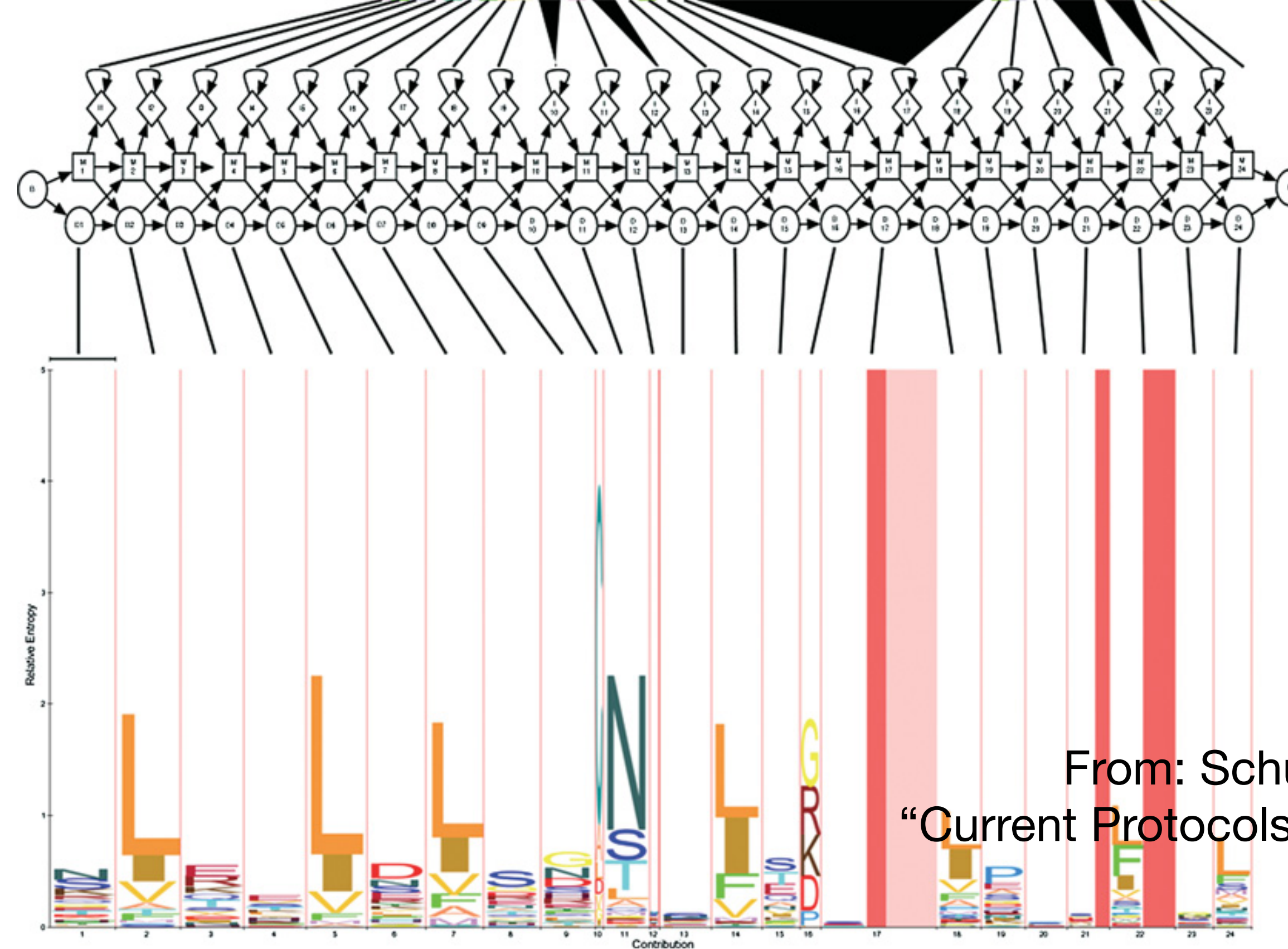
Comprehensive compilation of both multiple sequence alignments and profile HMMs of protein families.

<http://pfam.sanger.ac.uk/>

PFAM consists of two databases:

- **Pfam-A** is a manually curated collection of protein families in the form of multiple sequence alignments and profile HMMs. HMMER software is used to perform searches.
- **Pfam-B** contains additional protein sequences that are automatically aligned. Pfam-B serves as a useful supplement that makes the database more comprehensive.
- Pfam-A also contains higher-level groupings of related families, known as **clans**

Q9ARB2_LINUS/823-844 MLEYLDIGRA..P.RIV.H.....LDG...LENL
 Q9M8N0_ARATH/320-341 RLTFNLNLSFC..S.KLT.G.....LAF...FSII
 FLJI_HUMAN/318-339 NLEEFMAAN..N..NLE.L.....VPES..LCRC
 Q9VN74_DROME/90-112 ALHSLVIENC...TIV.H.....INDAA.FNQE
 Q8L8I7_PNTA/792-814 NLQTIQMYRX..E.SLQ.V.....LPDS..FGNL
 Q9FHL8_ARATH/301-324 NLWSLNLSR..N..LFSDP.....LPVVG.ARGF
 SLIK6_MOUSE/65-87 RPFHLSLLN..N..GLT.M.....LHTND.FSGL
 Q8NIJ8_EMEN/978-1000 TLTSLNIIAS..A..KLV.Q.....FRDTL.FDSL
 Q9LUQ2_ARATH/92-113 AMKSLDVSF..N..SIS.E.....LPEQ..IGSA
 Q9FH93_ARATH/169-188 RLTSNLNDF..N..RFNGT.....LPS....LN
 Q898G0_CLOTE/268-288 YLERINLDK..N..KI.KN.....IEE...LEAN
 Q8H6V2_MAIZE/678-699 NLRILSIVDC..V.SLQ.K.....LPP...SDSF
 Q9AR40_LINUS/692-713 DLKVLNINQ..T..EIT.T.....LKGE..VESL
 Q9LE82_ARATH/350-377 HLTEIYMSY..L..NLEDEGT.....EALSEAL.LKSA
 Q9H5N5_HUMAN/255-278 HLQVLDLHQC...SLT.AD.....DVMSL...TQVI
 Q8L4C7_ARATH/185-207 KLEYLDIWG..S..NVT.N.....QGAVS..ILKF
 Q9VSA4_DROME/1115-1136 QLKALRLQC..N..AI.GSH.....GLEAL..LCGQ
 TLR1_MOUSE/376-398 RLKTLNLSQK..N..QL.KN.....LENII.LTSA
 Q9TXJ6_LEMA/445-465 GLRDIDLSH..T..KVH.N.....IDA...LQAS
 FXL13_MOUSE/409-448 KLIYLDLSGC..T.QVL.VEKCPRISVVLIQSPHISDSA.FKAL
 Q9TXJ6_LEMA/927-948 ALTVVNANSC..V.NLT.S.....IEA...LESA
 Q9M4X9_CHLRE/1417-1444 LLAVLHLHD..NP.RLA.ADG.....VAGLAAA..LPGL
 Q945S6_LYCPM/656-677 NLRHLDVSN..T..RRL.K.....MPLH..LSRL



From: Schuster-Bockler *et al.*
 “Current Protocols in Bioinformatics”
 Supplement 18.

YOUR TURN!

- There are **four required** and **one optional** hands-on sections including:

1. Limits of using BLAST [~10 mins]

2. Using PSI-BLAST [~30 mins]

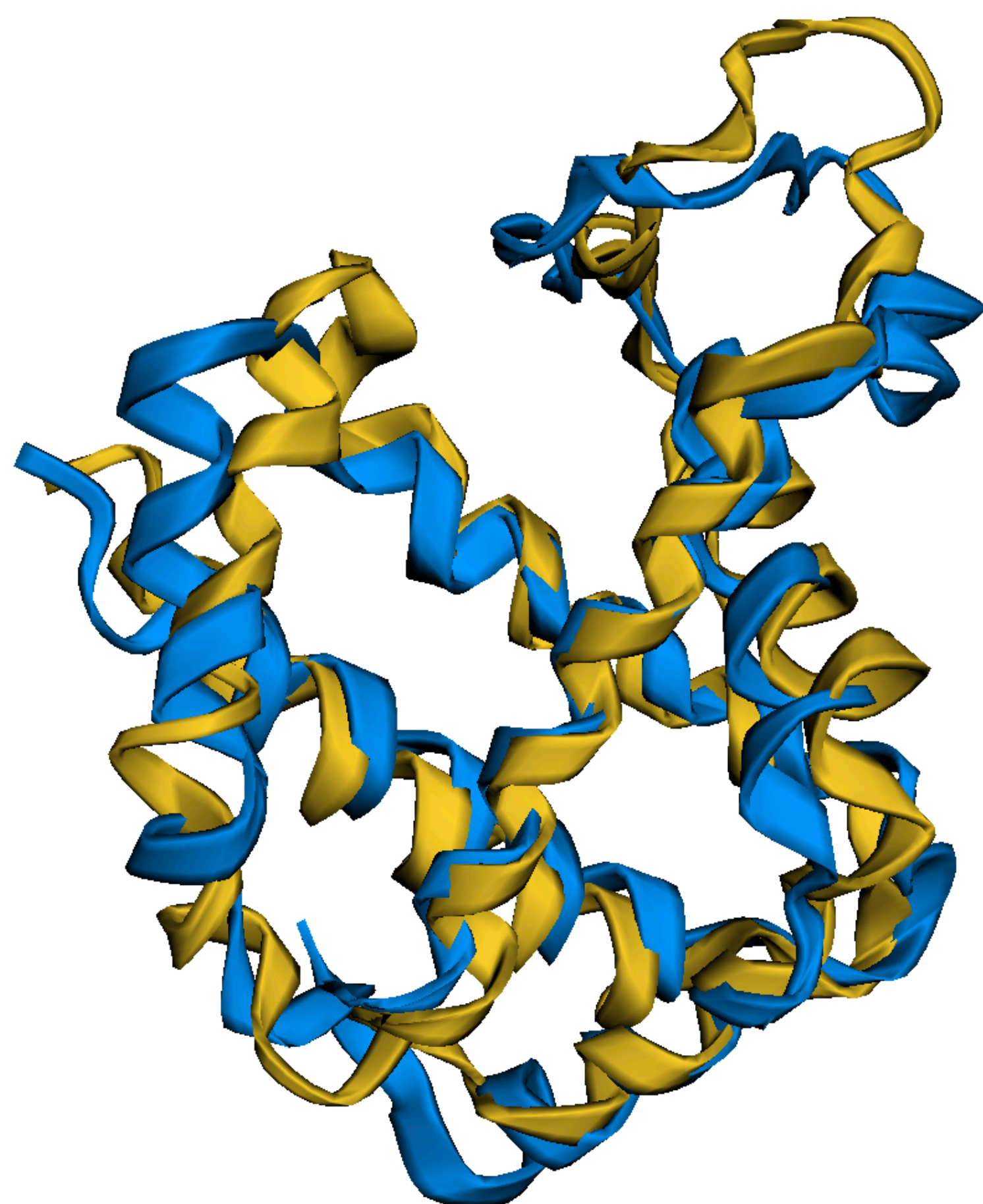
3. Examining conservation patterns [~20 mins]

— BREAK [15 mins]—

4. [Optional] Using HMMER [~10 mins]

5. **Divergence of protein sequence and structure** [~25 mins]

- ▶ Please do answer the last review question (**Q20**).
- ▶ We encourage discussion at your **Table** and on **Piazza**!



ALIGNMENT

CONTACT MAP

Align 2hbsB.pdb 146 with 4mpmB.pdb 148

```
Twists 0 ini-len 136 ini-rmsd 3.05 opt-equ 143 opt-rmsd 2.65 chain-rmsd 3.05
```

Score 318.72 align-len 150 gaps 7 (4.67%)

P-value 3.26e-14 Afp-num 14073 Identity 20.67% Similarity 40.00%

```
Block 0 afp 17 score 318.72 rmsd 3.05 gap 9 (0.06%)
```

[illegible]

Summary

- **Find a gene project:** You can start working on this now. Submit your responses to Q1-Q4 to get feedback.
- **PSI-BLAST algorithm:** Application of iterative position specific scoring matrices (PSSMs) to improve BLAST sensitivity
- **Hidden Markov models (HMMs):** More versatile probabilistic model for detection of remote similarities
- **Structure comparisons as gold standards:** Structure is more conserved than sequence

Homework: DataCamp!

Install **R** and **RStudio** (see website)

Complete the **Introduction to R** course on **DataCamp**
(Check Piazza for your DataCamp invite and sign up with your
UCSD email (i.e. first part of your email address) please.

Let me know **NOW** if you don't have access to DataCamp!