

Structural Bioinformatics II

Class 10

Barry Grant

UC San Diego

<http://thegrantlab.org>

Today's Menu

- Overview of structural bioinformatics
 - Motivations, goals and challenges
- Representing, interpreting & modeling protein structure
 - Visualizing & interpreting protein structures
 - Analyzing protein structures
 - Modeling protein structure

Finish last days Lab 09

bioboot.github.io

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Class 9

Code

1. Introduction to the RCSB Protein Data Bank (PDB)
2. Visualizing the HIV-1 protease structure
3. Introduction to Bio3D in R
4. Comparative structure analysis of Adenylate Kinase
5. Optional further visualization
6. Normal mode analysis [optional]

Class 9

Structural Bioinformatics (Pt. 1)

Barry Grant <<http://thegrantlab.org/teaching/>>
2022-10-25 (17:20:23 on Tue, Oct 25)

1: Introduction to the RCSB Protein Data Bank (PDB)

The PDB archive is the major repository of information about the 3D structures of large biological molecules, including proteins and nucleic acids. Understanding the shape of these molecules helps to understand how they work. This knowledge can be used to help deduce a structure's role in human health and disease, and in drug development. The structures in the PDB range from tiny proteins and bits of DNA or RNA to complex molecular machines like the ribosome composed of many chains of protein and RNA.

In the first section of this lab we will interact with the main US based PDB website (note there are also sites in Europe and Japan).

Visit: <http://www.rcsb.org/> and answer the following questions

NOTE: The "Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type" on the PDB home page should allow you to determine most of these answers.

PDB statistics

bioboot.github.io

Schedule - BIMM 143

Class 11

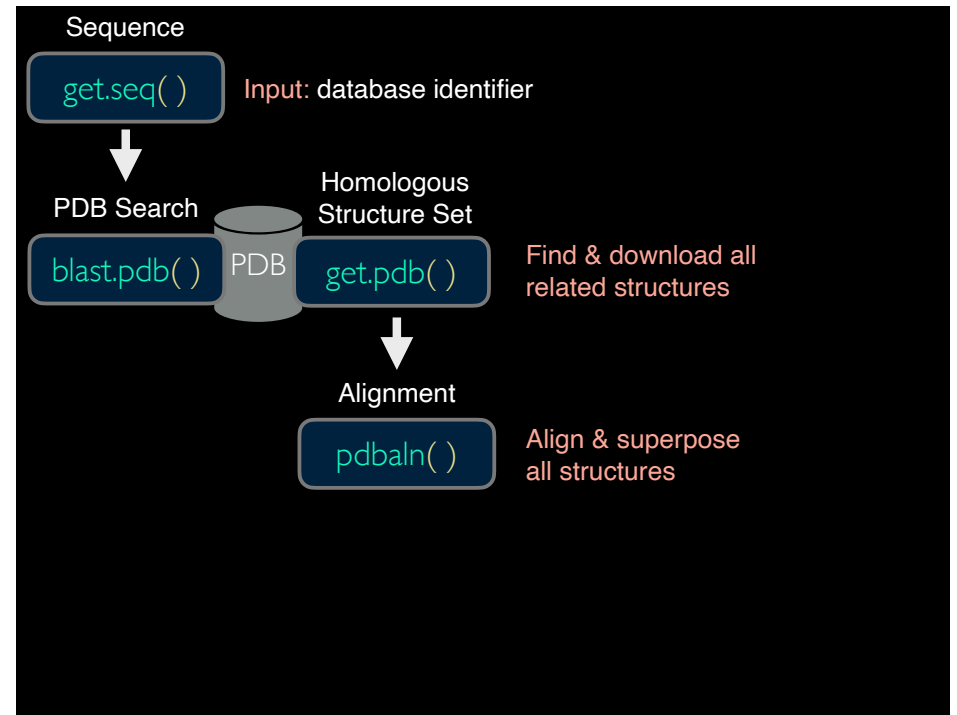
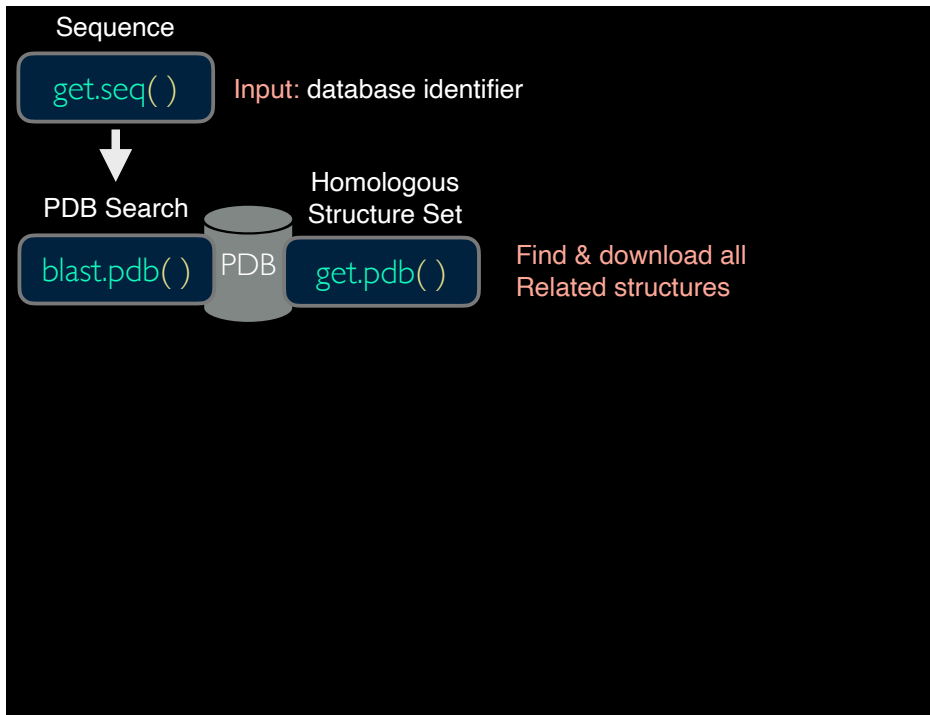
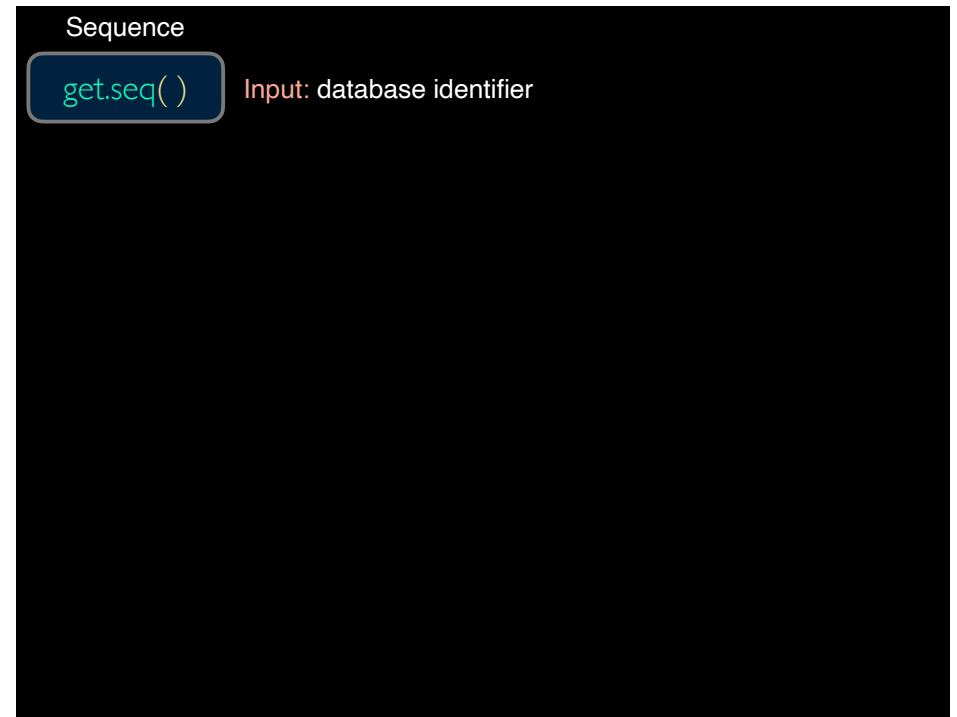
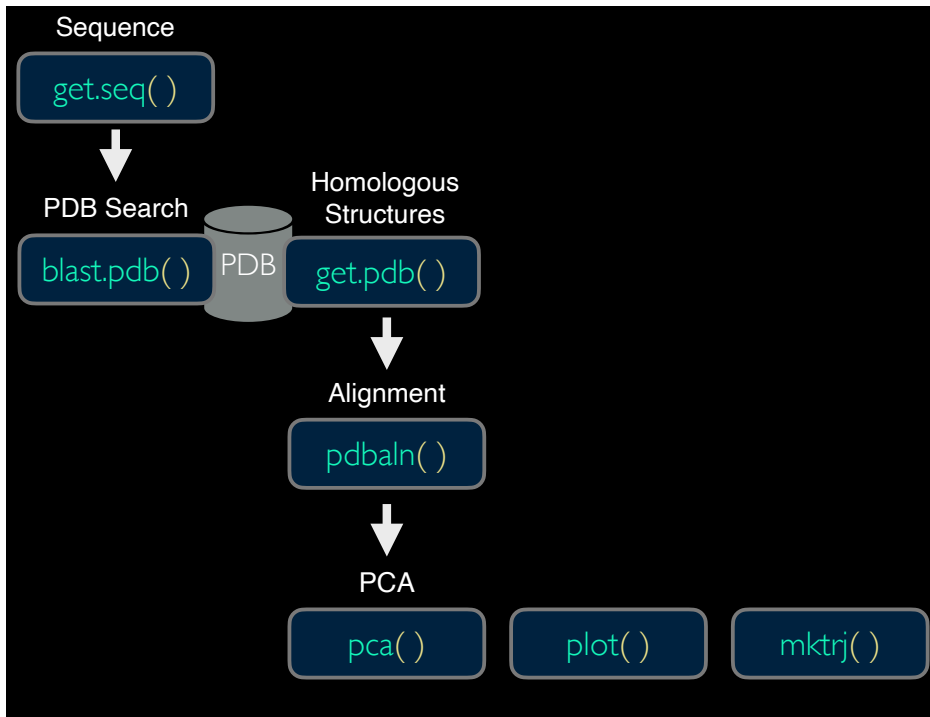
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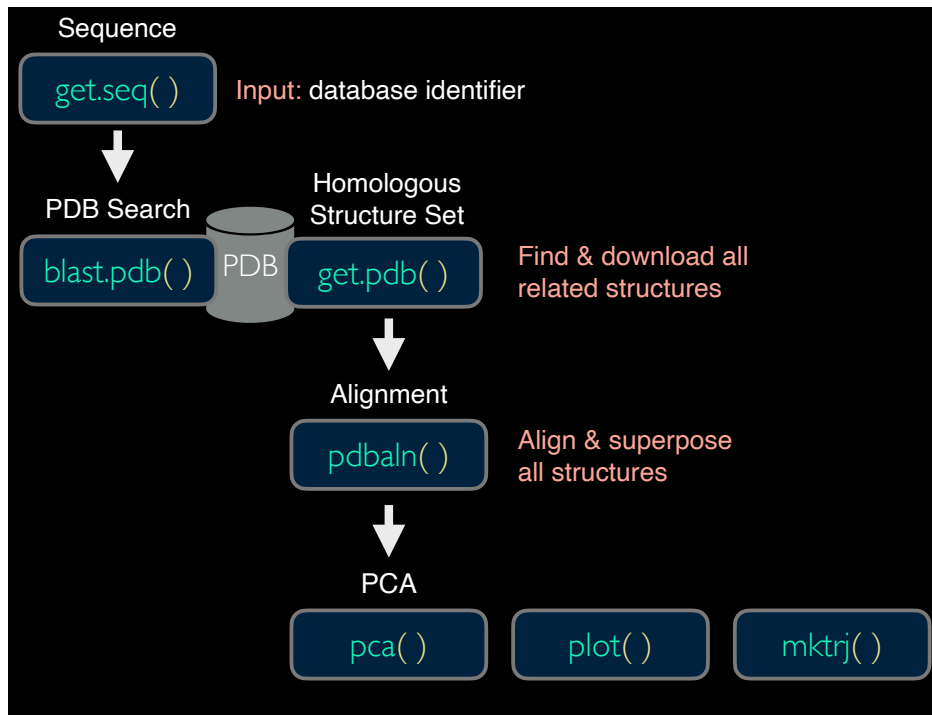
4. Comparative structure analysis of Adenylate Kinase

The goal of this section is to perform **principal component analysis (PCA)** on the complete collection of Adenylate kinase structures in the protein data-bank (PDB).

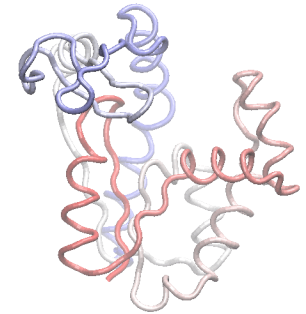
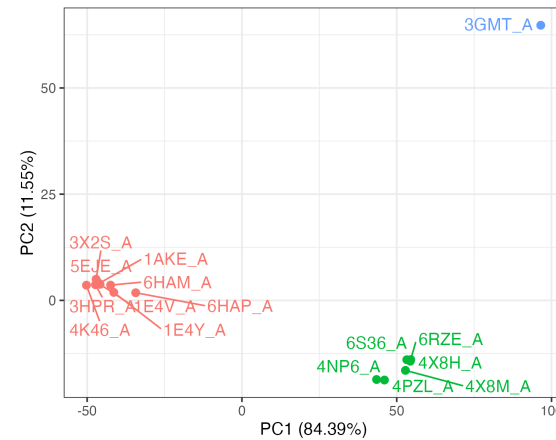
Adenylate kinase (often called simply Adk) is a ubiquitous enzyme that functions to maintain the equilibrium between cytoplasmic nucleotides essential for many cellular processes. Adk operates by catalyzing the reversible transfer of a phosphoryl group from ATP to AMP. This reaction requires a rate limiting conformational transition (i.e. change in shape). Here we analyze all currently available Adk structures in the PDB to reveal detailed features and mechanistic principles of these essential shape changing transitions.

1. Introduction to the RCSB Protein Data Bank (PDB)
2. Visualizing the HIV-1 protease structure
3. Introduction to Bio3D in R
4. Comparative structure analysis of Adenylate Kinase
 - Overview
 - Setup
 - Search and retrieve ADK structures
 - Align and superpose structures
 - Optional: Viewing our superposed structures
 - Annotate collected PDB structures [Optional]
 - Principal component analysis
5. Optional further visualization
6. Normal mode analysis





PCA Results



Today's Menu

Overview of structural bioinformatics

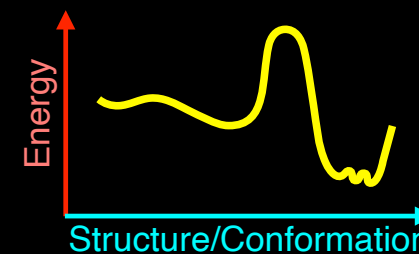
- Motivations, goals and challenges

Representing, interpreting & modeling protein structure

- Visualizing & interpreting protein structures
- Analyzing protein structures
- Modeling protein structure
 - Physics based approaches
 - Knowledge based approaches
 - Structure prediction and drug discovery

Key concept:

Potential functions describe a systems **energy** as a function of its **structure**



Two main approaches:

- (1). Physics-Based
- (2). Knowledge-Based

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For **physics** based potentials
energy terms come from physical theory

$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

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Sum of **bonded** and **non-bonded**
atom-type and position based terms

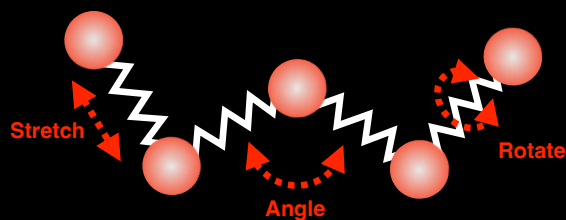
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E_{bonded} is itself a sum of three terms:

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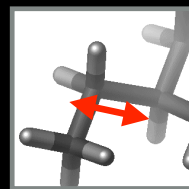
$$E_{\text{bond.stretch}} + E_{\text{bond.angle}} + E_{\text{bond.rotate}}$$



$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

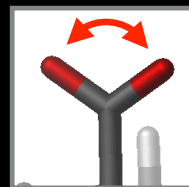
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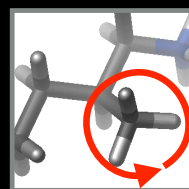
Bond Stretch

$$E_{\text{bond.stretch}}$$



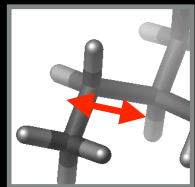
Bond Angle

$$E_{\text{bond.angle}}$$



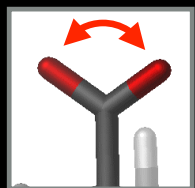
Bond Rotate

$$E_{\text{bond.rotate}}$$



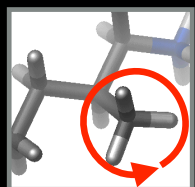
Bond Stretch

$$\sum_{\text{bonds}} K_i^{bs} (b_i - b_o)$$



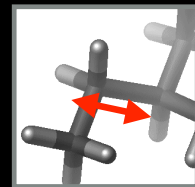
Bond Angle

$$\sum_{\text{angles}} K_i^{ba} (\theta_i - \theta_o)$$



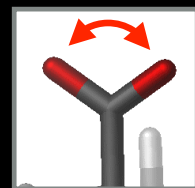
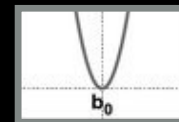
Bond Rotate

$$\sum_{\text{dihedrals}} K_i^{br} [1 - \cos(n_i \phi_i - \phi_o)]$$



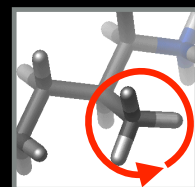
Bond Stretch

$$\sum_{\text{bonds}} K_i^{bs} (b_i - b_o)$$



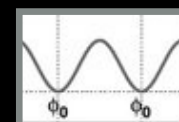
Bond Angle

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Bond Rotate

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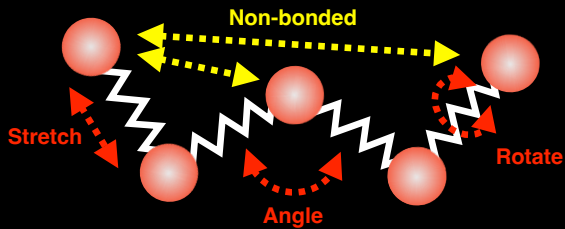
$$V(R) = E_{\text{bonded}} + E_{\text{non.bonded}}$$

$E_{\text{non.bonded}}$ is a sum of two terms:

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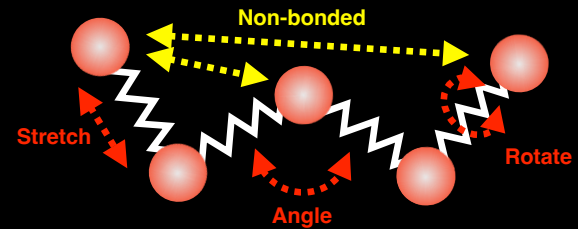
$$E_{\text{van.der.Waals}} + E_{\text{electrostatic}}$$



$$V(R) = E_{bonded} + E_{non.bonded}$$

$E_{non.bonded}$ is a sum of two terms:

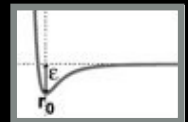
$$E_{van.der.Waals} + E_{electrostatic}$$



$$E_{electrostatic} = \sum_{pairs.i,j} \frac{q_i q_j}{\epsilon r_{ij}^2}$$



$$E_{van.der.Waals} = \sum_{pairs.i,j} \left[\epsilon_{ij} \left(\frac{r_{o,ij}}{r_{ij}} \right)^{12} - 2\epsilon_{ij} \left(\frac{r_{o,ij}}{r_{ij}} \right)^6 \right]$$



Total potential energy

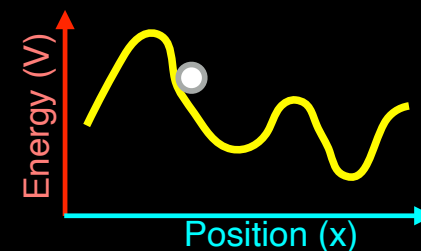
The potential energy can be given as a sum of terms for: Bond stretching, Bond angles, Bond rotations, van der Waals and Electrostatic interactions between atom pairs

$$V(R) = \left. \begin{array}{l} E_{bond.stretch} \\ + E_{bond.angle} \\ + E_{bond.rotate} \end{array} \right\} E_{bonded}$$

$$+ \left. \begin{array}{l} E_{van.der.Waals} \\ + E_{electrostatic} \end{array} \right\} E_{non.bonded}$$

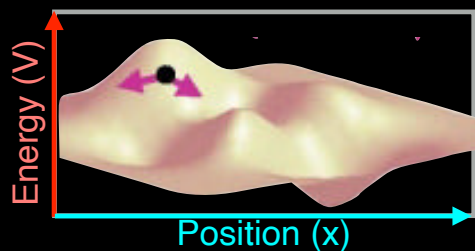
Potential energy surface

Now we can calculate the **potential energy surface** that fully describes the energy of a molecular system as a function of its geometry



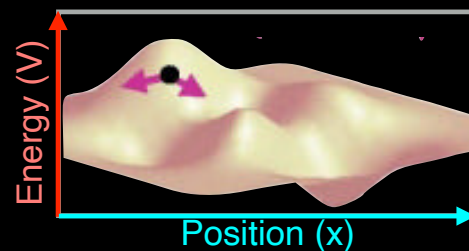
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Key concept:

Now we can calculate the **potential energy surface** that fully describes the energy of a molecular system as a function of its geometry



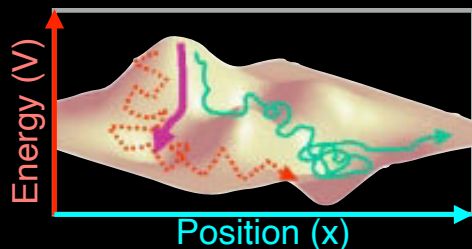
- The **forces** are the gradients of the energy

$$F(x) = -dV/dx$$

Moving Over The Energy Surface

- Energy Minimization** drops into local minimum
- Molecular Dynamics** uses thermal energy to move smoothly over surface
- Monte Carlo Moves** are random. Accept with probability:

$$\exp(-\Delta V/dx)$$



PHYSICS-ORIENTED APPROACHES

Weaknesses

- Fully physical detail becomes computationally intractable
- Approximations are unavoidable (Quantum effects approximated classically, water may be treated crudely)
- Parameterization still required

Strengths

- Interpretable, provides guides to design
- Broadly applicable, in principle at least
- Clear pathways to improving accuracy

Status

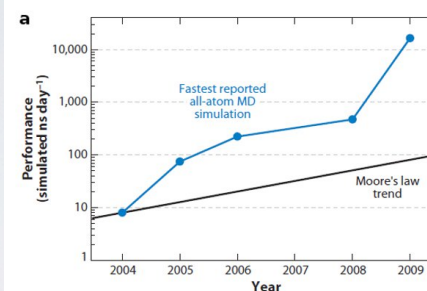
- Useful, widely adopted but far from perfect
- Multiple groups working on fewer, better approx
 - Force fields, quantum entropy, water effects
- Moore's law: hardware improving

HOW COMPUTERS HAVE CHANGED

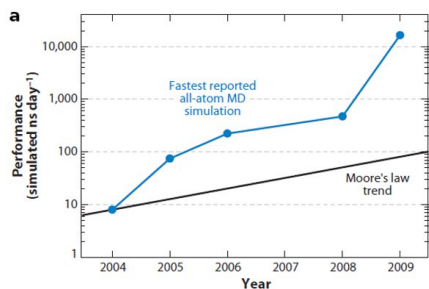
DATE	COST	SPEED	MEMORY	SIZE
1967	\$40M	0.1 MHz	1 MB	WALL
2013	\$4,000	1 GHz	10 GB	LAPTOP
CHANGE	10,000	10,000	10,000	10,000

If cars were like computers then a new Volvo would cost \$3, would have a top speed of 1,000,000 km/hr, would carry 50,000 adults and would park in a shedbox.

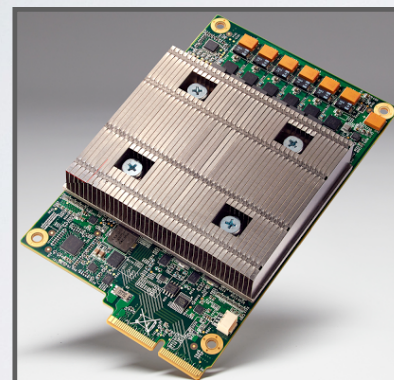
SIDE-NOTE: **ANTON** SUPERCOMPUTER



SIDE-NOTE: **GPUS** (GRAPHICAL PROCESSING UNITS)



SIDE-NOTE: **TPU** (TENSOR PROCESSING UNITS) FOR AI



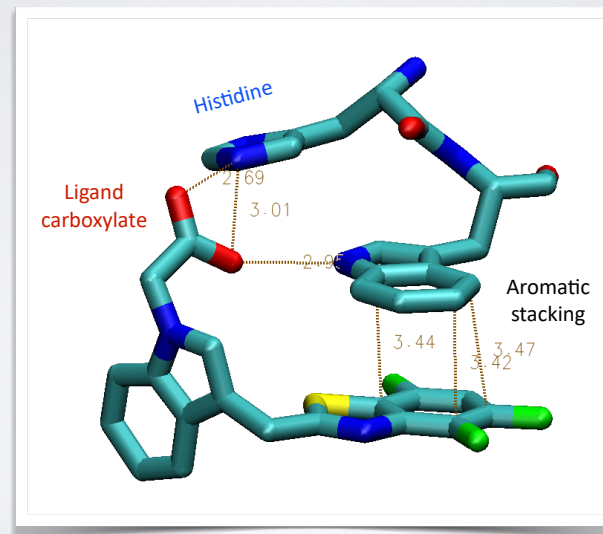
POTENTIAL FUNCTIONS DESCRIBE A SYSTEMS ENERGY AS A FUNCTION OF ITS STRUCTURE

Two main approaches:

(1). Physics-Based

(2). Knowledge-Based

KNOWLEDGE-BASED DOCKING POTENTIALS



ENERGY DETERMINES PROBABILITY (STABILITY)

Basic idea: Use probability as a proxy for energy



Boltzmann:

$$p(r) \propto e^{-E(r)/RT}$$



Inverse Boltzmann:

$$E(r) = -RT \ln[p(r)]$$

Example: ligand carboxylate O to protein histidine N

Find all protein-ligand structures in the PDB with a ligand carboxylate O

1. For each structure, histogram the distances from O to every histidine N
2. Sum the histograms over all structures to obtain $p(r_{O-N})$
3. Compute $E(r_{O-N})$ from $p(r_{O-N})$

KNOWLEDGE-BASED POTENTIALS

Weaknesses

Accuracy limited by availability of data

Strengths

Relatively easy to implement

Computationally fast

Status

Useful, far from perfect

May be at point of diminishing returns

(not always clear how to make improvements)

- Break -

The future? Combining AI and Physics based approaches

AlphaFold Protein Structure Database

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AlphaFold Protein Structure Database

Developed by DeepMind and EMBL-EBI

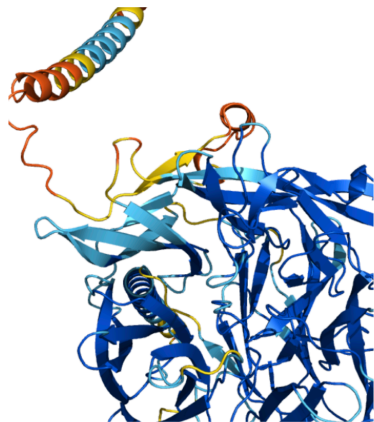
Search for protein, gene, UniProt accession or organism

Examples: [Free fatty acid receptor 2](#) [At1g58602](#) [Q5VSL9](#) [E. coli](#) Help: [AlphaFold DB search help](#)

AlphaFold DB provides open access to protein structure

AlphaFold is an AI system developed by DeepMind that predicts a protein's 3D structure from its amino acid sequence. It regularly achieves accuracy competitive with experiment.

DeepMind and EMBL's European Bioinformatics Institute (EMBL-EBI) have partnered to create AlphaFold DB to make these predictions freely available to the scientific community. The first release covers the human proteome and the proteomes of several other key organisms. In the coming months we plan to expand the database to cover a large proportion of all catalogued proteins (the over 100 million in UniRef90).



Q813H7: May protect the malaria parasite against attack by the immune system. Mean pLDDT 85.57.

nature

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NEWS | 30 November 2020

'It will change everything': DeepMind's AI makes gigantic leap in solving protein structures

Google's deep-learning program for determining the 3D shapes of proteins stands to transform biology, say scientists.

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One of biology's biggest mysteries 'largely solved' by AI

By Helen Briggs
BBC Science Correspondent

30 November 2020

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'The game has changed.' AI triumphs at solving protein structures

In milestone, software predictions finally match structures calculated from experimental data

30 NOV 2020 BY ROBERT F. SERVICE

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The Guardian For 200 years

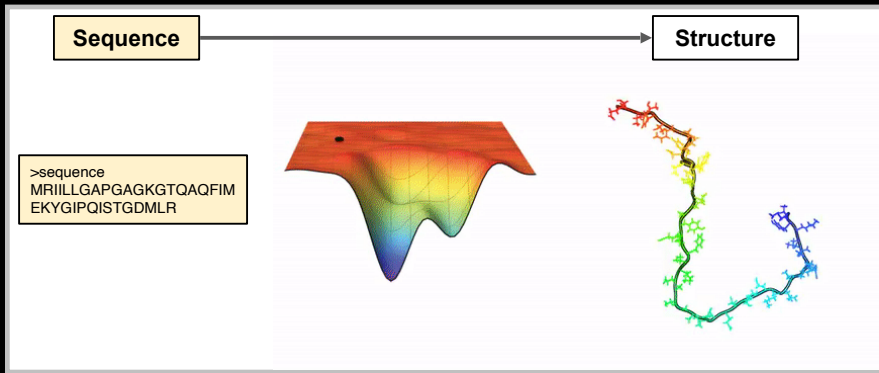
DeepMind AI cracks 50-year-old problem of protein folding

Program solves scientific problem in 'stunning advance' for understanding machinery of life

30 NOV 2020

Protein Folding Problem

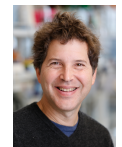
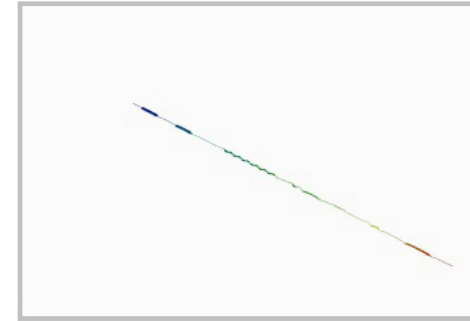
For a given **sequence**, find **structure** with lowest free energy



[Video credit: C. Fennell]

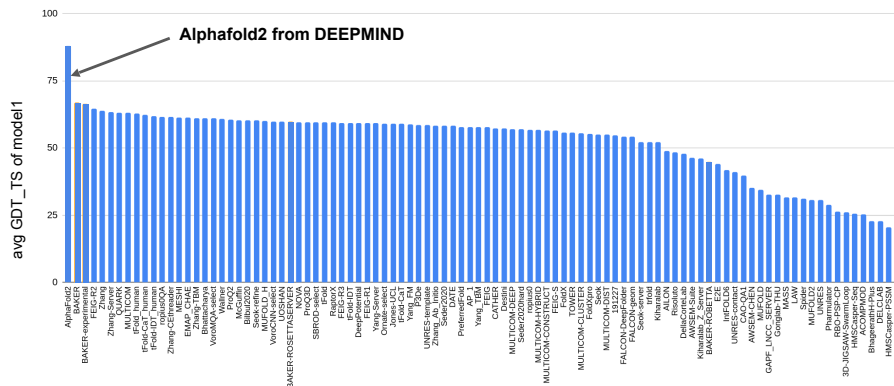
Dill, K.A. and MacCallum, J.L., 2012. The protein-folding problem, 50 years on. *science*, 338(6110), pp.1042-1046.

Rosetta - Protein "folding" with Energy function + fragments recombination



David Baker

Results from CASP14 (Critical Assessment of protein Structure Prediction)

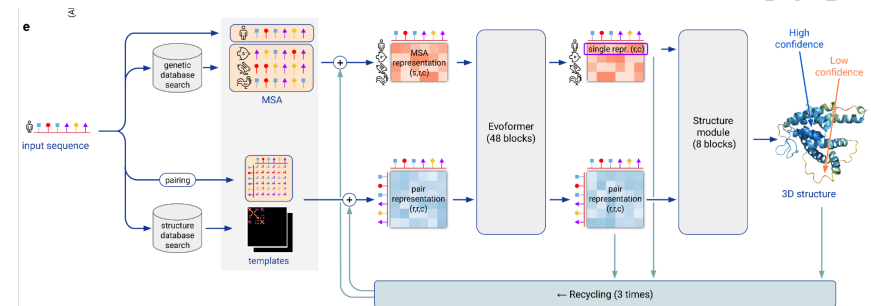


nature

<https://doi.org/10.1038/s41586-021-03819-2>

Accelerated Article Preview

Highly accurate protein structure prediction with AlphaFold



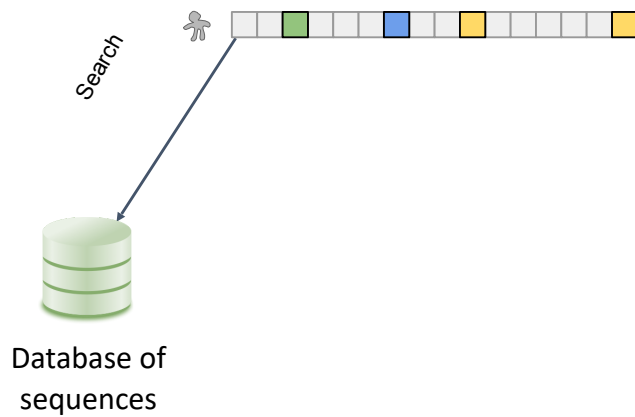
John Jumper, Richard Evans, Alexander Pritzel, Tim Green, Michael Figurnov, Olaf Ronneberger, Kathryn Tunyasuvunakool, Russ Bates, Augustin Židek, Anna Potapenko, Alex Bridgland, Clemens Meyer, Simon A. A. Kohl, Andrew J. Ballard, Andrew Cowie, Bernardino Romera-Paredes, Stanislav Nikolov, Rishub Jain, Jonas Adler, Trevor Back, Stig Petersen, David Reiman, Ellen Clancy, Michal Zielinski, Martin Steinegger, Michalina Pacholska, Tamas Berghammer, Sebastian Bodenstein, David Silver, Oriol Vinyals, Andrew W. Senior, Koray Kavukcuoglu, Pushmeet Kohli & Demis Hassabis

Multiple Sequence Alignments (MSAs)
are key inputs to these winning methods
(**alphafold2** and **RoseTTAFold**)

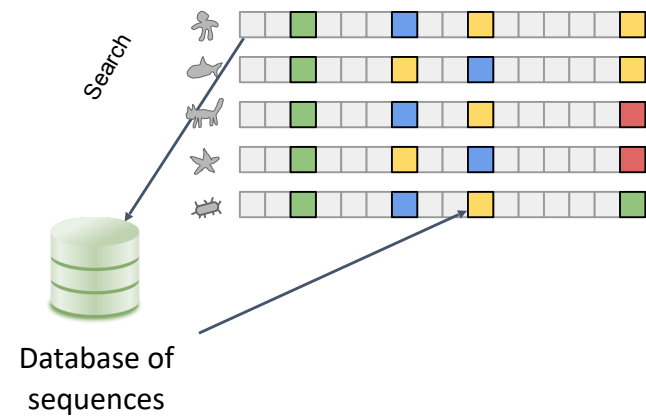
Start with a single sequence



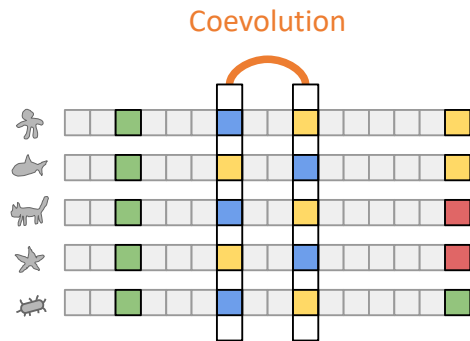
Search against a database of sequences



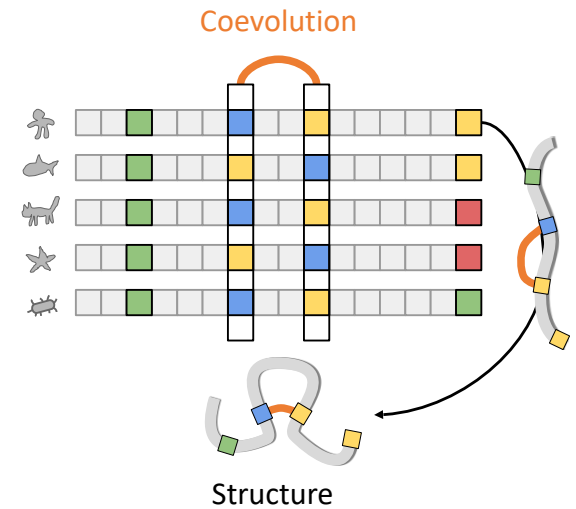
Generate a multiple sequence alignment



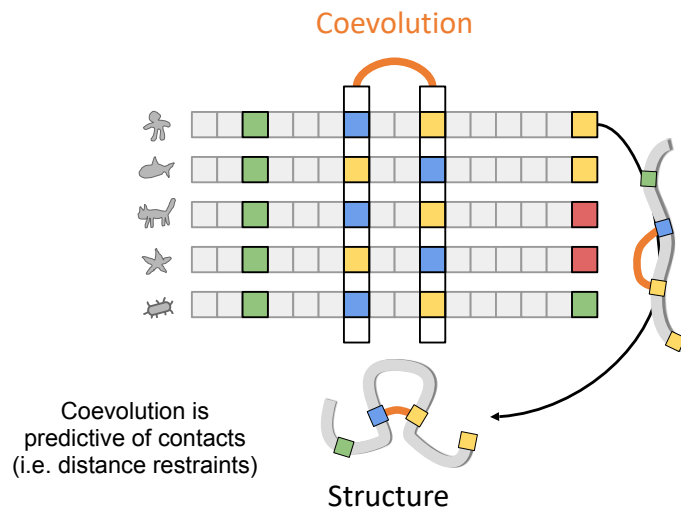
Analyze the MSA for coevolution



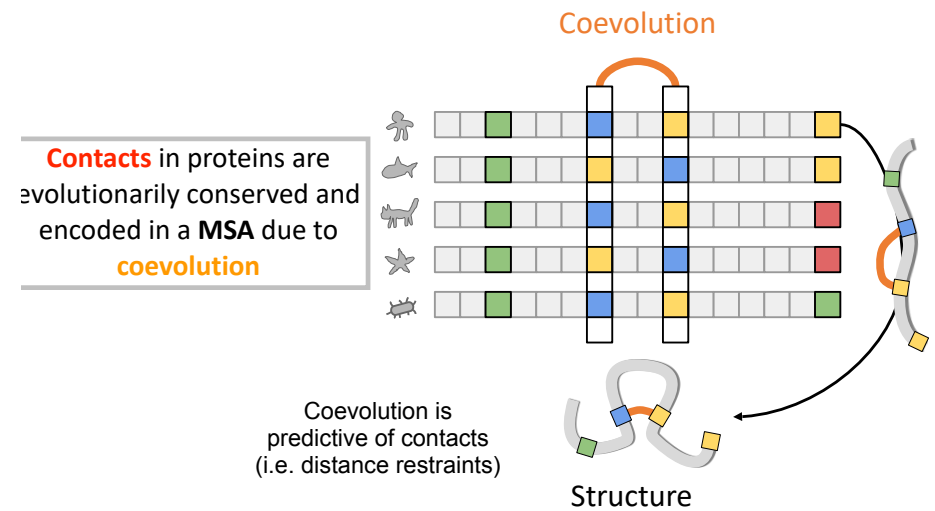
Use coevolution as restraints in folding simulations!



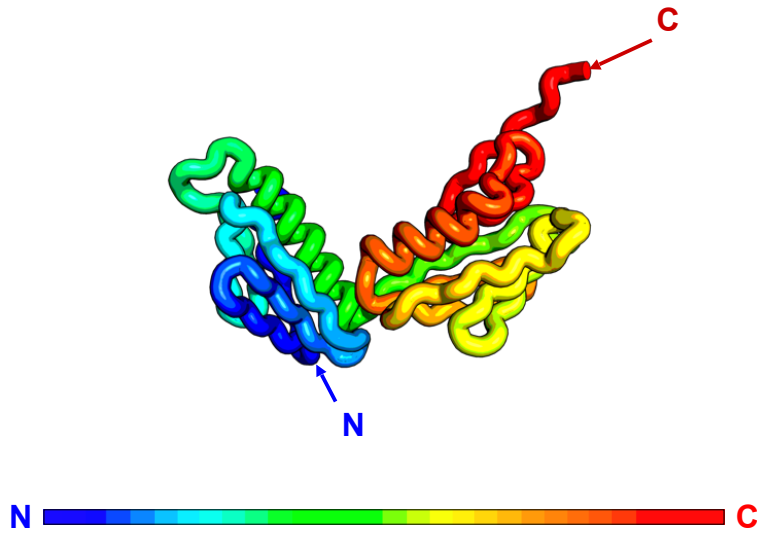
Use coevolution as restraints in folding simulations!



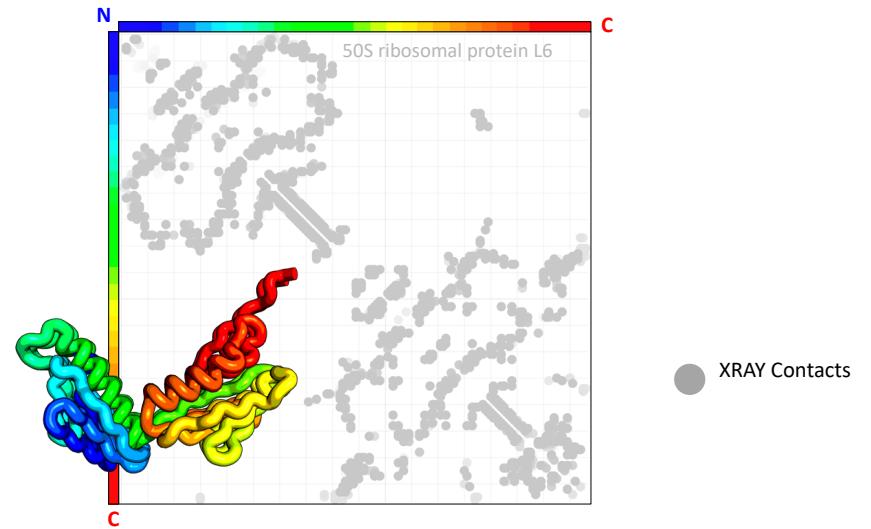
By measuring **coevolution**, we can infer **contacts**!



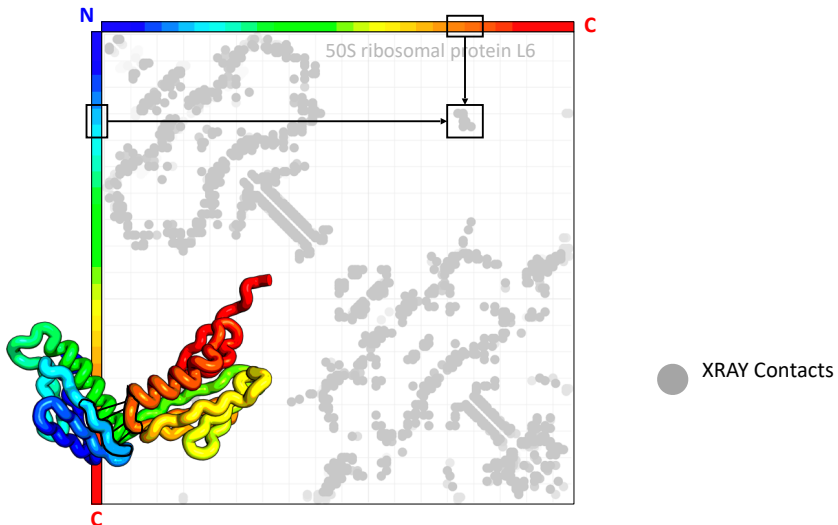
Review - How to read a contact/distance matrix?



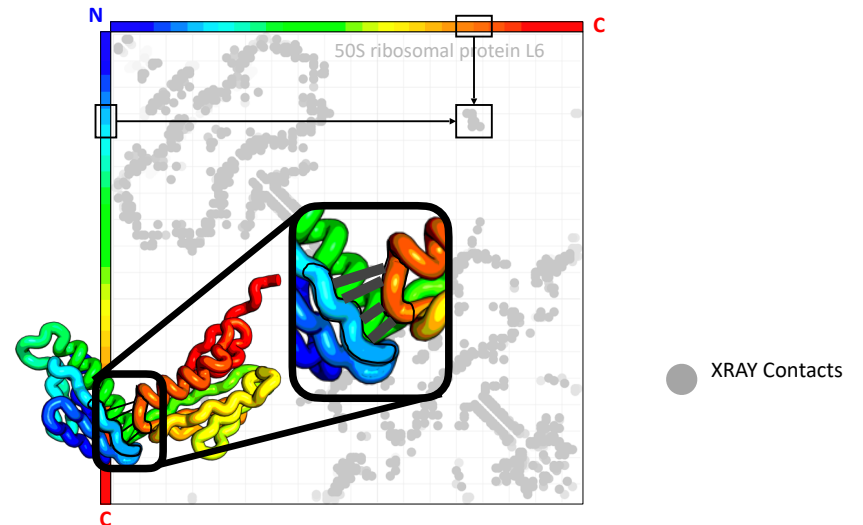
Contact map



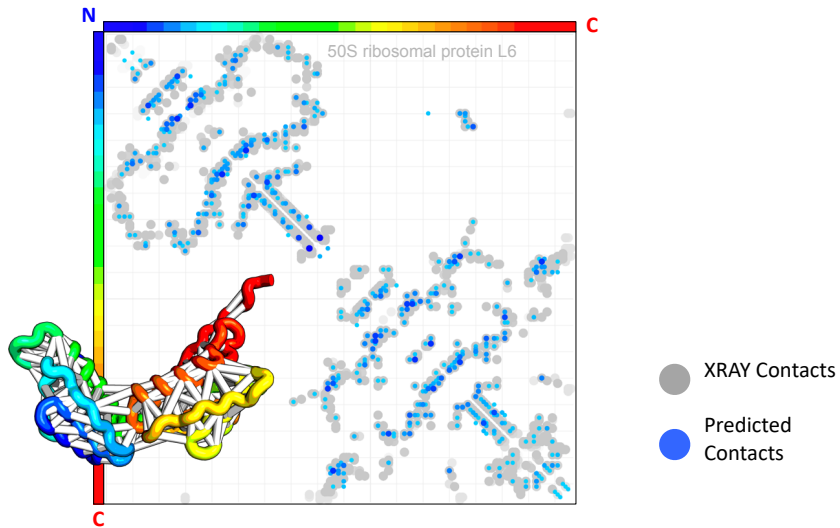
How to read a contact map



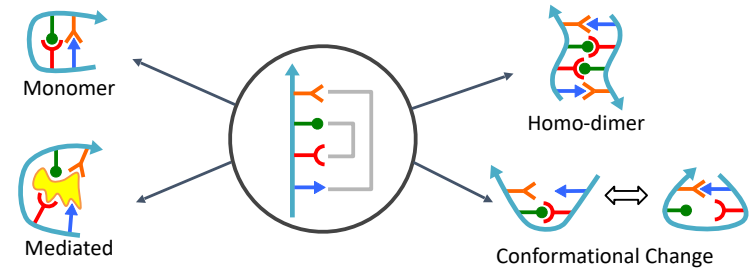
How to read a contact map



Overlay of predicted contacts on real contacts



The origin of contacts

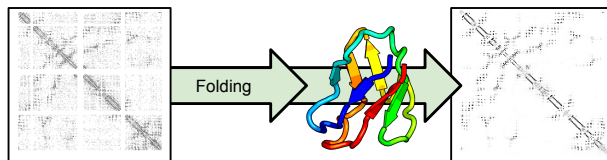


Anishchenko, I., Ovchinnikov, S., Kamisetty, H. and Baker, D., 2017. Origins of coevolution between residues distant in protein 3D structures. *PNAS*, 114(34), pp.9122-9127.

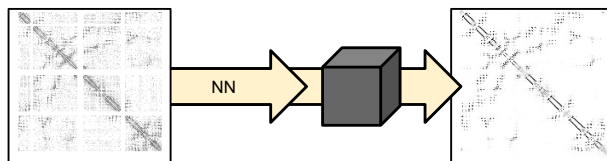
Slide Credit: Sergey Ovchinnikov (@sokrypton)

How to solve this problem?

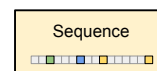
- Enumerate folds and see which matches contacts best
- Try different number (or combination) of restraints
- Lots of sampling with ambiguous restraints



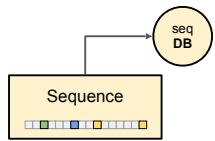
- Use NN to filter/enhance contacts before trying to fold



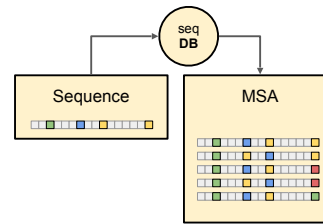
Alphafold2



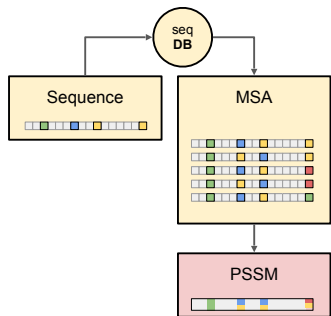
AlphaFold2



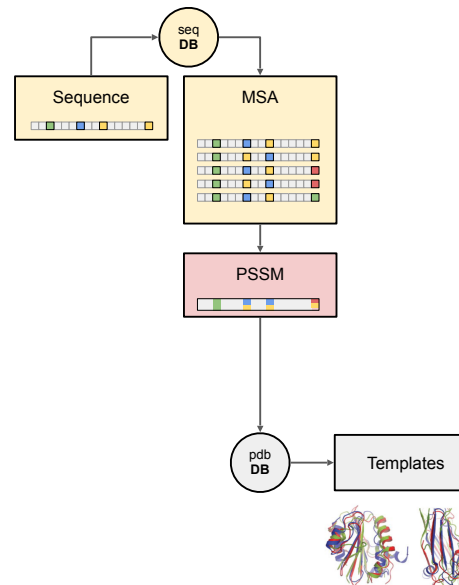
AlphaFold2



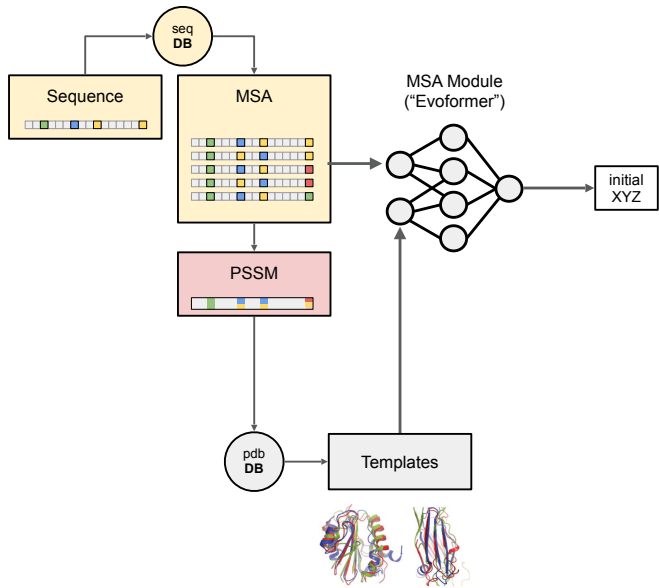
AlphaFold2



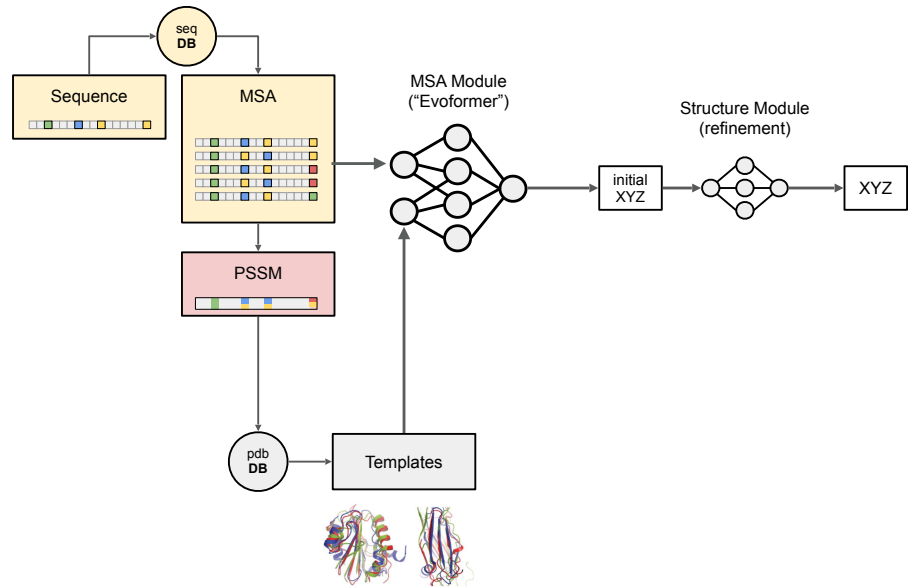
AlphaFold2



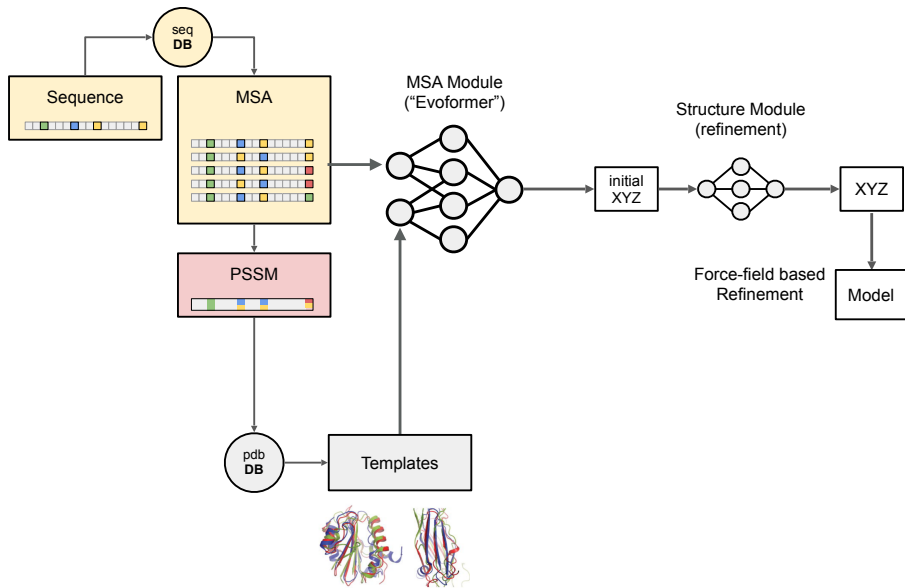
AlphaFold2



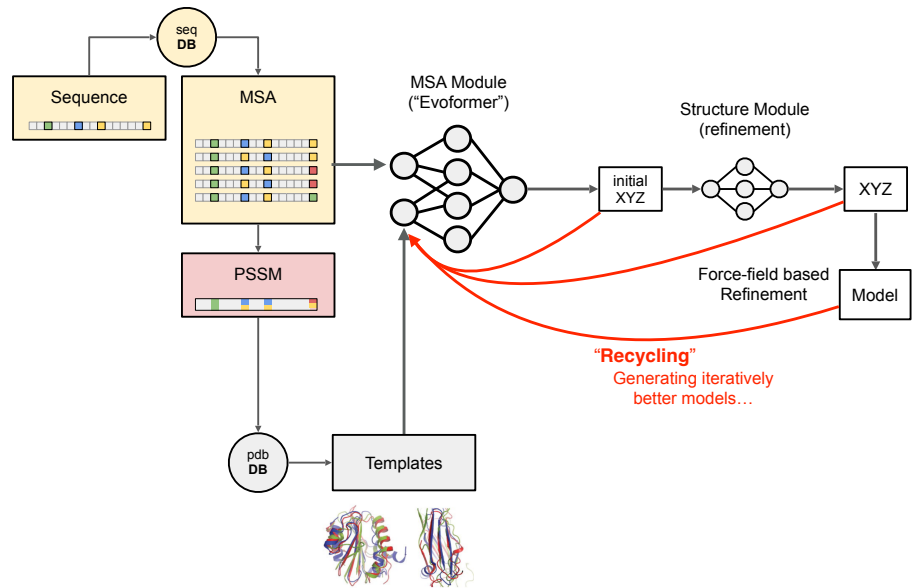
AlphaFold2



AlphaFold2

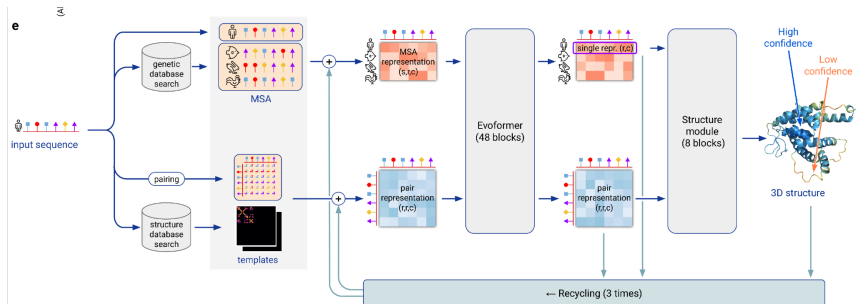


AlphaFold2 - New Critical detail **Recycling**



Accelerated Article Preview

Highly accurate protein structure prediction with AlphaFold



Received: 11 May 2021

Accepted: 12 July 2021

Accelerated Article Preview Published online 15 July 2021

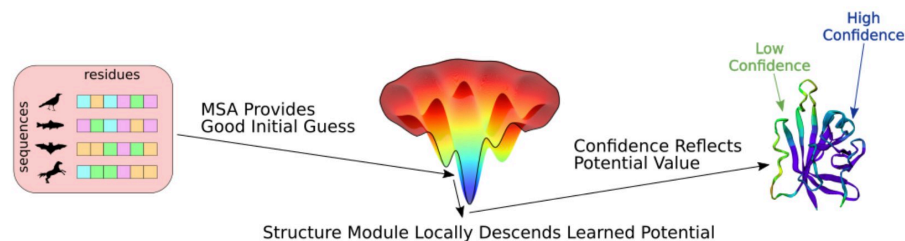
Cite this article as: Jumper, J. et al. Highly accurate protein structure prediction with AlphaFold. *Nature* <https://doi.org/10.1038/s41586-021-03819-2> (2021).

John Jumper, Richard Evans, Alexander Pritzel, Tim Green, Michael Figurnov, Olaf Ronneberger, Kathryn Tunyasuvunakool, Russ Bates, Augustin Zidek, Anna Potapenko, Alex Bridgland, Clemens Meyer, Simon A. A. Kohl, Andrew J. Ballard, Andrew Cowie, Bernardino Romera-Paredes, Stanislav Nikolov, Rishub Jain, Jonas Adler, Trevor Back, Stig Petersen, David Reiman, Ellen Clancy, Michal Zieliński, Martin Steinegger, Michalina Pacholska, Tamas Berghammer, Sebastian Bodenstein, David Silver, Oriol Vinyals, Andrew W. Senior, Koray Kavukcuoglu, Pushmeet Kohli & Demis Hassabis

This is a PDF file of a peer-reviewed paper that has been accepted for publication.

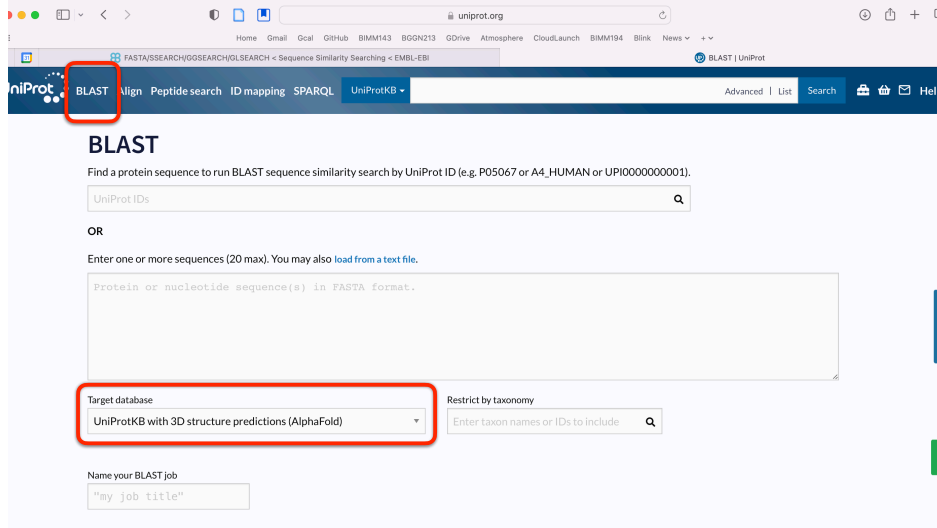
Hypothesis:

AlphaFold uses input MSA/Templates to "solve" the global search problem. The rest of the model refines the structure using the learned energy potential.



Search UniProt with AlphaFold

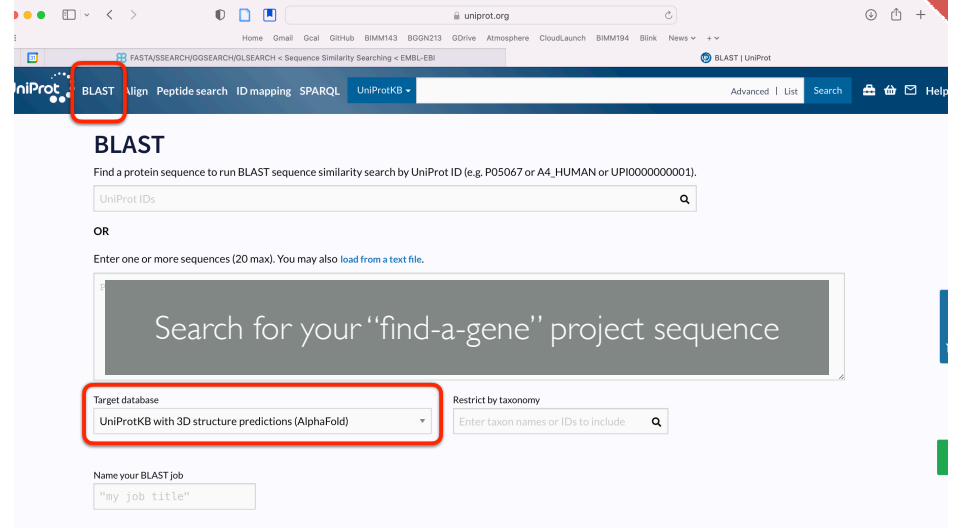
<https://www.uniprot.org/blast>



Search UniProt with AlphaFold

<https://www.uniprot.org/blast>

Do it Yourself!



<https://www.ebi.ac.uk/Tools/sss/fasta/>

Do it Yourself!

PROTEIN DATABASES

2 Databases Selected

X Clear Selection

- UniProt Knowledgebase (The UniProt Knowledgebase includes UniProtKB/Swiss-Prot and UniProtKB/TrEMBL)
- UniProtKB/Swiss-Prot (The manually annotated section of UniProtKB)
- UniProtKB/Swiss-Prot isoforms (The manually annotated isoforms of UniProtKB/Swiss-Prot)
- UniProtKB/TrEMBL (The automatically annotated section of UniProtKB)
- UniProtKB Reference Proteomes plus Swiss-Prot
- UniProtKB COVID-19
- UniProtKB Taxonomi
- UniProt Clusters
- Patents
- Structures
 - Protein Structure Sequences (PDBe protein structure sequences)
 - AlphaFold DB
 - UniProtKB PDB
- Other Protein Databases

Search for your Find-a-gene project sequence in AlphaFold DB

<https://www.ebi.ac.uk/Tools/sss/fasta/>

PROTEIN DATABASES

2 Databases Selected

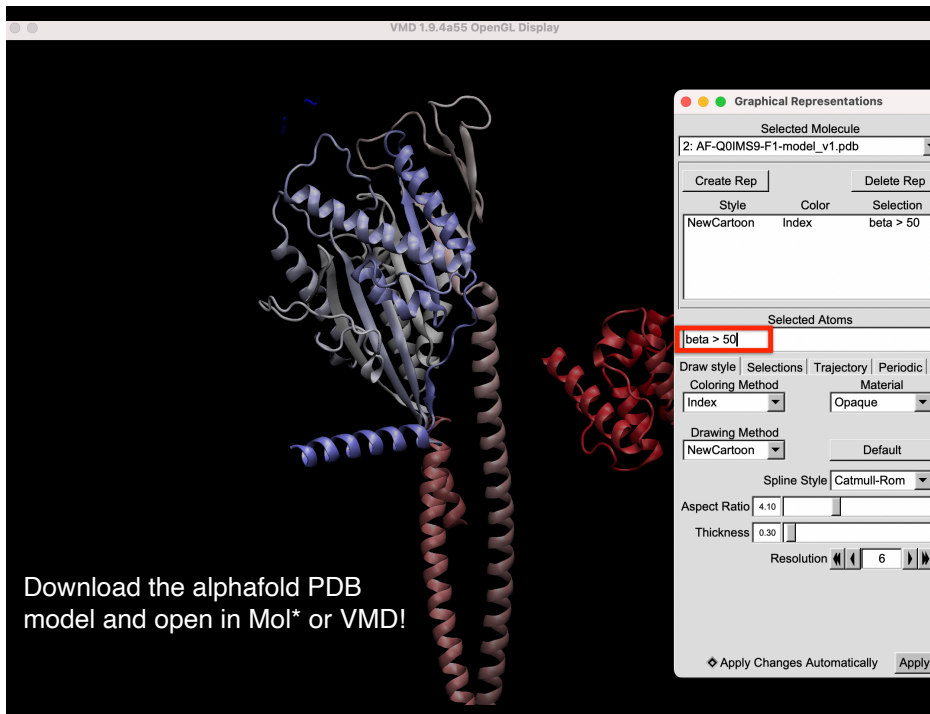
X Clear Selection

- UniProt Knowledgebase (The UniProt Knowledgebase includes UniProtKB/Swiss-Prot and UniProtKB/TrEMBL)
- UniProtKB/Swiss-Prot (The manually annotated section of UniProtKB)
- UniProtKB/Swiss-Prot isoforms (The manually annotated isoforms of UniProtKB/Swiss-Prot)
- UniProtKB/TrEMBL (The automatically annotated section of UniProtKB)
- UniProtKB Reference Proteomes plus Swiss-Prot
- UniProtKB COVID-19
- UniProtKB Taxonomi
- UniProt Clusters
- Patents
- Structures
 - Protein Structure Sequences (PDBe protein structure sequences)
 - AlphaFold DB
 - UniProtKB PDB
- Other Protein Databases

Search for your Find-a-gene project sequence in alpha fold DB

Or: "KIN-I4Q"

Or: Q8W3K0



AlphaFold low confidence regions

- AlphaFold produces a per-residue confidence score (**pLDDT**) between 0 and 100 that is written to the B-factor column.
- To remove low confidence regions (with low pLDDT scores)

```
p <- read.pdb("AF-model.pdb")
# Find atoms with good confidence score (pLDDT)
atoms <- which( p$atom$b > 70 )
# Trim to selected atoms
p2 <- trim.pdb(p, as.select( atoms ) )
write.pdb(p2, file="high_confidence_model.pdb")
```

<https://github.com/sokrypton/ColabFold>

Evolutionary scale modeling (ESM)

For short monomeric proteins (< 400 amino acids) consider using the new **ESMFold**


<https://esmatlas.com/>

[No need for GPU & comparatively fast]

Alternative: Language Models

- **AlphaFold** (and related methods) need to search through large protein databases to identify related sequences.
- They require a large group of evolutionarily related sequences as input so that they can extract the patterns that are linked to structure.
- **ESM-fold** uses a language model that learns these evolutionary patterns during its training on protein sequences, enabling faster structure prediction from a single sequence.

Brand new!



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Revision Summary

bioRxiv posts many COVID-19-related papers. A reminder: they have not been formally peer-reviewed and should not guide health-related behavior or be reported in the press as conclusive.

New Results Follow this preprint

Evolutionary-scale prediction of atomic level protein structure with a language model

Zeming Lin, Haili Akin, Roshan Rao, Brian Hie, Zhongkai Zhu, Wenting Lu, Nikita Smetanin, Robert Verkuil, Ori Kabeli, Yaniv Shmueli, Allan dos Santos Costa, Maryam Fazel-Zarandi, Tom Sercu, Salvatore Candido, Alexander Rives

doi: <https://doi.org/10.1101/2022.07.20.500902>

This article is a preprint and has not been certified by peer review [what does this mean?]

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Abstract
Full Text
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Abstract

Artificial intelligence has the potential to open insight into the structure of proteins at the scale of evolution. It has only recently been possible to extend protein structure prediction to two hundred million cataloged proteins. Characterizing the structures of the exponentially growing billions of protein sequences revealed by large scale gene sequencing experiments would necessitate a break-through in the speed of folding. Here we show that direct inference of structure from primary sequence using a large language model enables an order of

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>Neuraminidase
 VKLAGNSSLCPINGWAVYKSDNSIRIGSKGDFVIREPFISSCHLECRFFLTQGALLNDKXHMSTVKDR
 SPHRTLMSCPVGEAPSPNSRFESVAVWSASACHDGTSLWLTIGSPDNGAVAWLYKNGIITDTKSWRN
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Try an example:

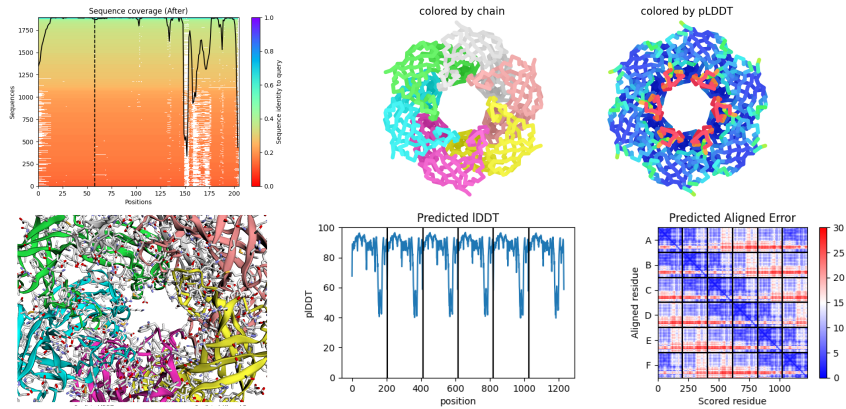
Plastic degradation protein - PETase
Antifreeze protein - 1EZO
AI-generated protein - BCYK

7-bladed propeller fold - Neuraminidase

redict protein structure with ESMFold Meta A

ColabFold

Making Protein folding accessible via Google Colab



github.com/sokrypton/ColabFold

<https://github.com/sokrypton/ColabFold>