



**BIMM 143**  
**Structural Bioinformatics**

Lecture 11

**Barry Grant**  
**UC San Diego**

<http://thegrantlab.org/bimm143>

<http://www.ks.uiuc.edu/Development/Download/download.cgi>

*“Bioinformatics is the application of computers to the collection, archiving, organization, and analysis of biological data.”*

... A hybrid of biology and computer science

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**Bioinformatics is computer aided biology!**

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**Bioinformatics is computer aided biology!**

**Goal: Data to Knowledge**

So what is **structural bioinformatics**?

So what is **structural bioinformatics**?

**... computer aided structural biology!**

Aims to characterize and interpret biomolecules and their assemblies at the molecular & atomic level

**Why should we care?**

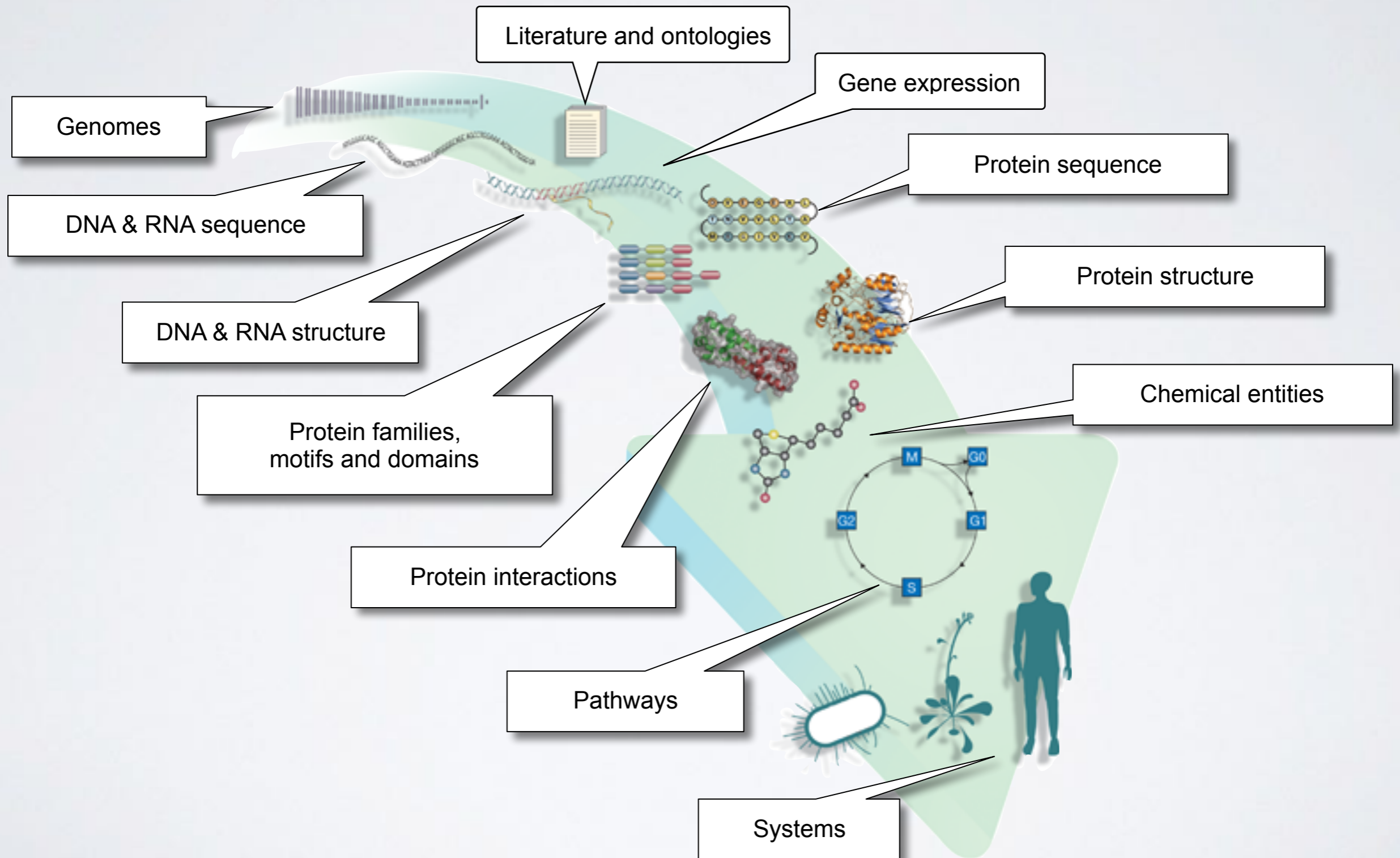
# Why should we care?

Because biomolecules are “nature’s robots”

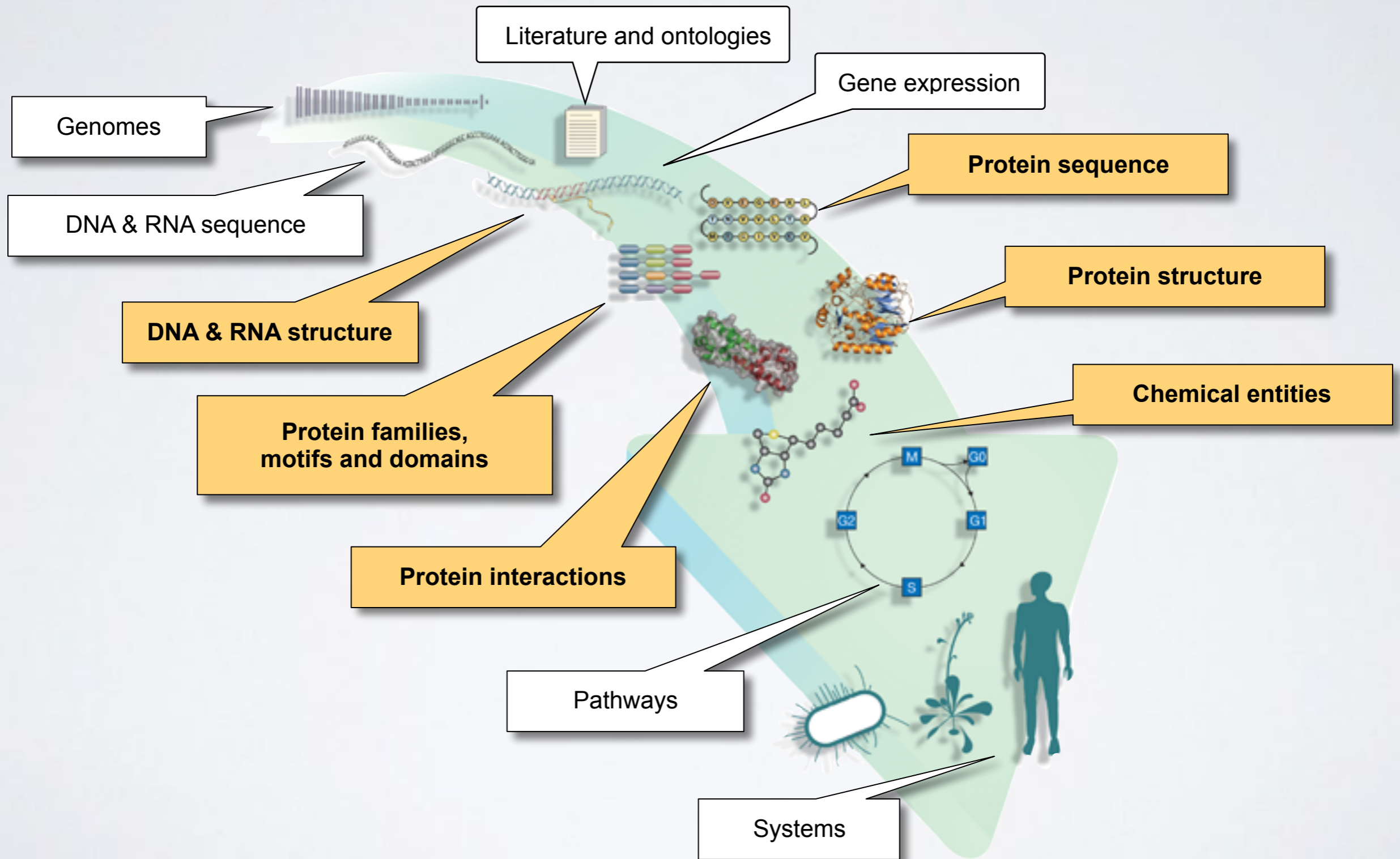
... and because it is only by coiling into **specific 3D structures** that they are able to perform their functions



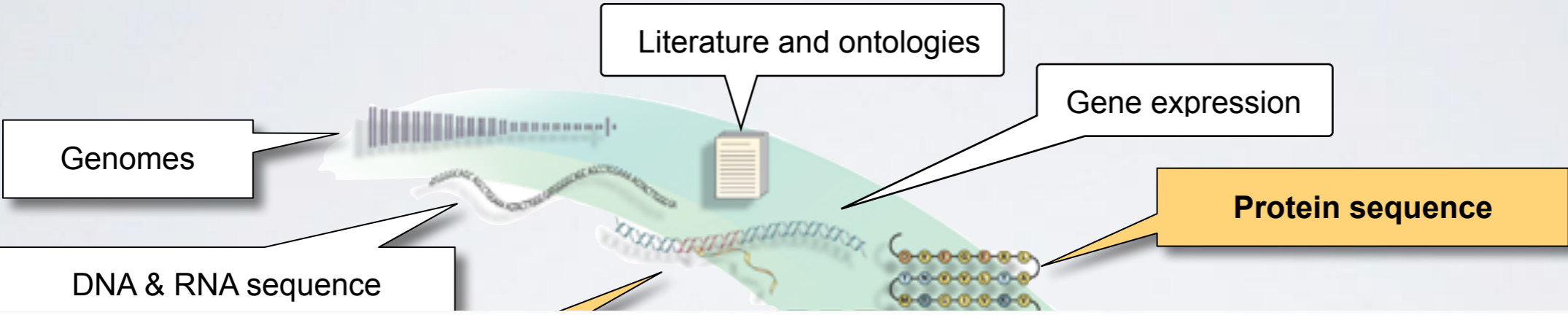
# BIOINFORMATICS DATA



# STRUCTURAL DATA IS CENTRAL



# STRUCTURAL DATA IS CENTRAL



**Sequence > Structure > Function**

DNA & RNA structure

Protein structure

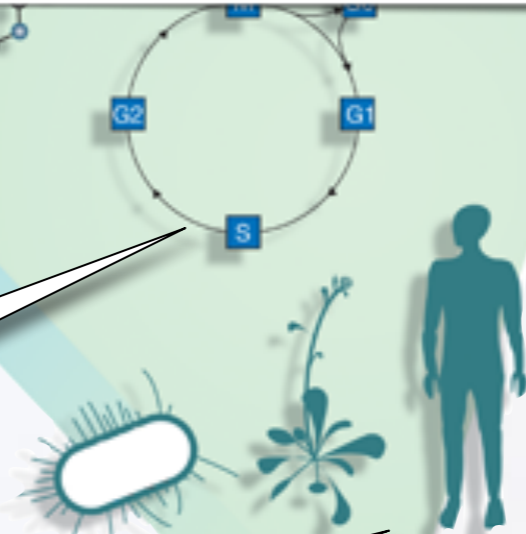
Protein families,  
motifs and domains

Chemical entities

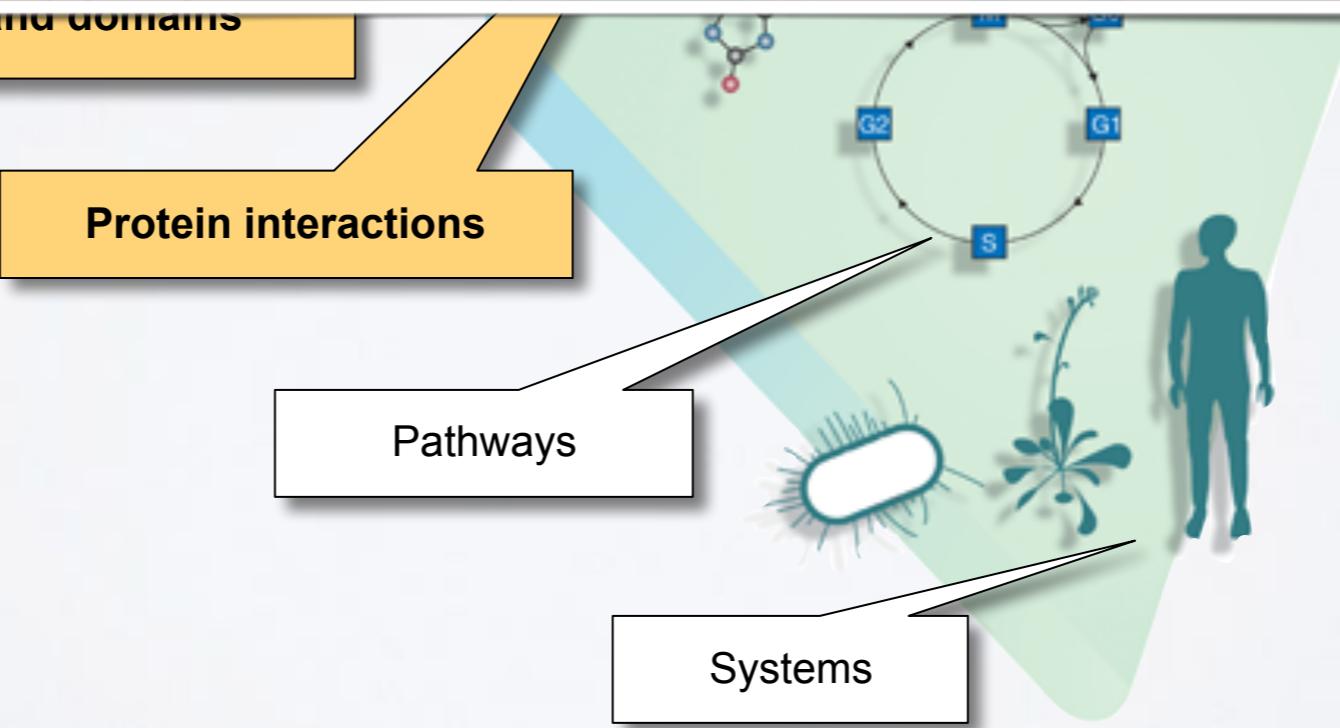
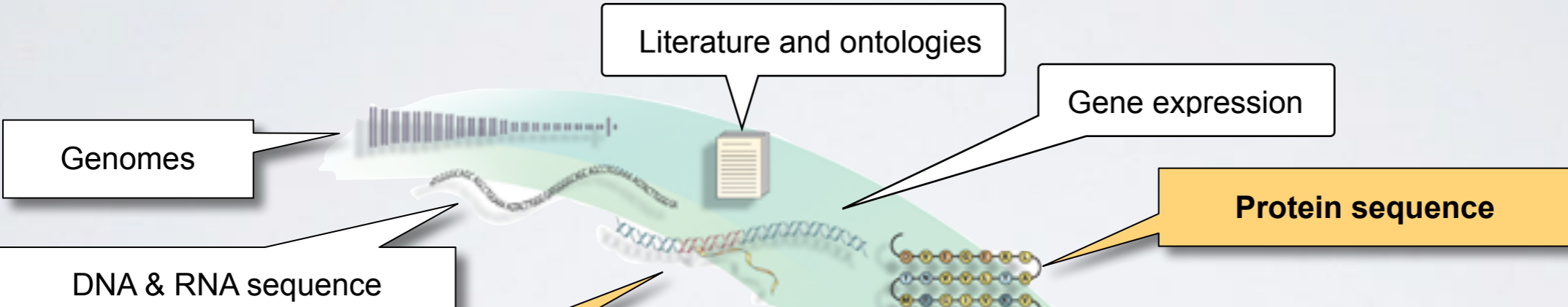
Protein interactions

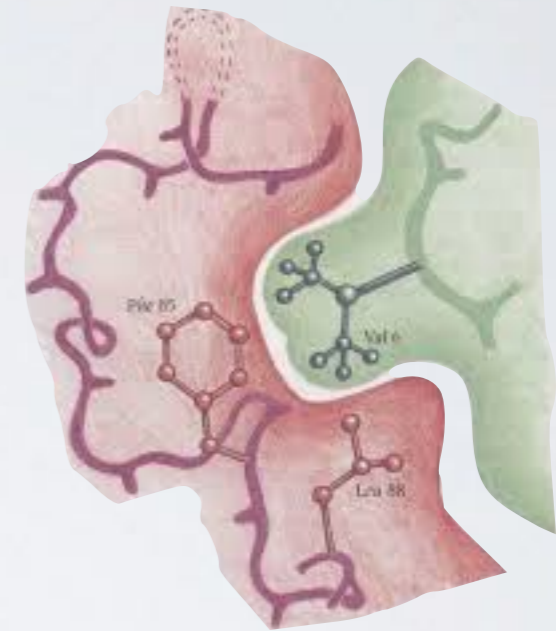
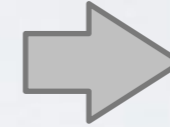
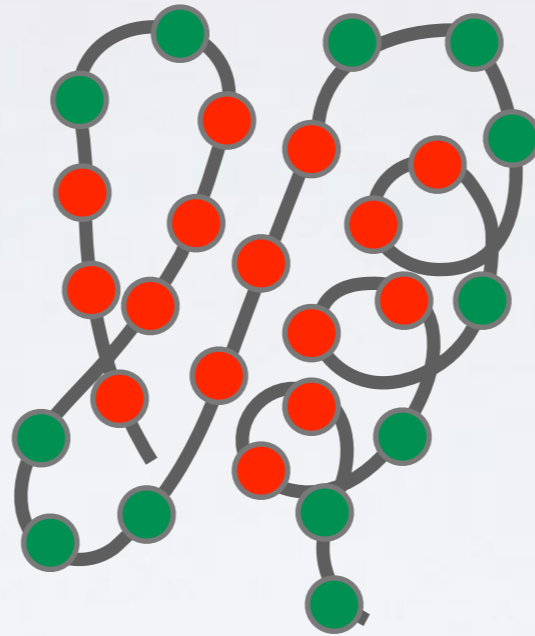
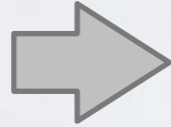
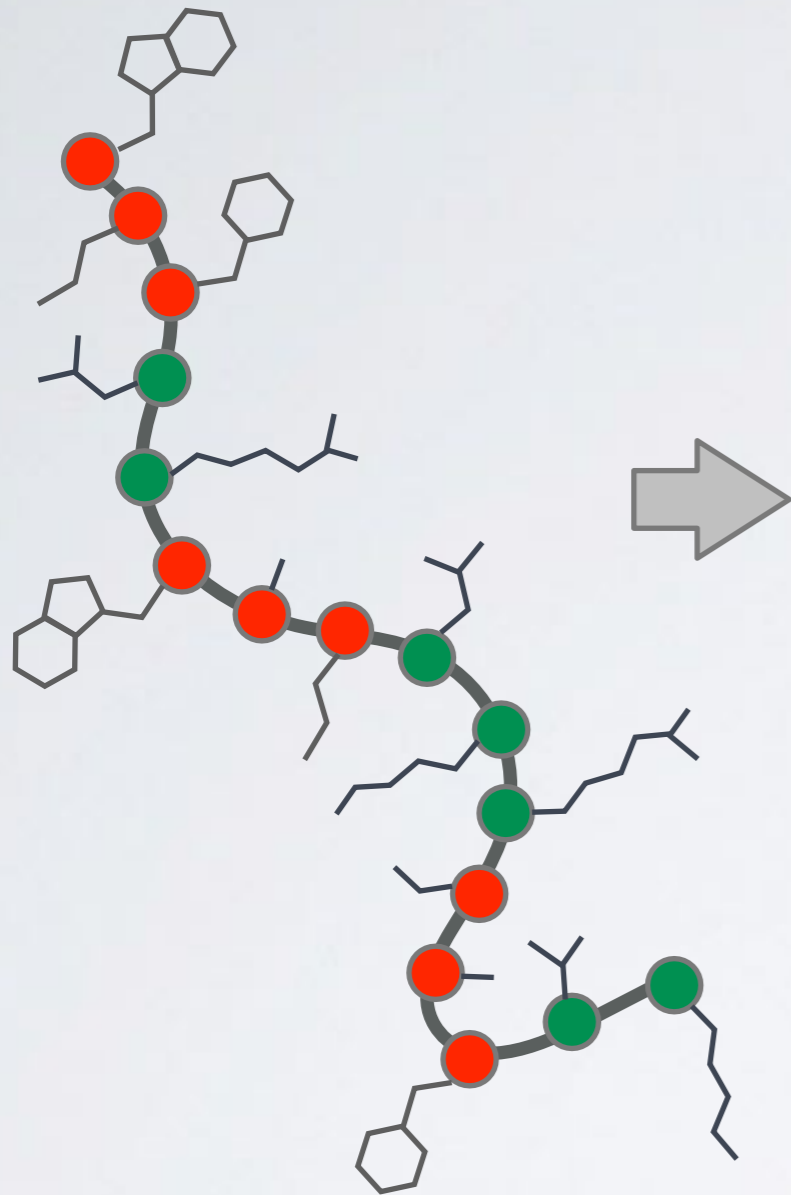
Pathways

Systems



# STRUCTURAL DATA IS CENTRAL





## Sequence

- Unfolded chain of amino acid chain
- Highly mobile
- Inactive

## Structure

- Ordered in a precise 3D arrangement
- Stable but dynamic

## Function

- Active in specific "conformations"
- Specific associations & precise reactions

In daily life, we use machines with functional *structure* and *moving parts*



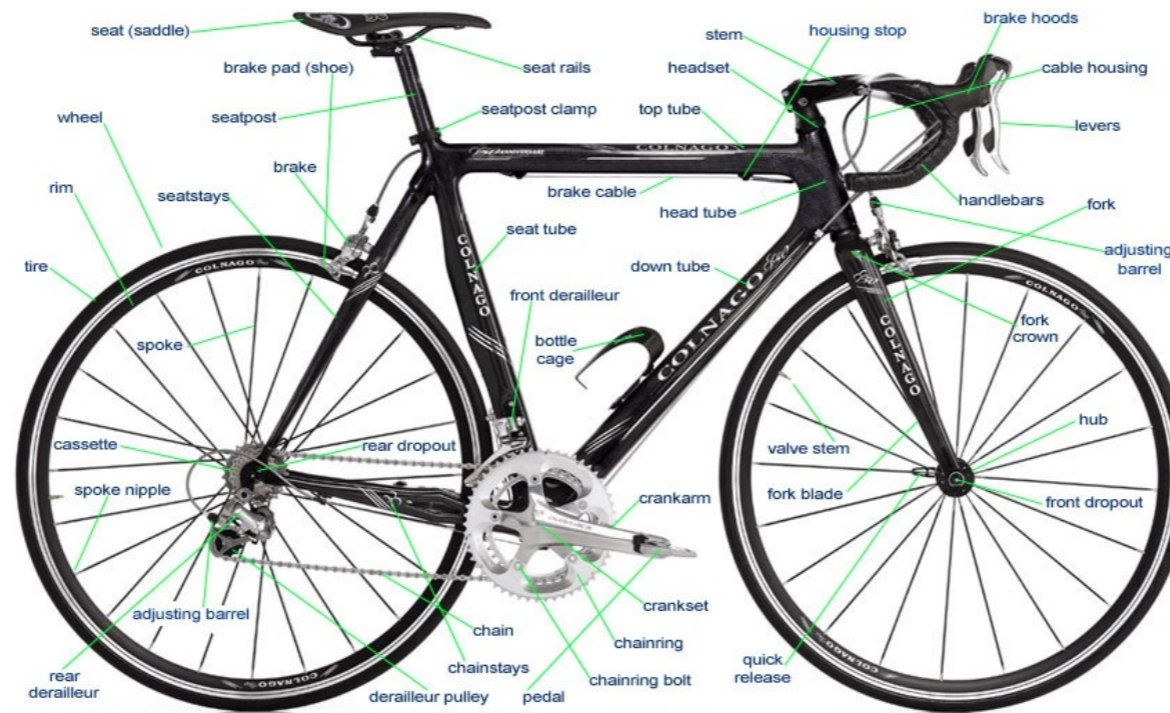
# Genomics is a great start ....

## Track Bike – DL 175

| REF. NO. | IBM NO. | DESCRIPTION  |
|----------|---------|--|
| 1        | 156011  | Track Frame 21", 22", 23", 24", Team Red               |
| 2        | 157040  | Fork for 21" Frame                                     |
| 2        | 157039  | Fork for 22" Frame                                     |
| 2        | 157038  | Fork for 23" Frame                                     |
| 2        | 157037  | Fork for 24" Frame                                     |
| 3        | 191202  | Handlebar TTT Competition Track Alloy 15/16"           |
| 4        |         | Handlebar Stem, TTT, Specify extension                 |
| 5        | 191278  | Expander Bolt  |
| 6        | 191272  | Clamp Bolt   |
| 7        | 145841  | Headset Complete 1 x 24 BSC                            |
| 8        | 145842  | Ball Bearings  |
| 9        | 190420  | 175 Raleigh Pistard Seta Tubular Prestavalve 27"       |
| 10       | 190233  | Rim, 27" AVA Competition (36H) Alloy Prestavalve       |
| 11       | 145973  | Hub, Large Flange Campagnolo Pista Track Alloy (pairs) |
| 12       | 190014  | Spokes, 11 5/8"  |
| 13       | 145837  | Sleeve   |
| 14       | 145636  | Ball Bearings  |
| 15       | 145170  | Bottom Bracket Axle                                    |
| 16       | 145838  | Cone for Sleeve  |
| 17       | 146473  | L.H. Adjustable Cup                                    |
| 18       | 145833  | Lockring   |
| 19       | 145239  | Straps for Toe Clips                                   |
| 20       | 145834  | Fixing Bolt  |
| 21       | 145835  | Fixing Washer  |
| 22       | 145822  | Dustcap  |
| 23       | 145823  | R.H. and L.H. Crankset with Chainwheel                 |
| 24       | 146472  | Fixed Cup  |
| 25       | 145235  | Toe Clips, Christophe, Chrome (Medium)                 |
| 26       | 145684  | Pedals, Extra Light, Pairs                             |
| 27       | 123021  | Chain  |
| 28       | 145980  | Seat Post  |
| 29       |         | Seat Post Bolt and Nut                                 |
| 30       | 167002  | Saddle, Brooks   |
| 31       | 145933  | Track Sprocket, Specify 12, 13, 14, 15, or 16 T.       |

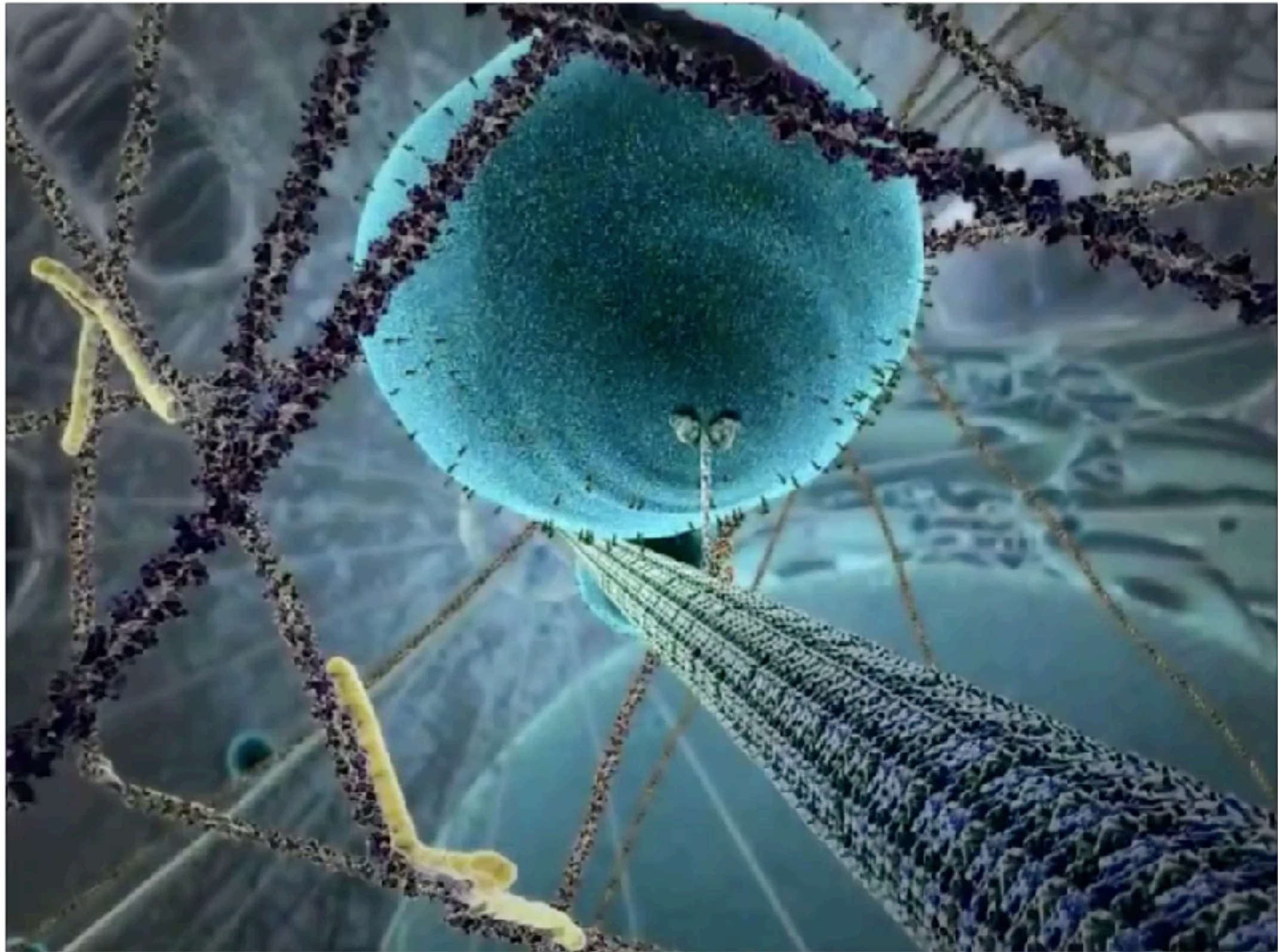
- But a parts list is not enough to understand how a bicycle works

# ... but not the end

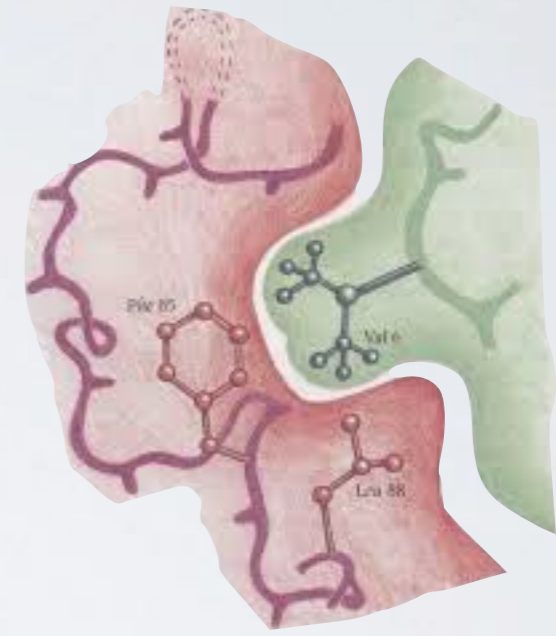
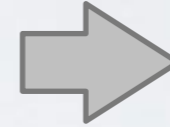
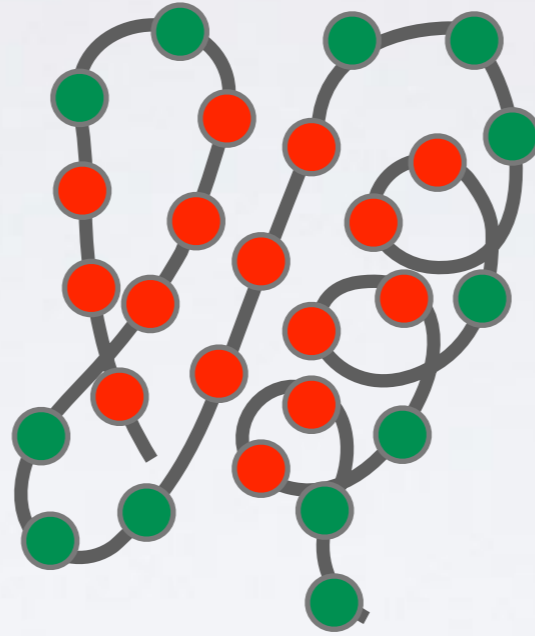
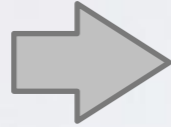
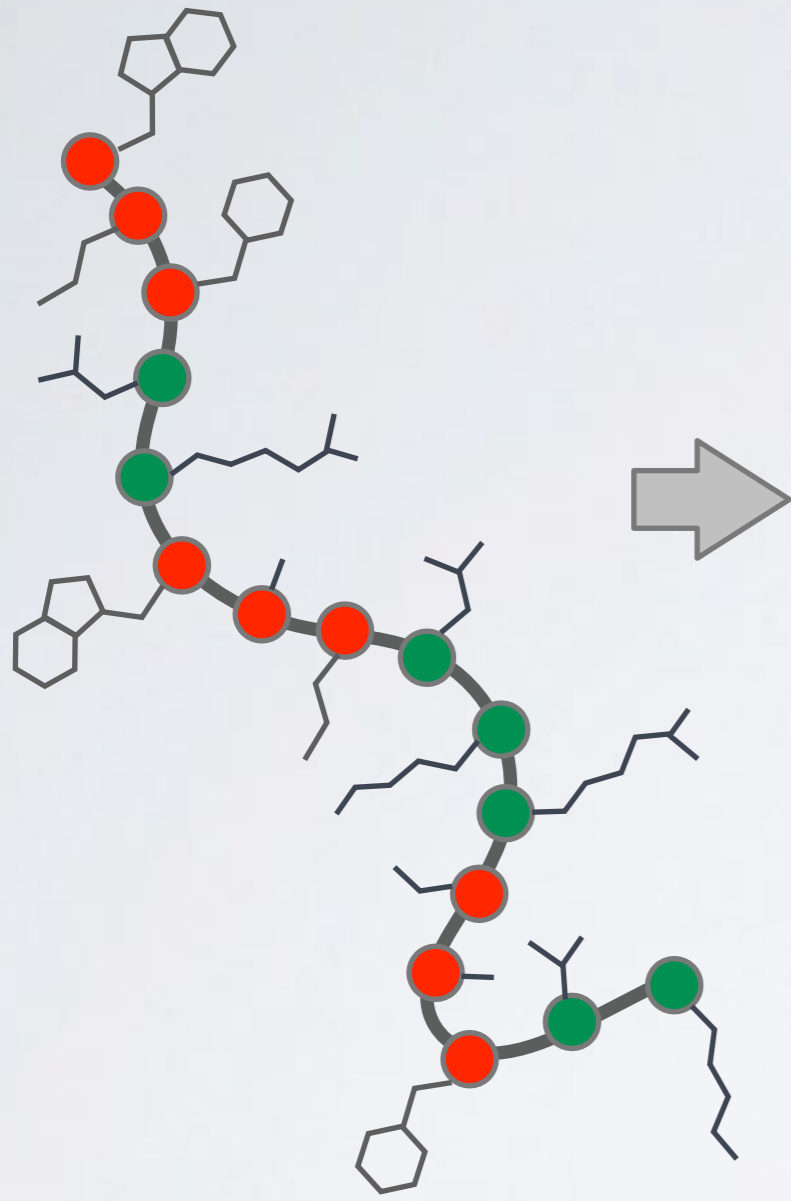


- We want the full spatiotemporal picture, and an ability to control it
- Broad applications, including drug design, medical diagnostics, chemical manufacturing, and energy





Extracted from The Inner Life of a Cell by Cellular Visions and Harvard  
[YouTube link: <https://www.youtube.com/watch?v=y-uuk4Pr2i8> ]



## Sequence

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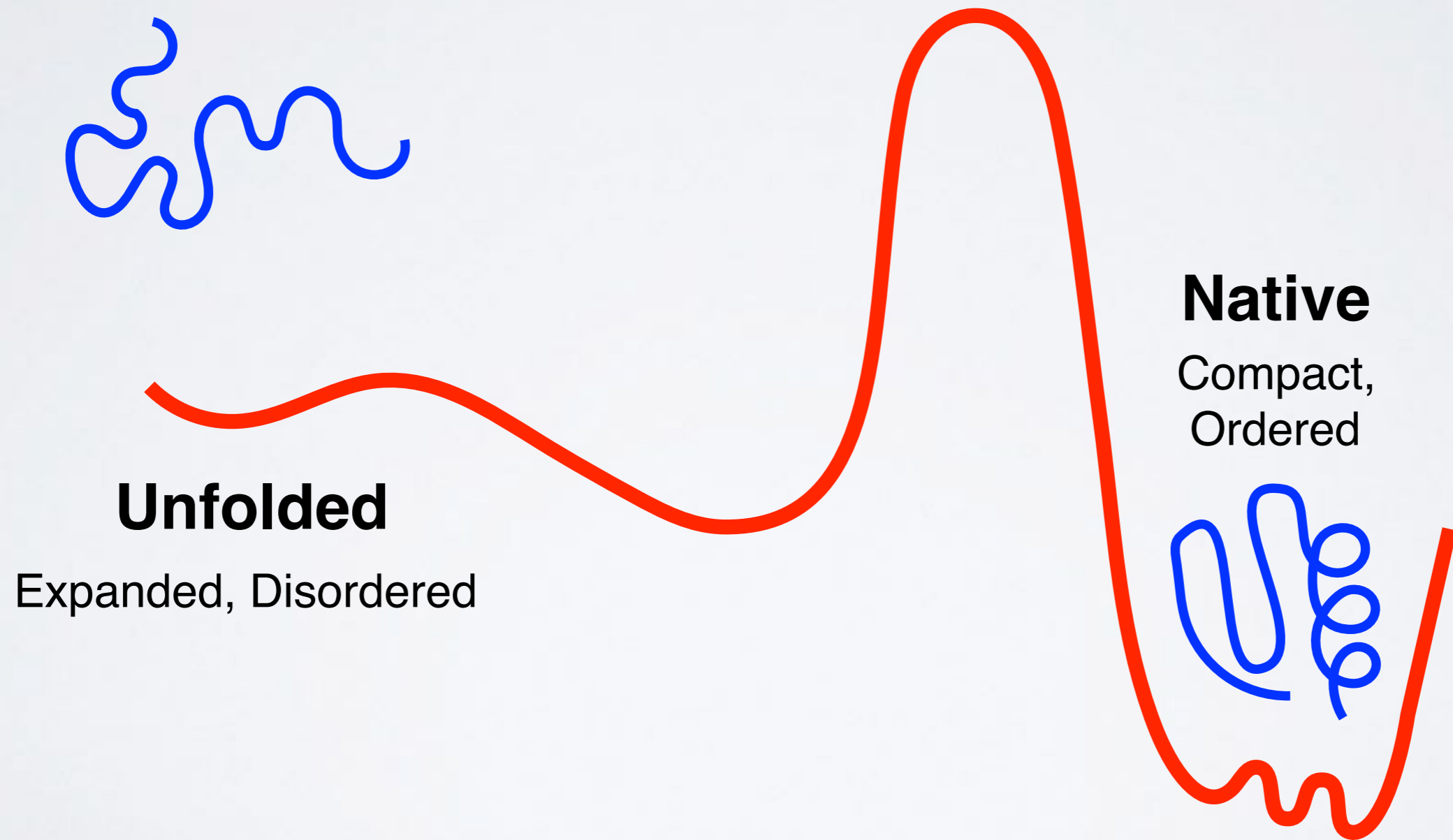
## Structure

- Ordered in a precise 3D arrangement
- Stable but dynamic

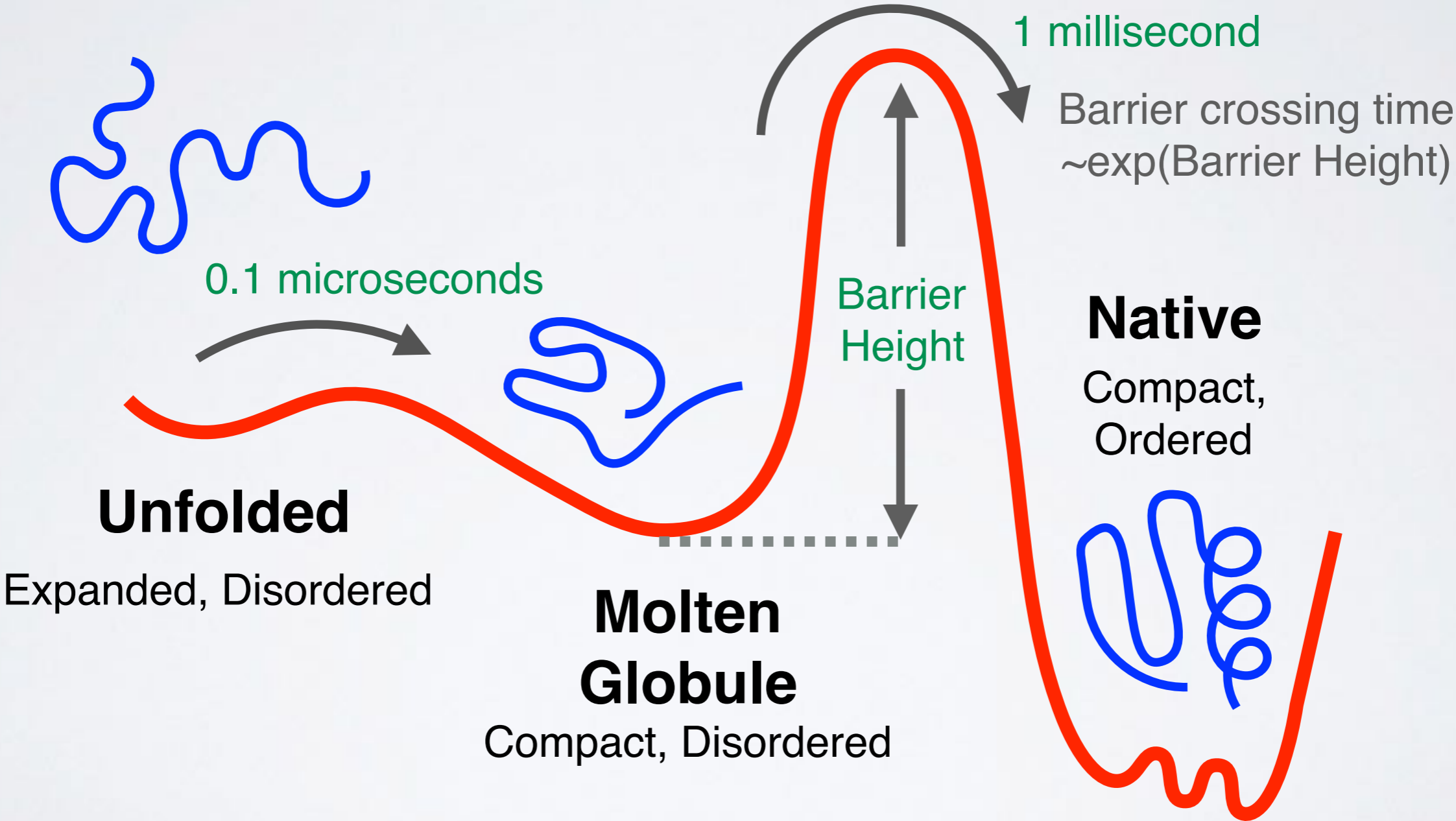
## Function

- Active in specific "conformations"
- Specific associations & precise reactions

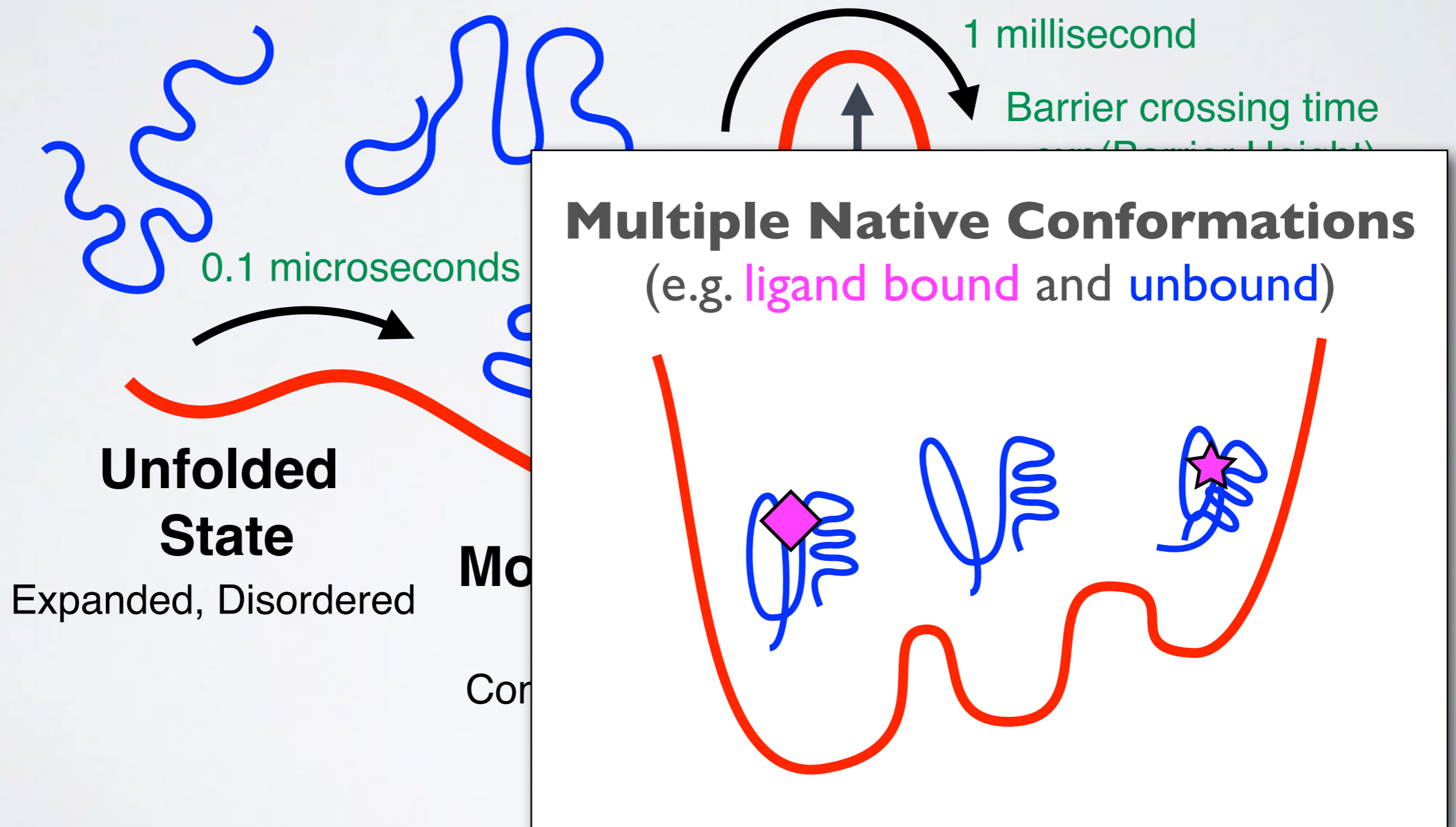
# KEY CONCEPT: ENERGY LANDSCAPE



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# OUTLINE:

- ▶ **Overview of structural bioinformatics**
  - Major motivations, goals and challenges
- ▶ **Fundamentals of protein structure**
  - Composition, form, forces and dynamics
- ▶ **Representing and interpreting protein structure**
  - Modeling energy as a function of structure
- ▶ **Example application areas**
  - Predicting functional dynamics & drug discovery

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- ▶ **Overview of structural bioinformatics**

- Major motivations, goals and challenges

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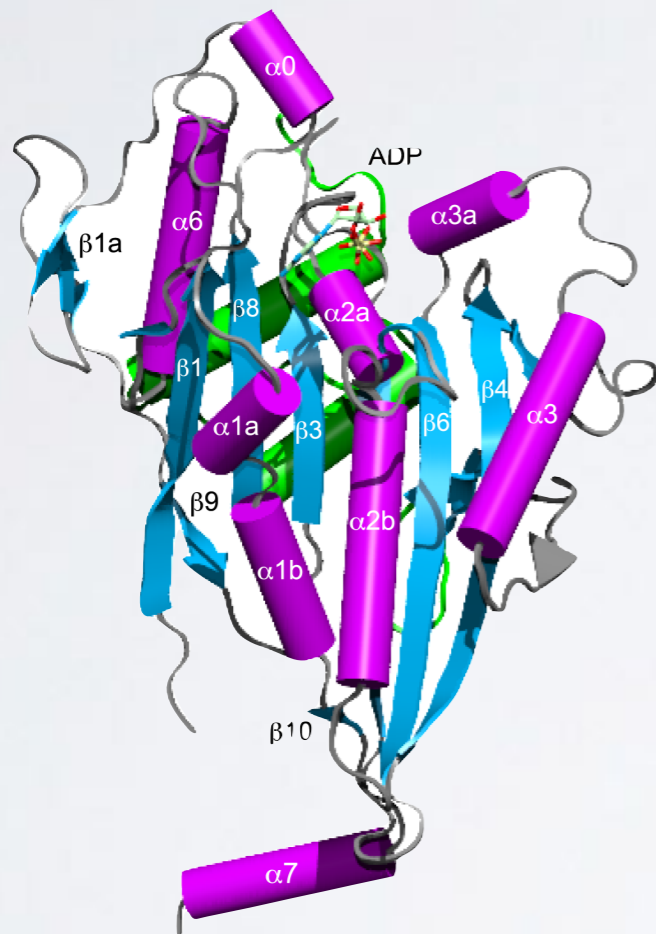
- ▶ **Representing and interpreting protein structure**

- Modeling energy as a function of structure

- ▶ **Example application areas**

- Predicting functional dynamics & drug discovery

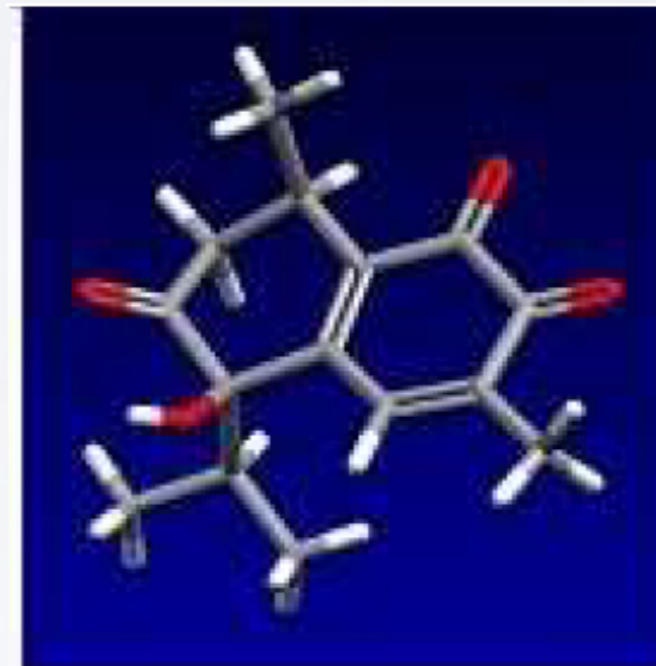
TRADITIONAL FOCUS **PROTEIN, DNA**  
AND **SMALL MOLECULE** DATA SETS  
WITH **MOLECULAR STRUCTURE**



Protein  
(PDB)



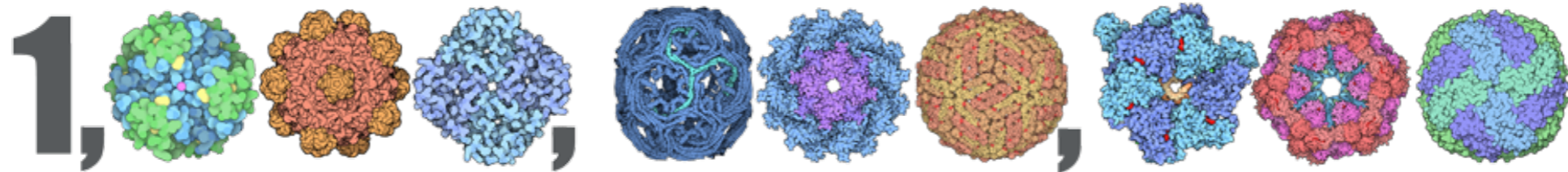
DNA  
(NDB)



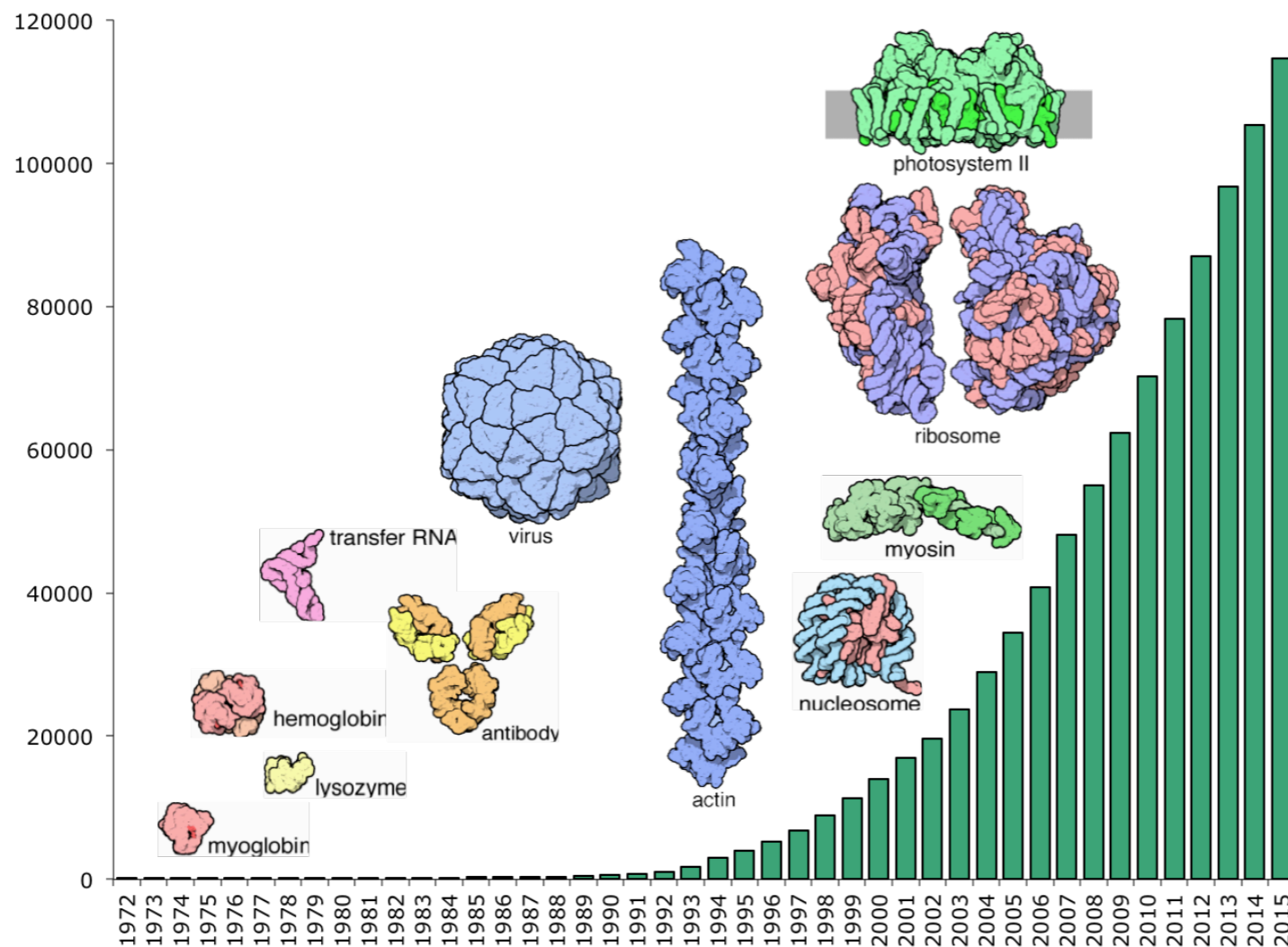
Small Molecules  
(CCDB)



# PDB – A Billion Atom Archive

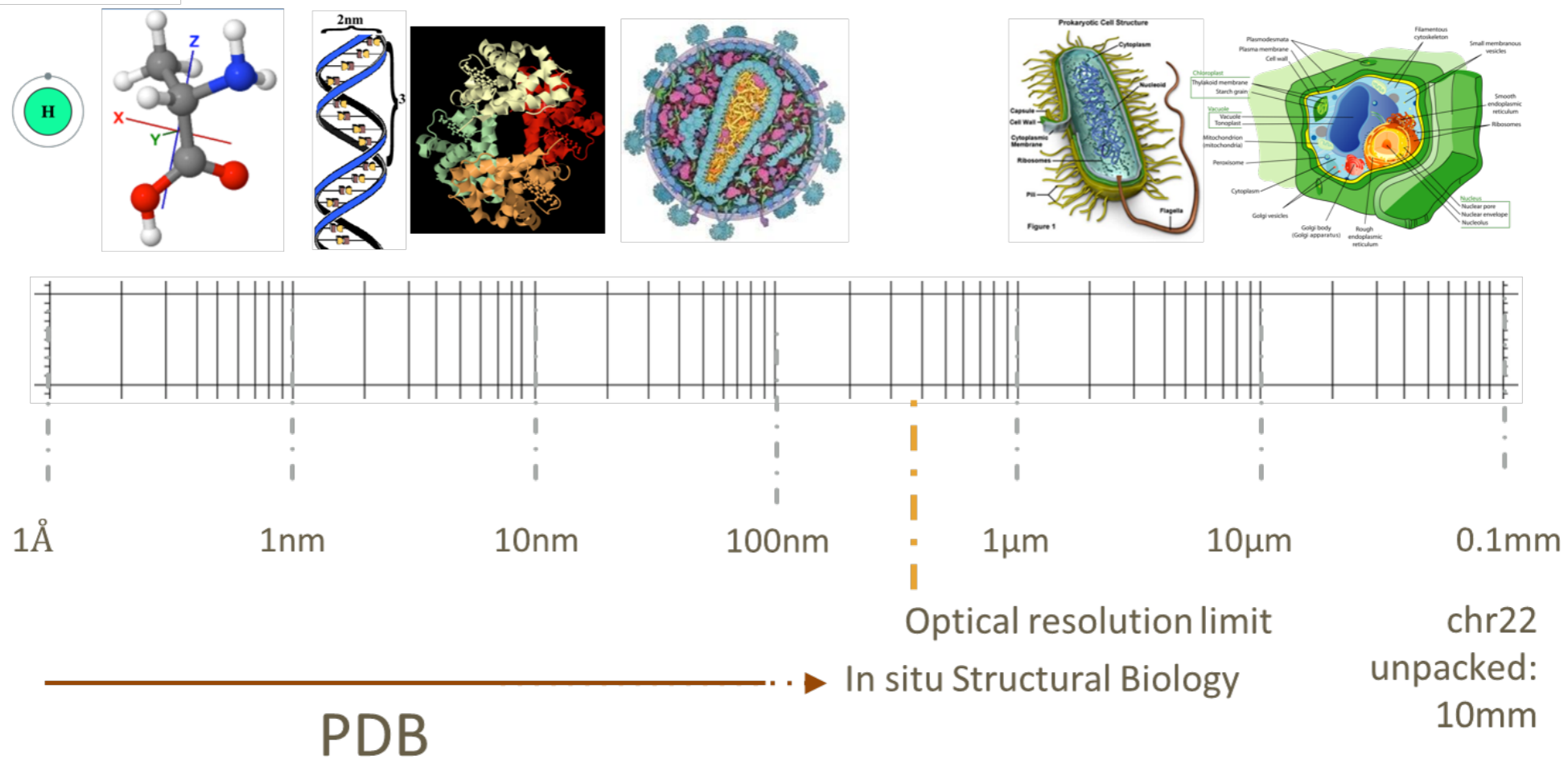


*> 1 billion atoms in the asymmetric units*

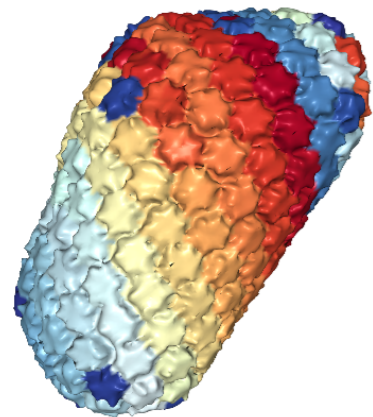


*~140,000  
Structures  
in May 2018*

# Growing Structure Size and Complexity

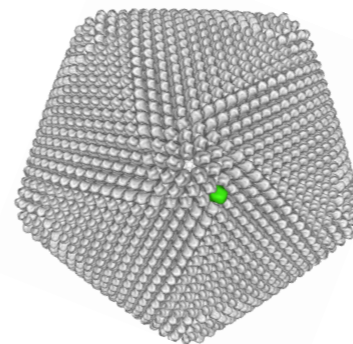


Largest asymmetric structure in PDB



HIV-1 capsid: PDB ID 3J3Q  
~2.4M unique atoms

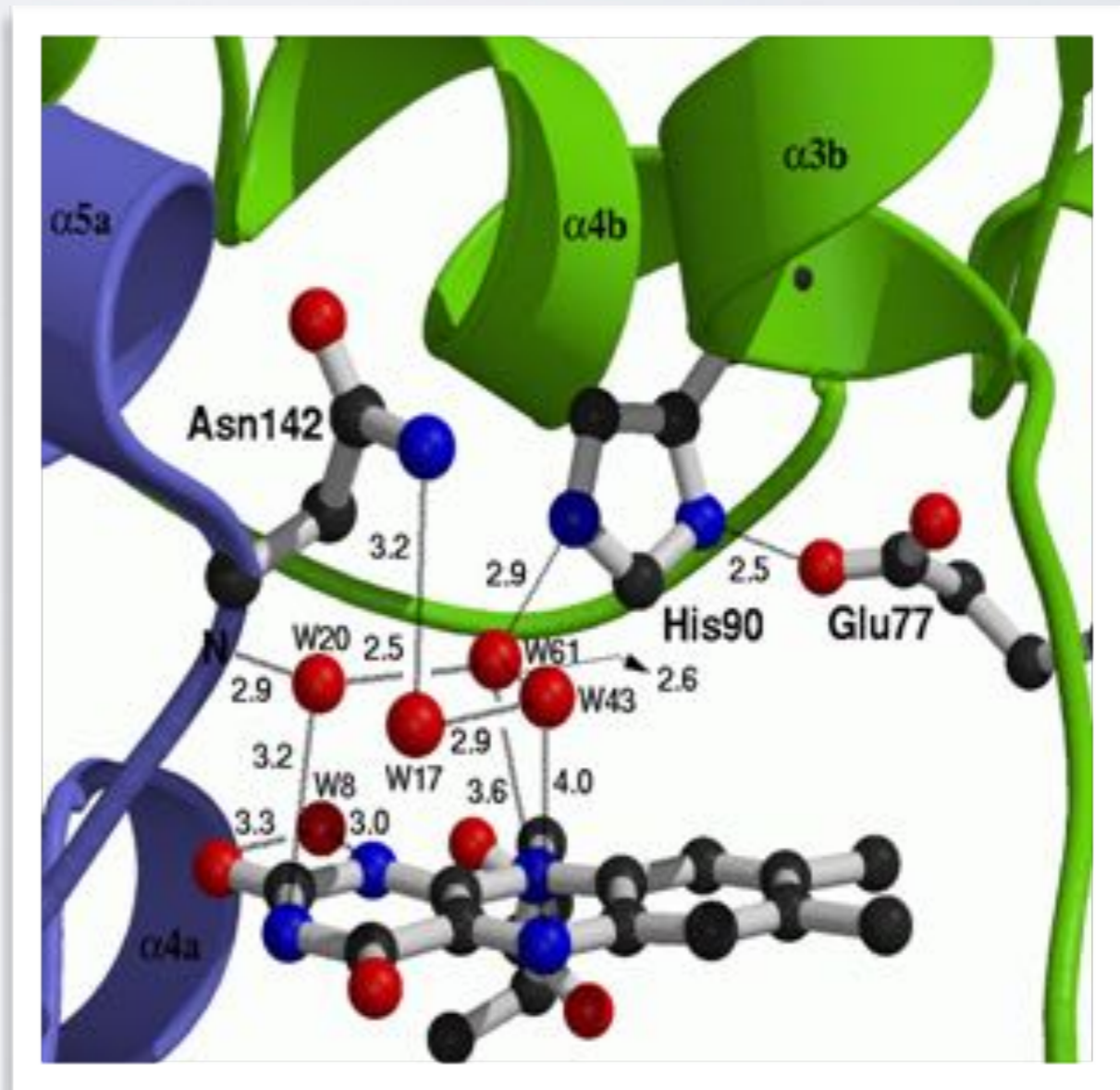
Largest symmetric structure in PDB



Faustovirus major capsid: PDB ID 5J7V  
~40M overall atoms

**Motivation 1:**  
Detailed understanding of  
molecular interactions

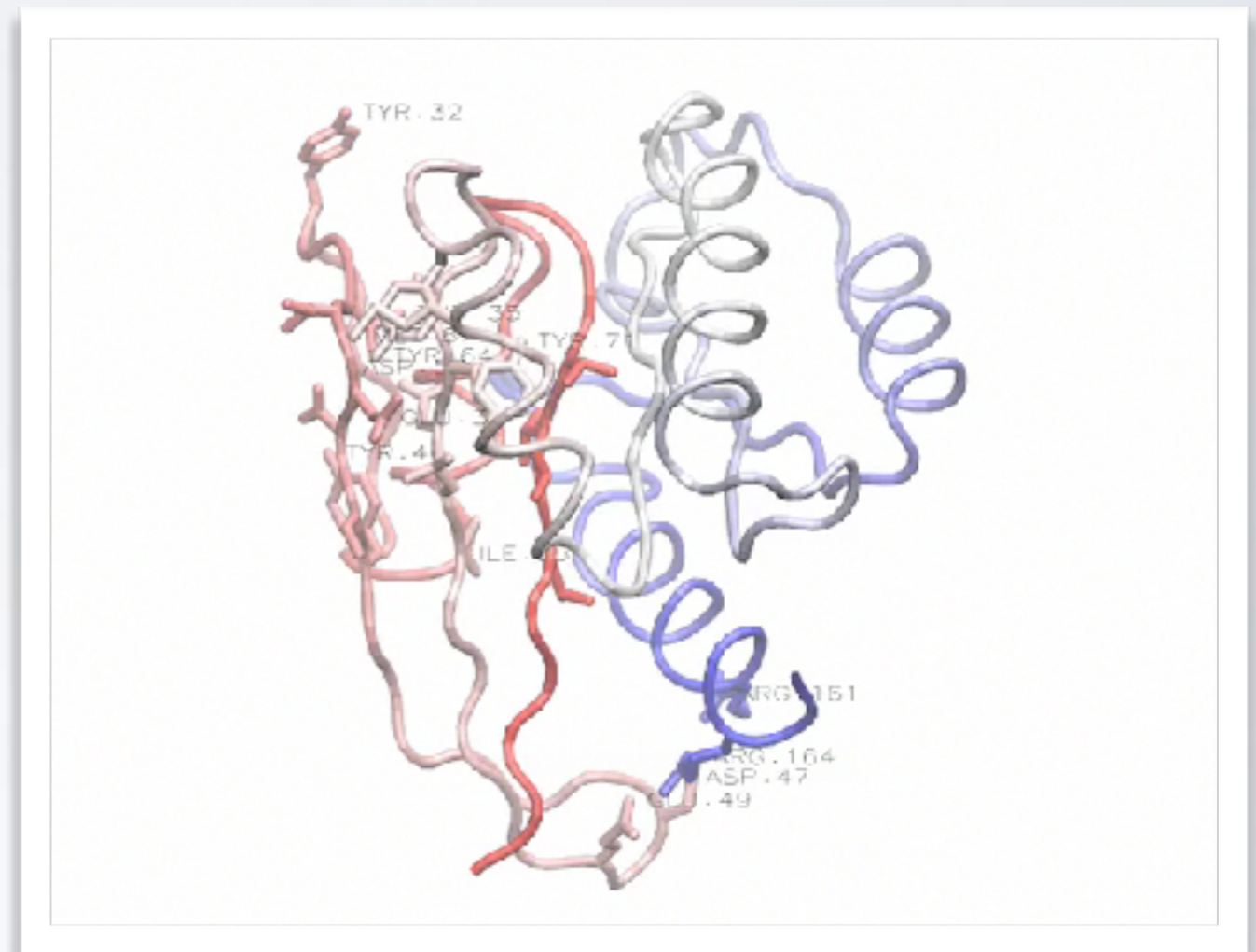
Provides an invaluable structural  
context for conservation and  
mechanistic analysis leading to  
functional insight.



## Motivation 1:

### Detailed understanding of molecular interactions

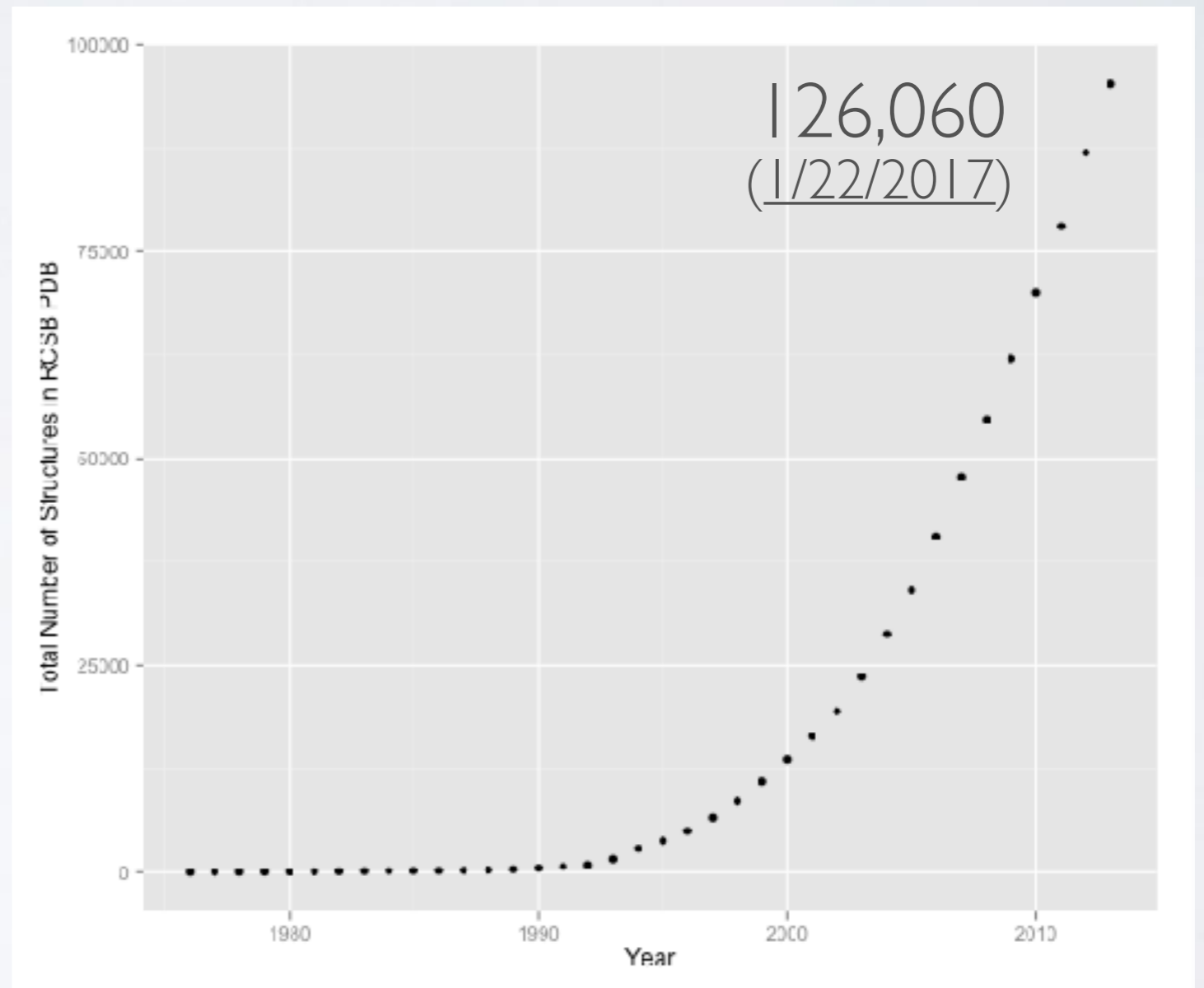
Computational modeling can provide detailed insight into functional interactions, their regulation and potential consequences of perturbation.



## Motivation 2:

Lots of structural data is becoming available

Structural Genomics has contributed to driving down the cost and time required for structural determination



Data from: <http://www.rcsb.org/pdb/statistics/>

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Structural Genomics has contributed to driving down the cost and time required for structural determination

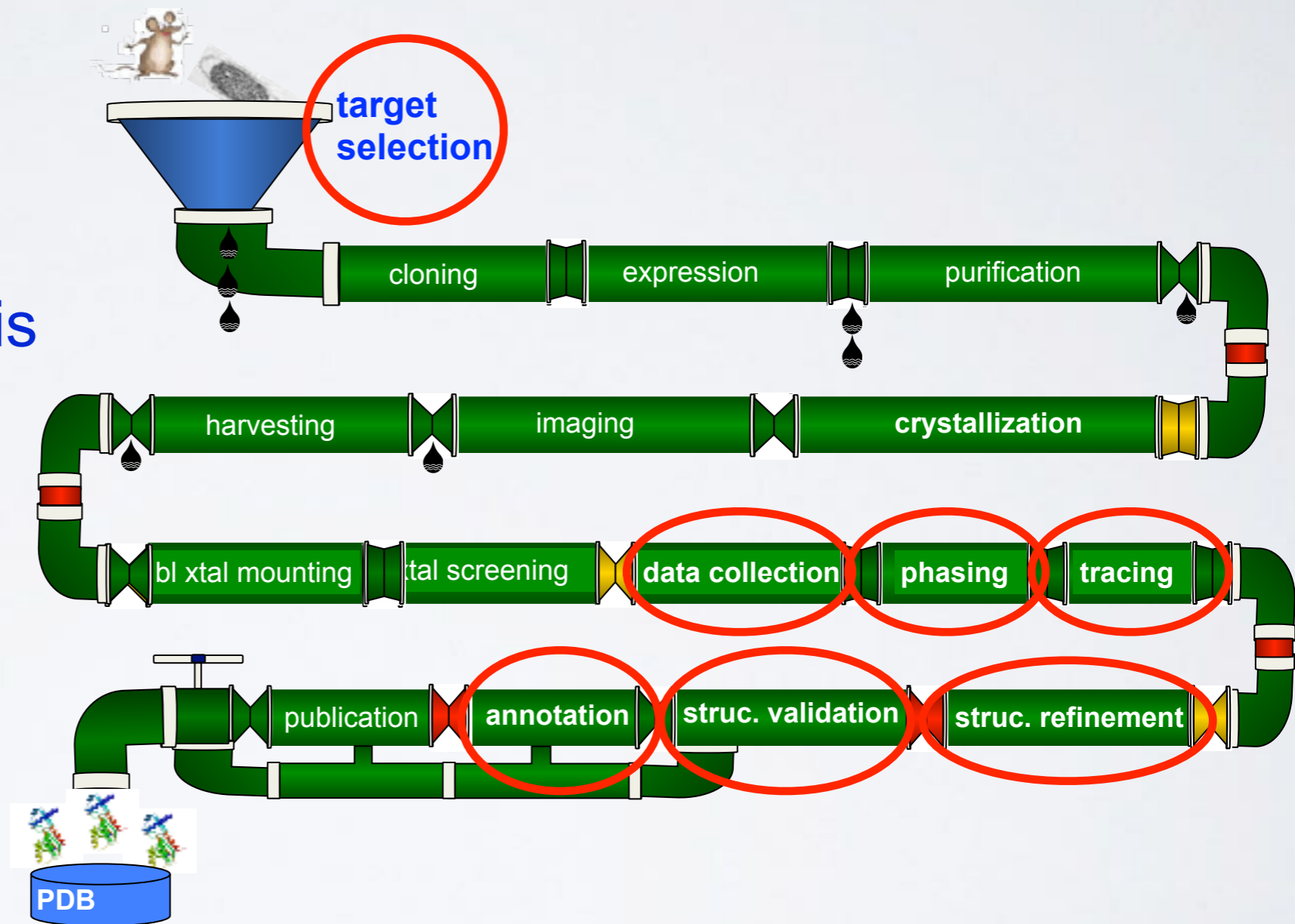
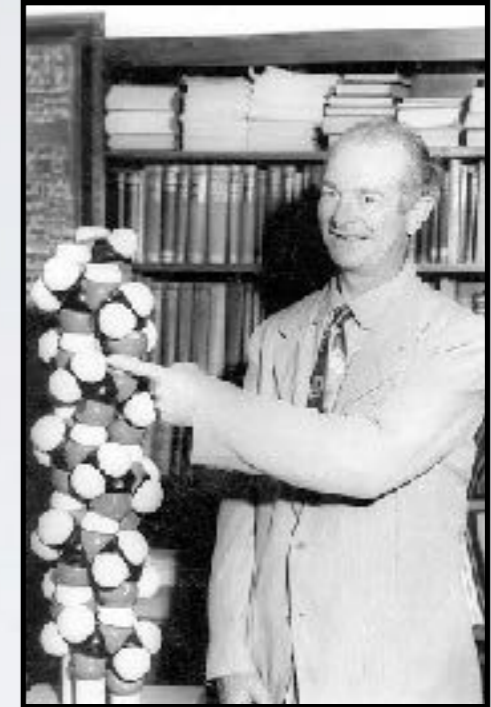
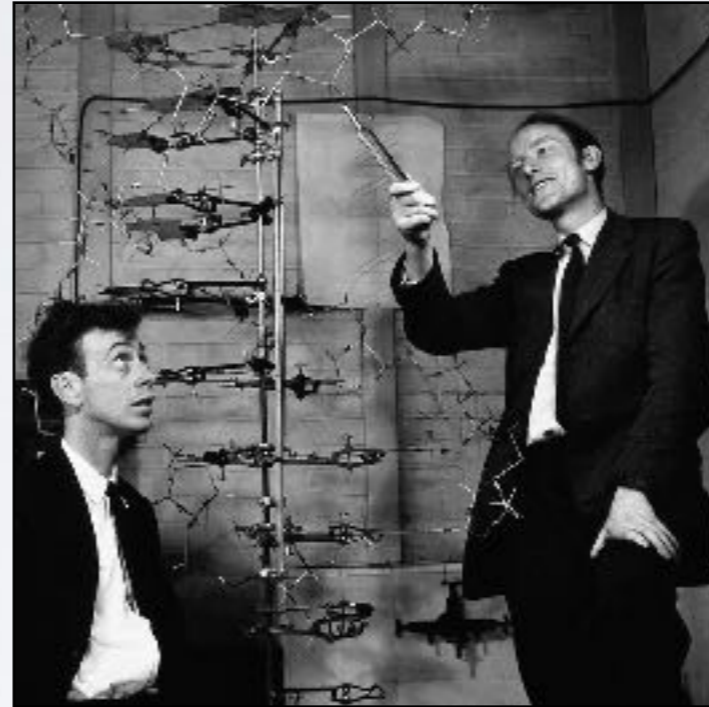


Image Credit: "Structure determination assembly line" Adam Godzik



### **Motivation 3:**

Theoretical and computational predictions have been, and continue to be, enormously valuable and influential!



# SUMMARY OF KEY **MOTIVATIONS**

## **Sequence > Structure > Function**

- Structure determines function, so understanding structure helps our understanding of function

## **Structure is more conserved than sequence**

- Structure allows identification of more distant evolutionary relationships

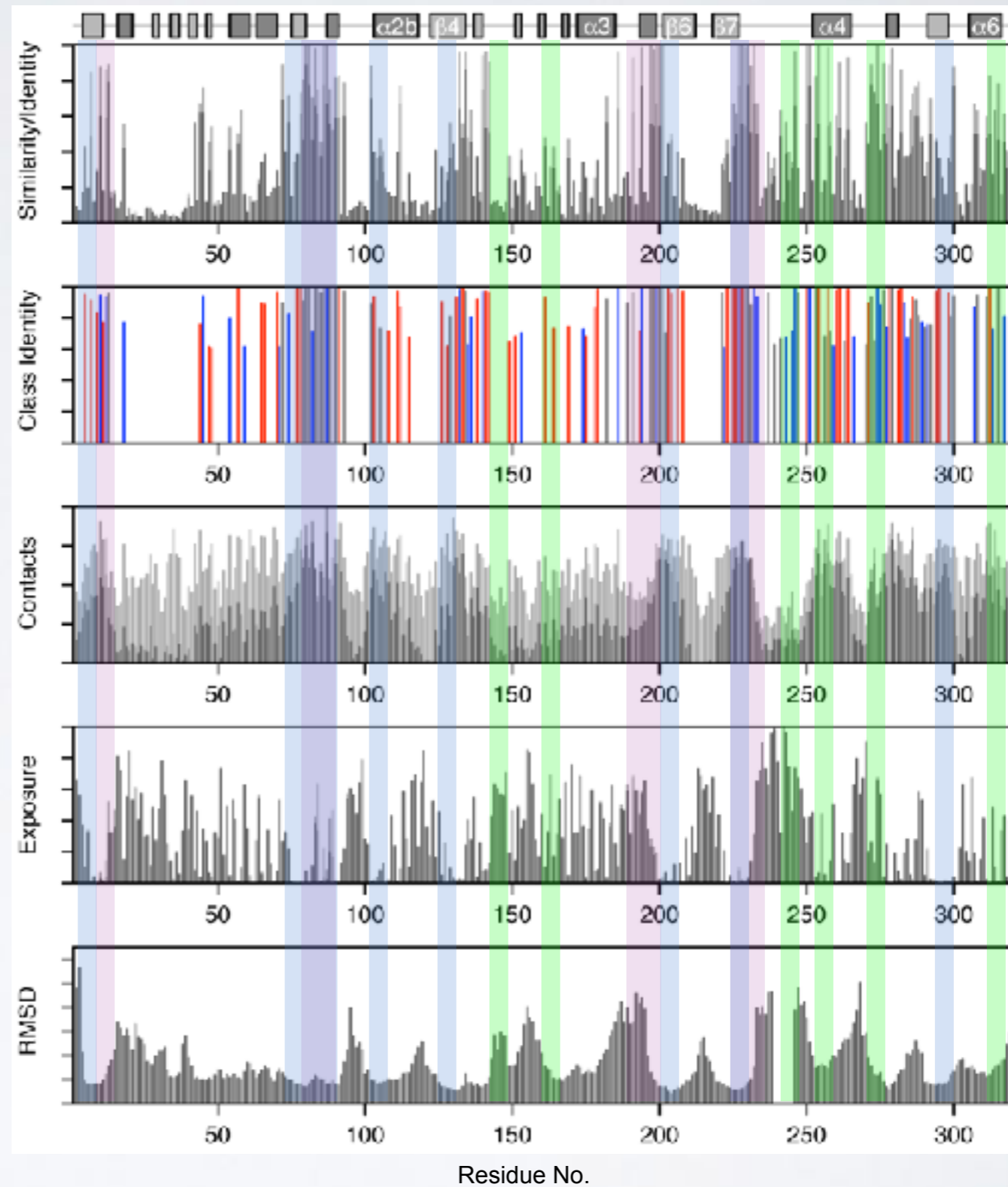
## **Structure is encoded in sequence**

- Understanding the determinants of structure allows design and manipulation of proteins for industrial and medical advantage



## Goals:

- Analysis
- Visualization
- Comparison
- Prediction
- Design



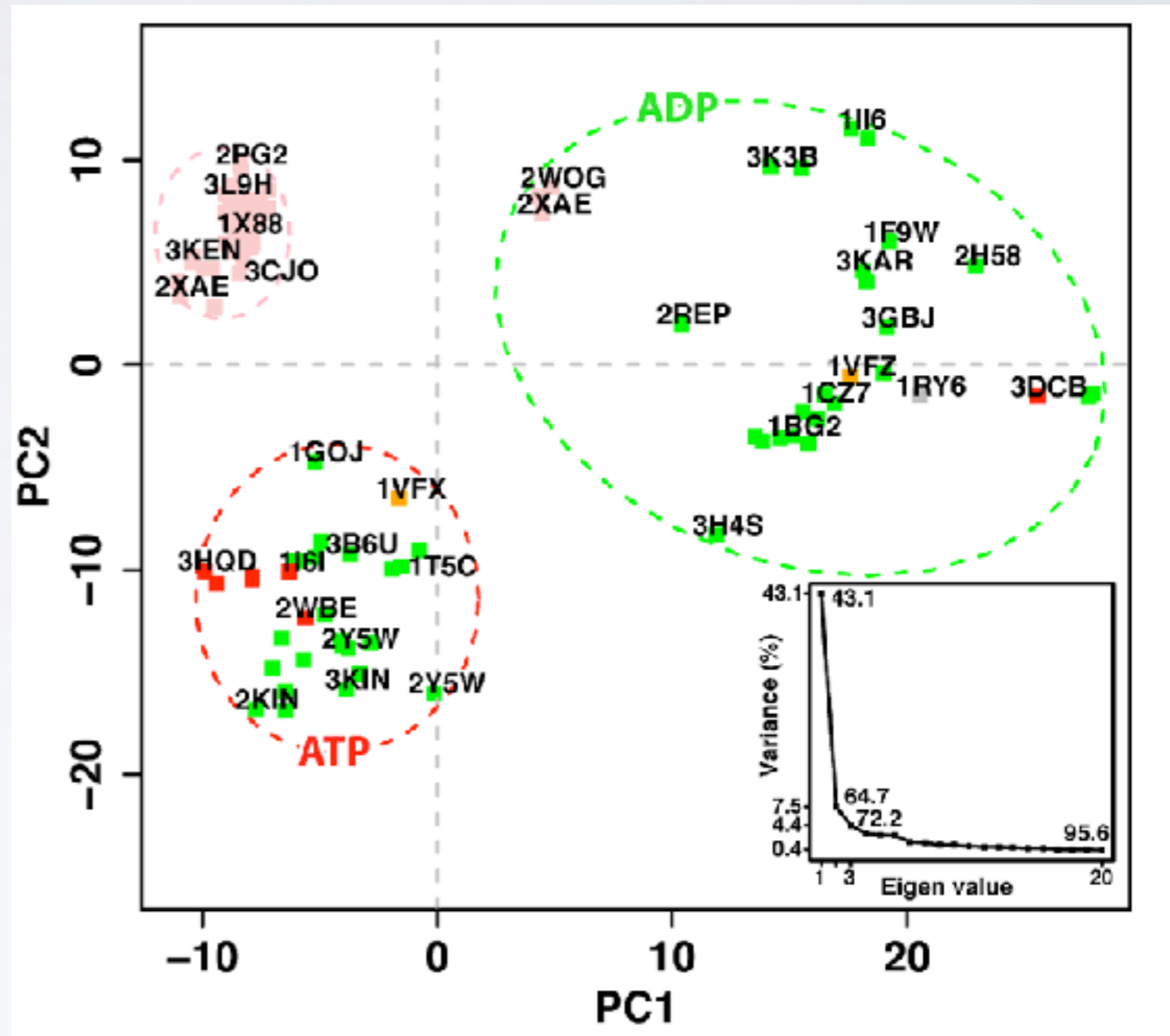
## Goals:

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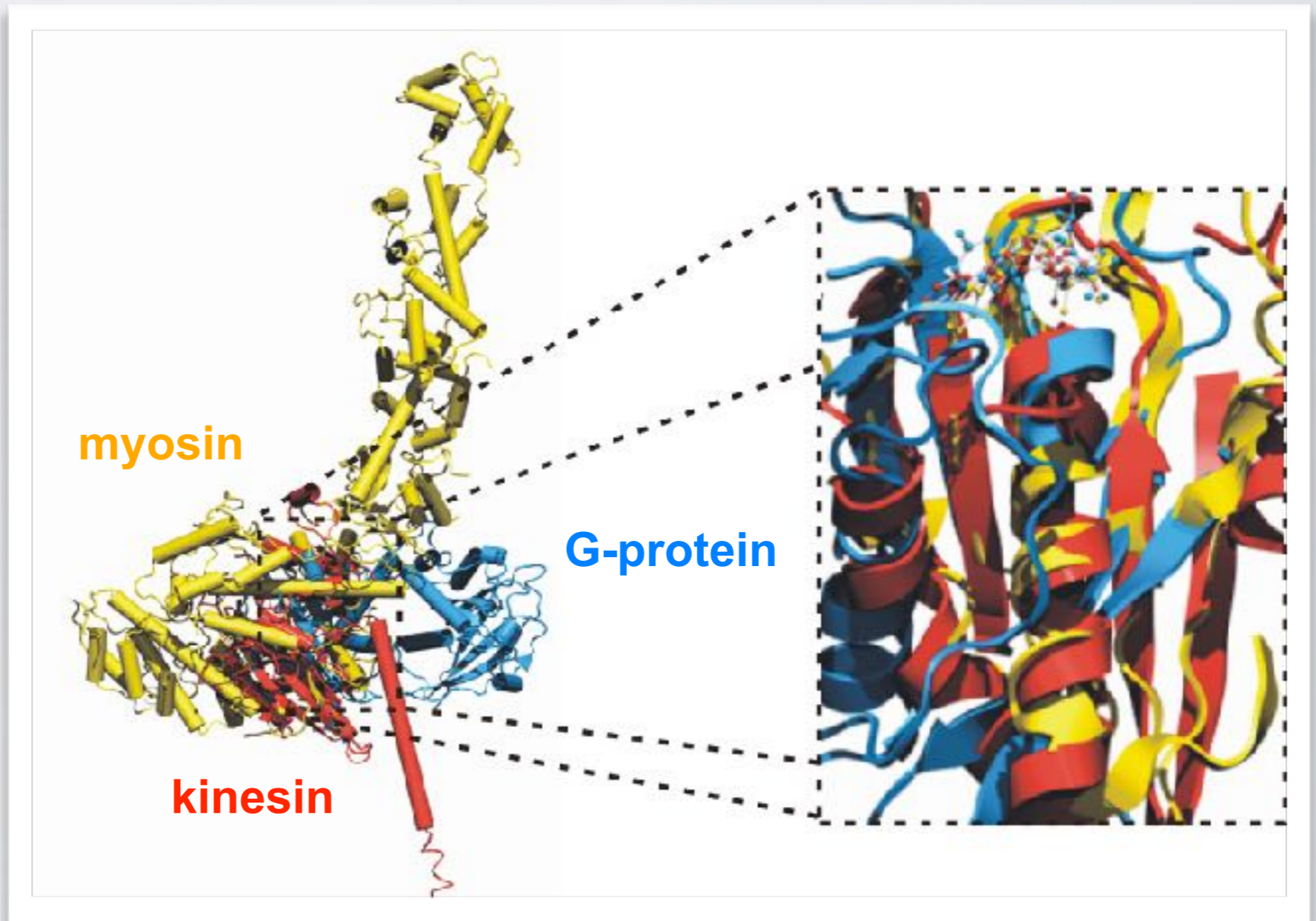
## Goals:

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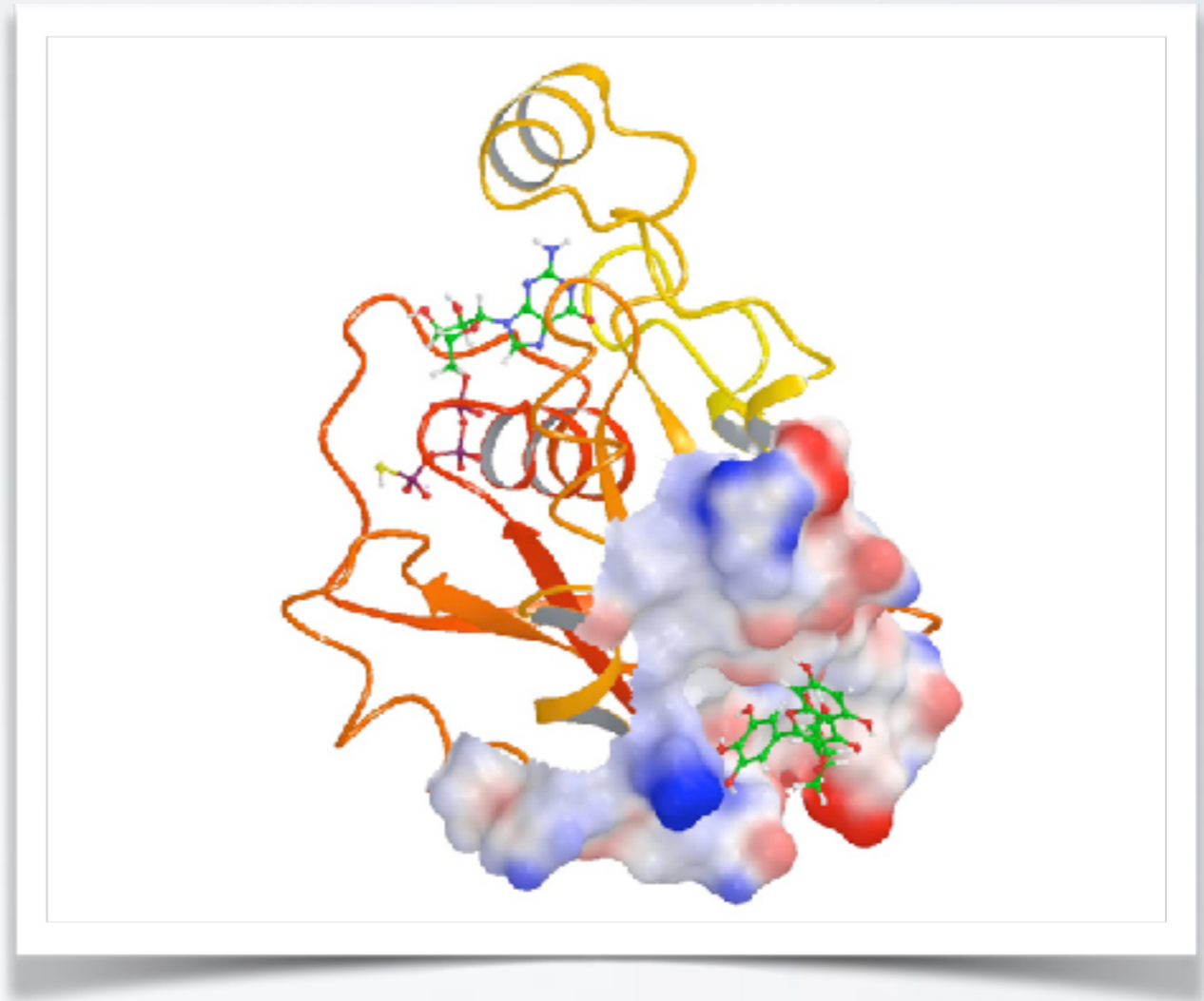
Goals:

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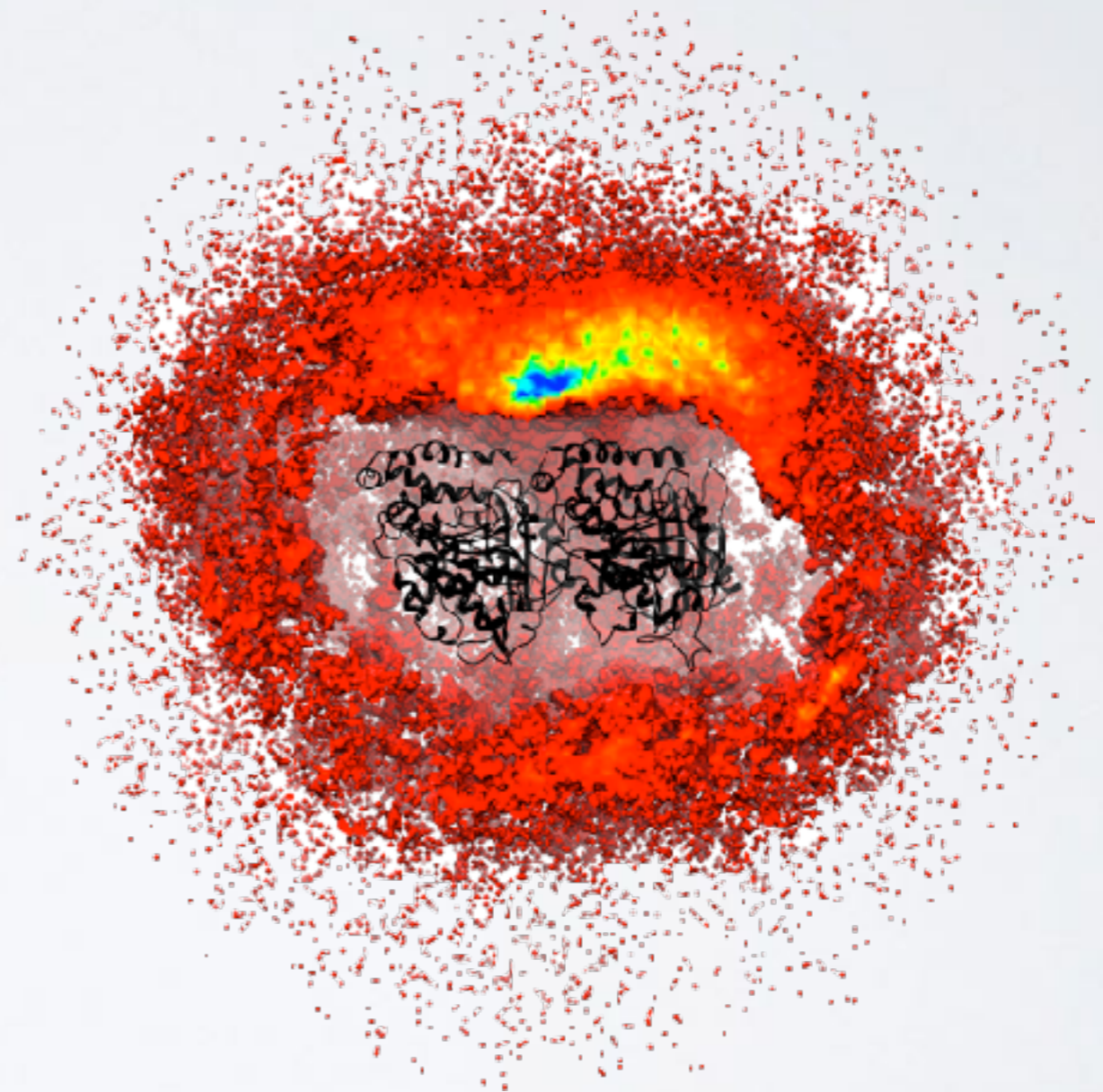
- Analysis
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Grant *et al.* PLoS One (2011, 2012)

## Goals:

- Analysis
- Visualization
- Comparison
- Prediction
- Design



Grant et al. PLoS Biology (2011)

# MAJOR RESEARCH AREAS AND CHALLENGES

Include but are not limited to:

- Protein classification
- Structure prediction from sequence
- Binding site detection
- Binding prediction and drug design
- Modeling molecular motions
- Predicting physical properties (stability, binding affinities)
- Design of structure and function
- etc...

With applications to Biology, Medicine, Agriculture and Industry

# NEXT UP:

- ▶ **Overview of structural bioinformatics**
  - Major motivations, goals and challenges

- ▶ **Fundamentals of protein structure**
  - Composition, form, forces and dynamics

- ▶ **Representing and interpreting protein structure**

- Modeling energy as a function of structure

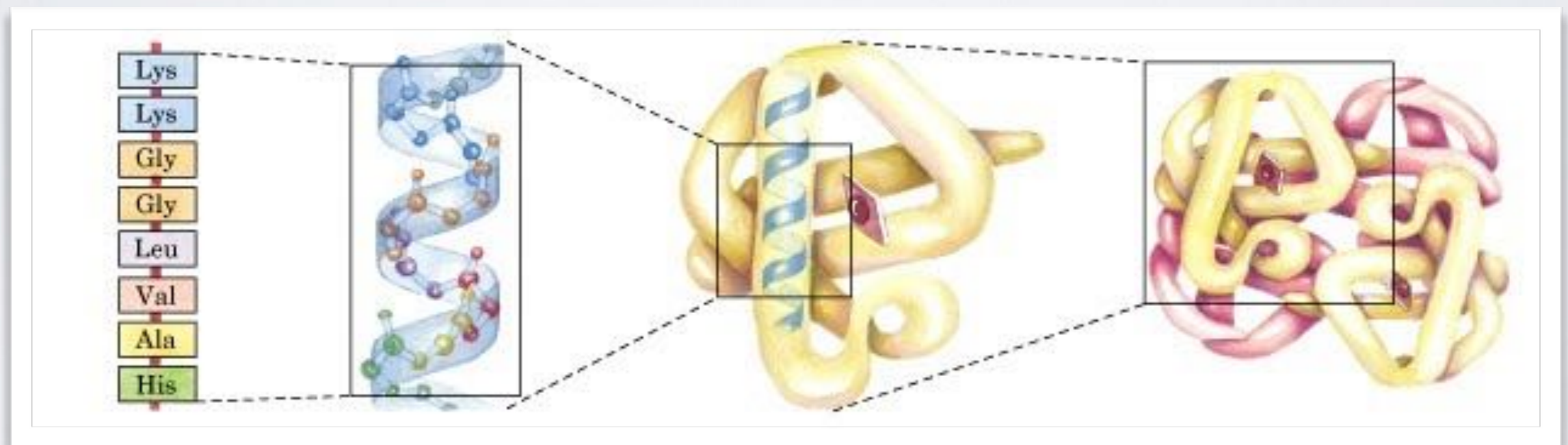
- ▶ **Example application areas**

- Predicting functional dynamics & drug discovery



# HIERARCHICAL STRUCTURE OF PROTEINS

Primary > Secondary > Tertiary > Quaternary



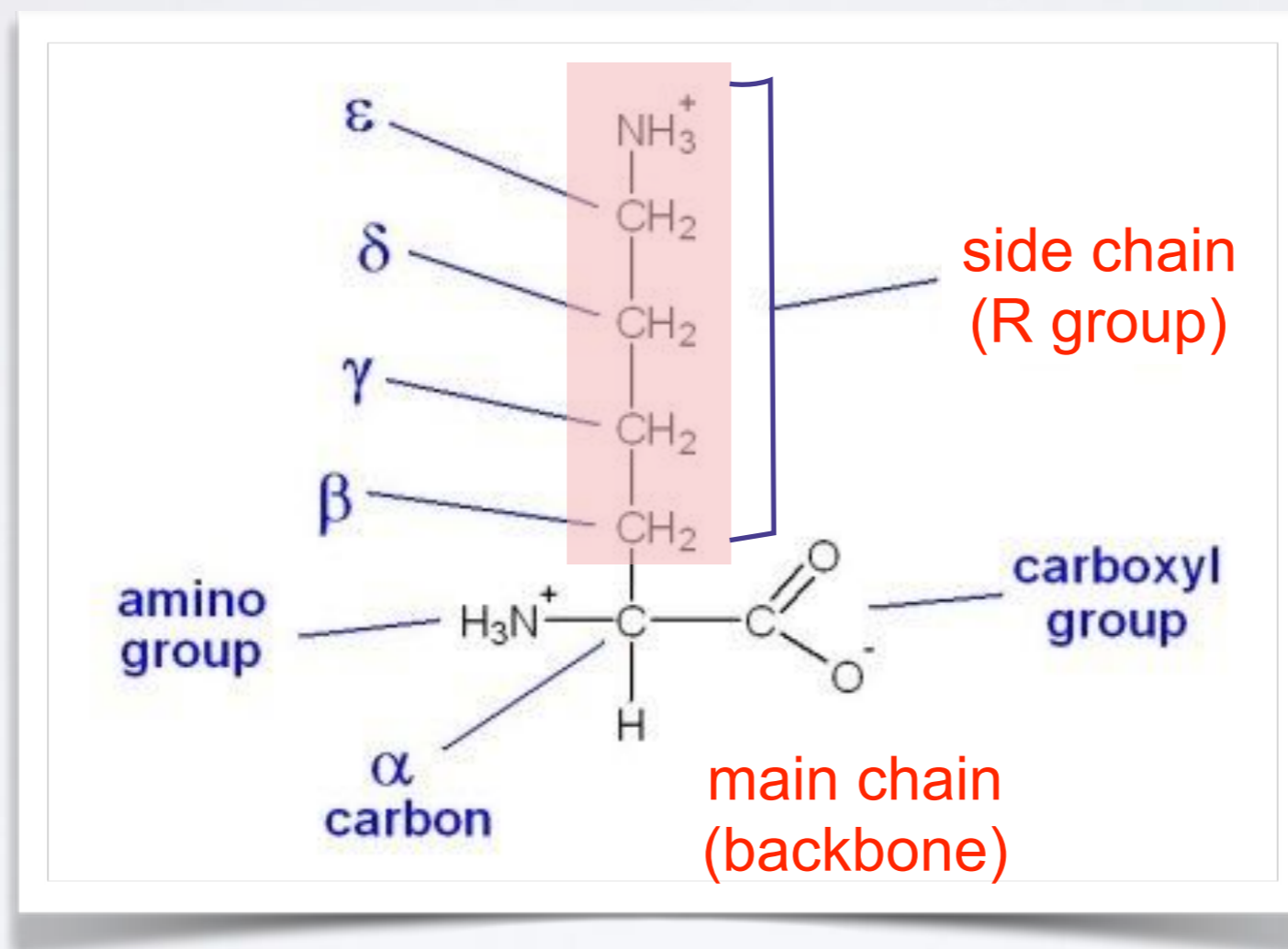
amino acid  
residues

Alpha  
helix

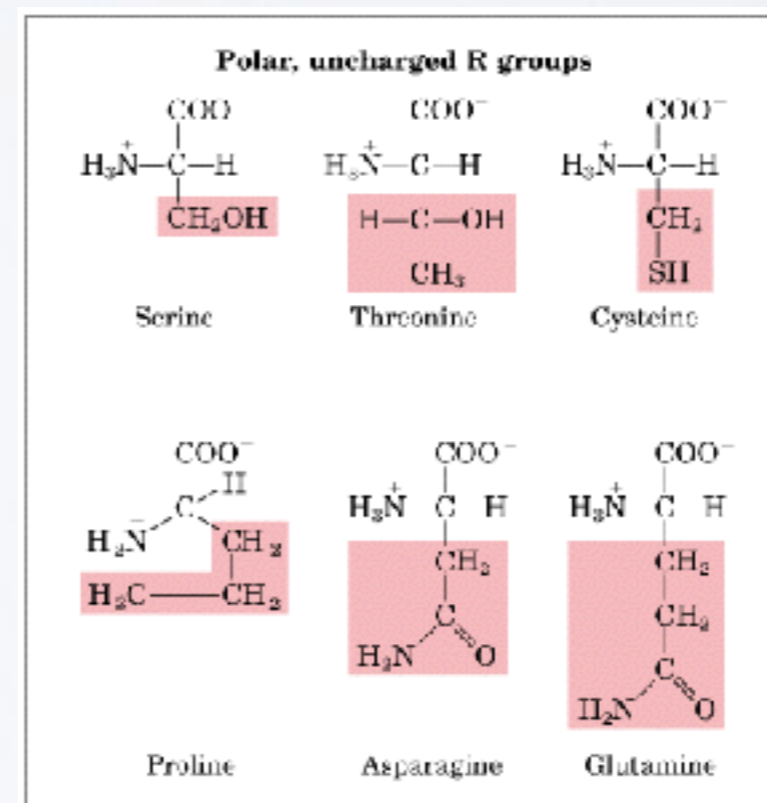
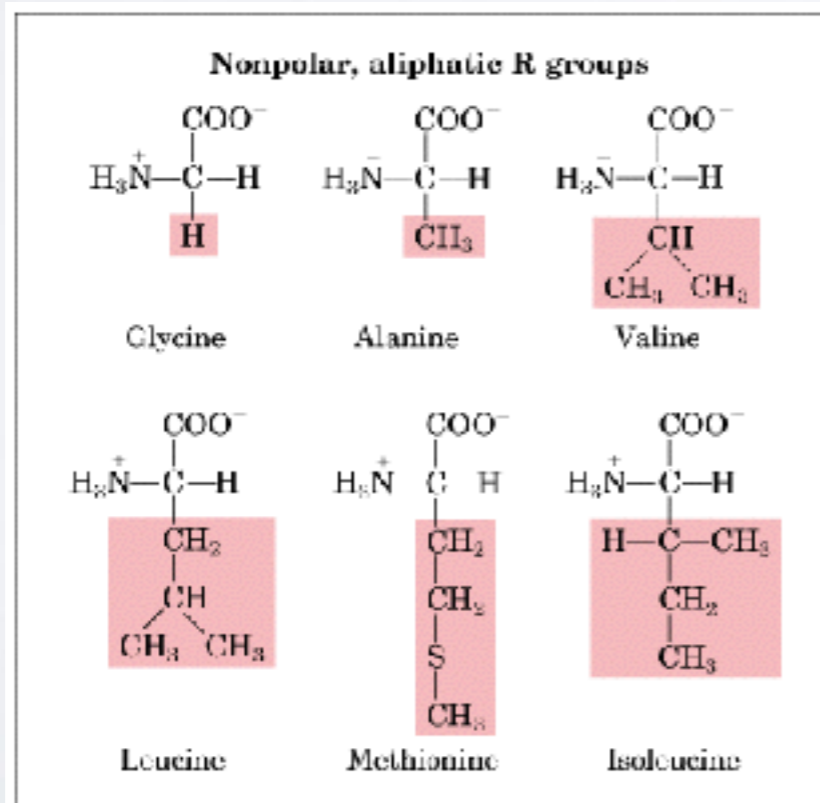
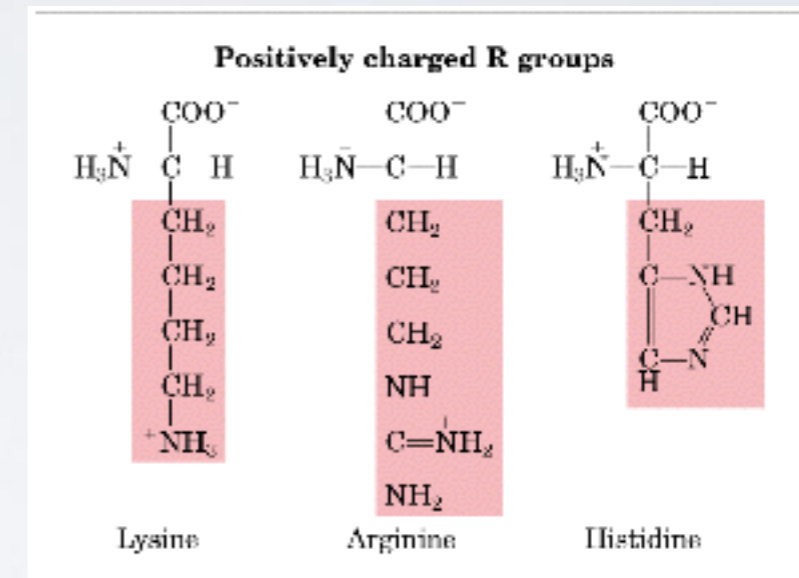
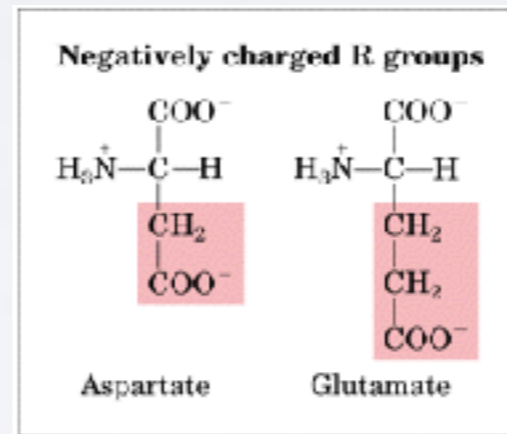
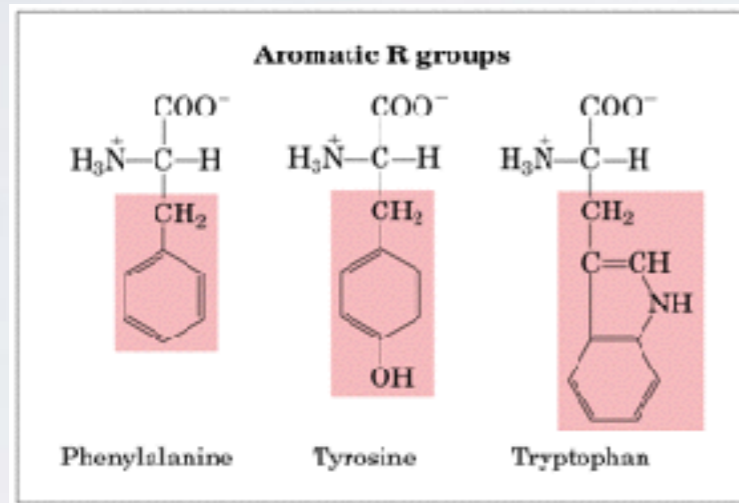
Polypeptide  
chain

Assembled  
subunits

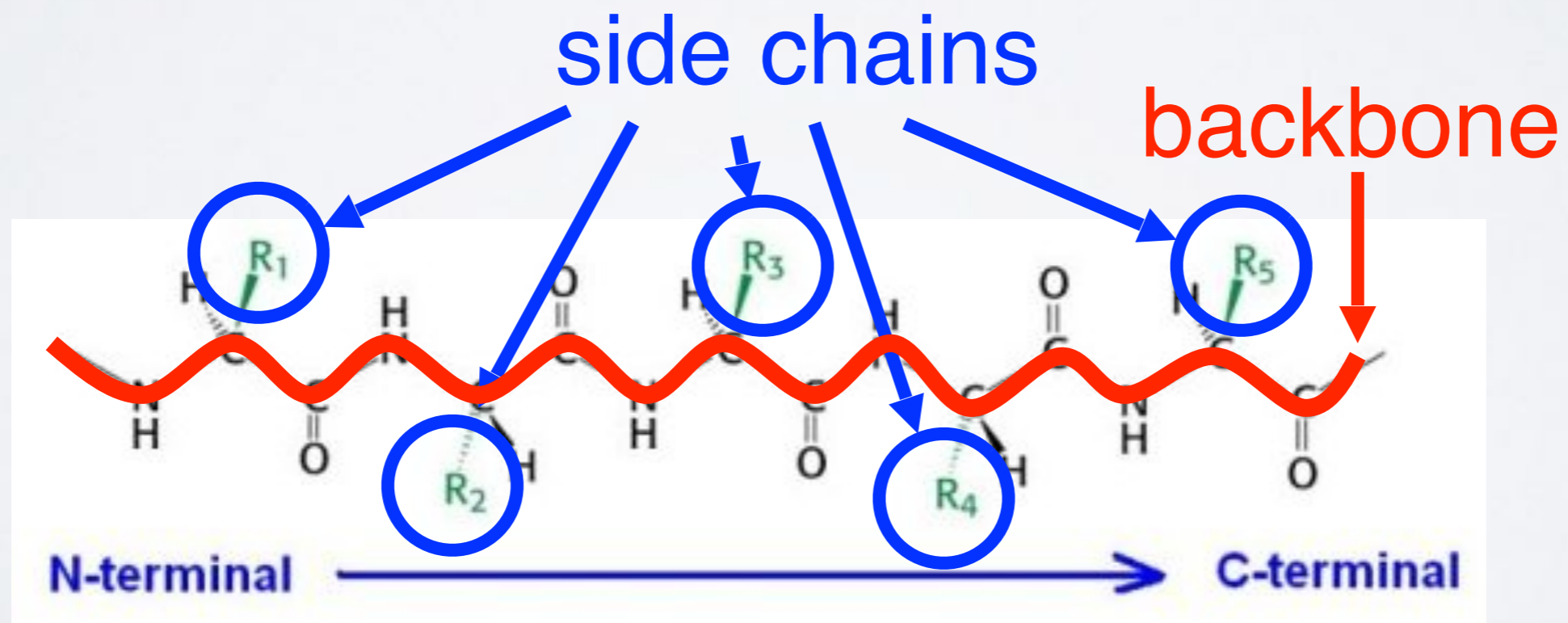
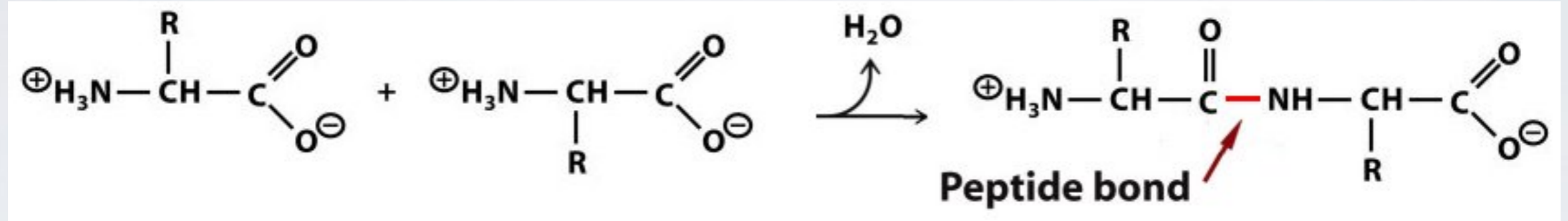
# RECAP: AMINO ACID NOMENCLATURE



# AMINO ACIDS CAN BE GROUPED BY THE PHYSIOCHEMICAL PROPERTIES



# AMINO ACIDS POLYMERIZE THROUGH **PEPTIDE BOND** FORMATION



# PEPTIDES CAN ADOPT DIFFERENT CONFORMATIONS BY VARYING THEIR **PHI & PSI BACKBONE TORSIONS**

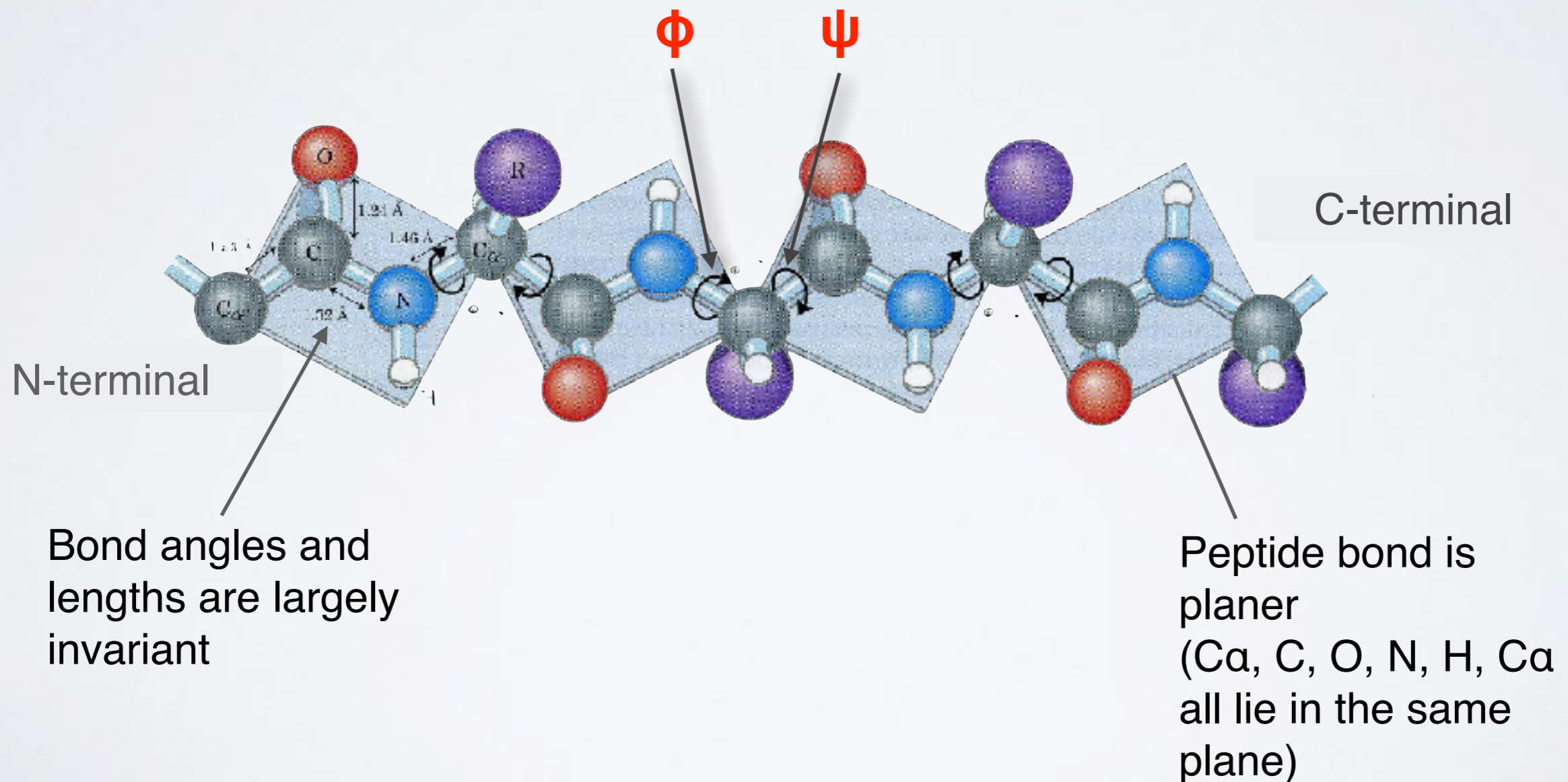
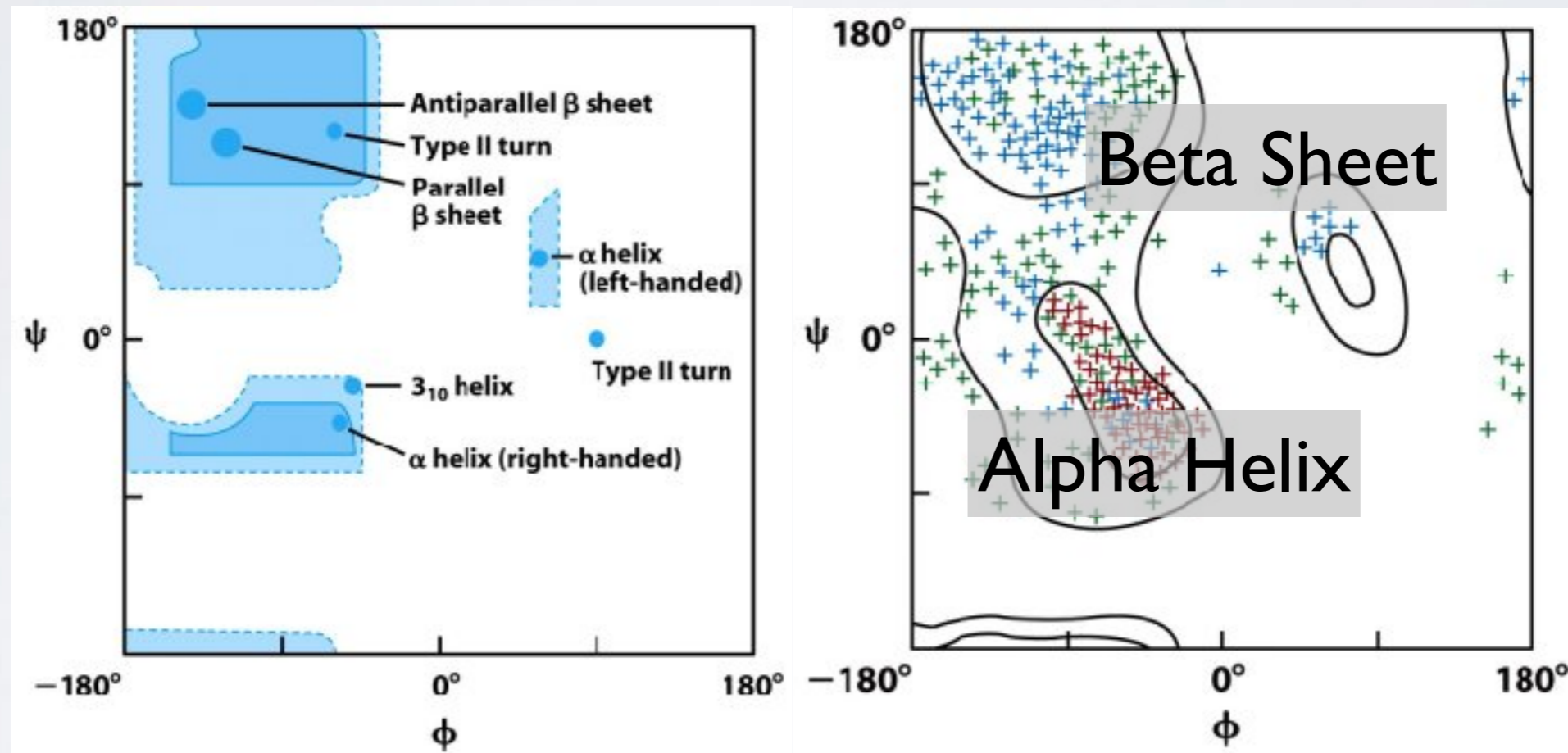


Image from: <http://www.ncbi.nlm.nih.gov/books/NBK21581/>

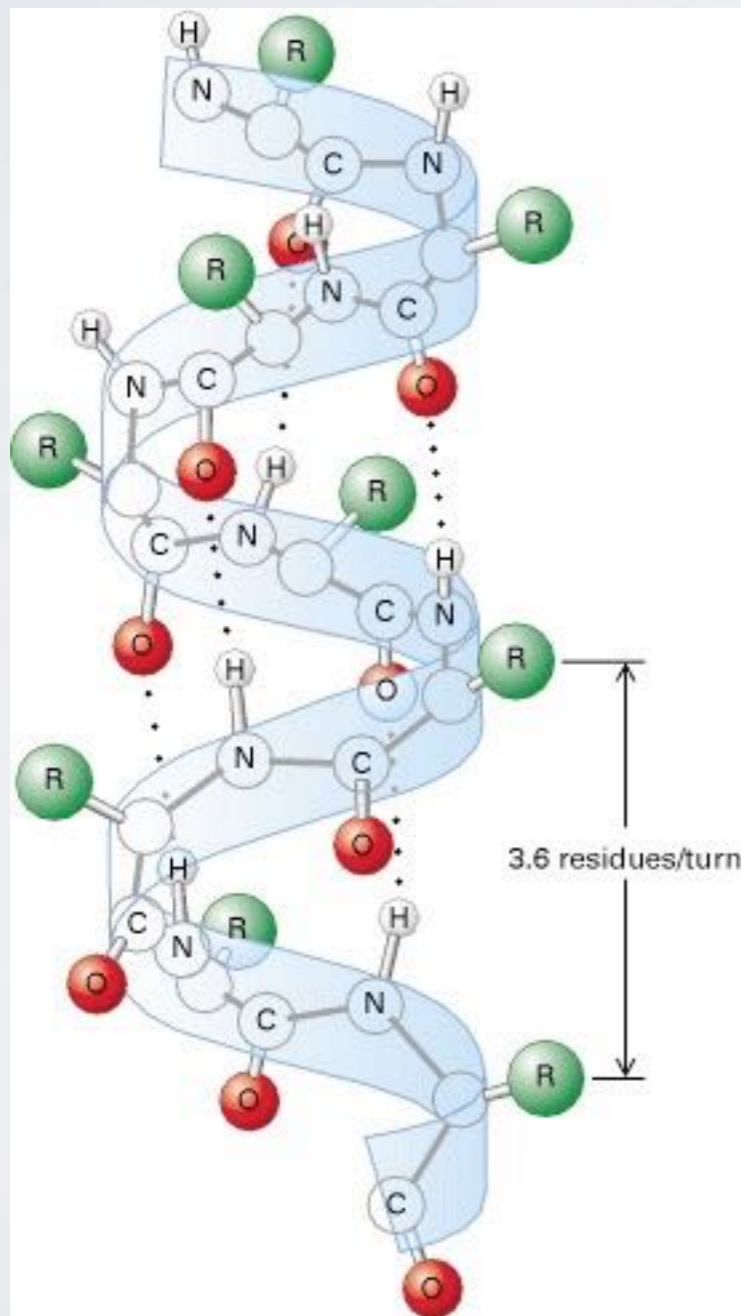
# PHI vs PSI PLOTS ARE KNOWN AS **RAMACHANDRAN DIAGRAMS**



- Steric hindrance dictates torsion angle preference
- Ramachandran plot show preferred regions of  $\phi$  and  $\psi$  dihedral angles which correspond to major forms of **secondary structure**

# MAJOR SECONDARY STRUCTURE TYPES

## **ALPHA HELIX** & BETA SHEET

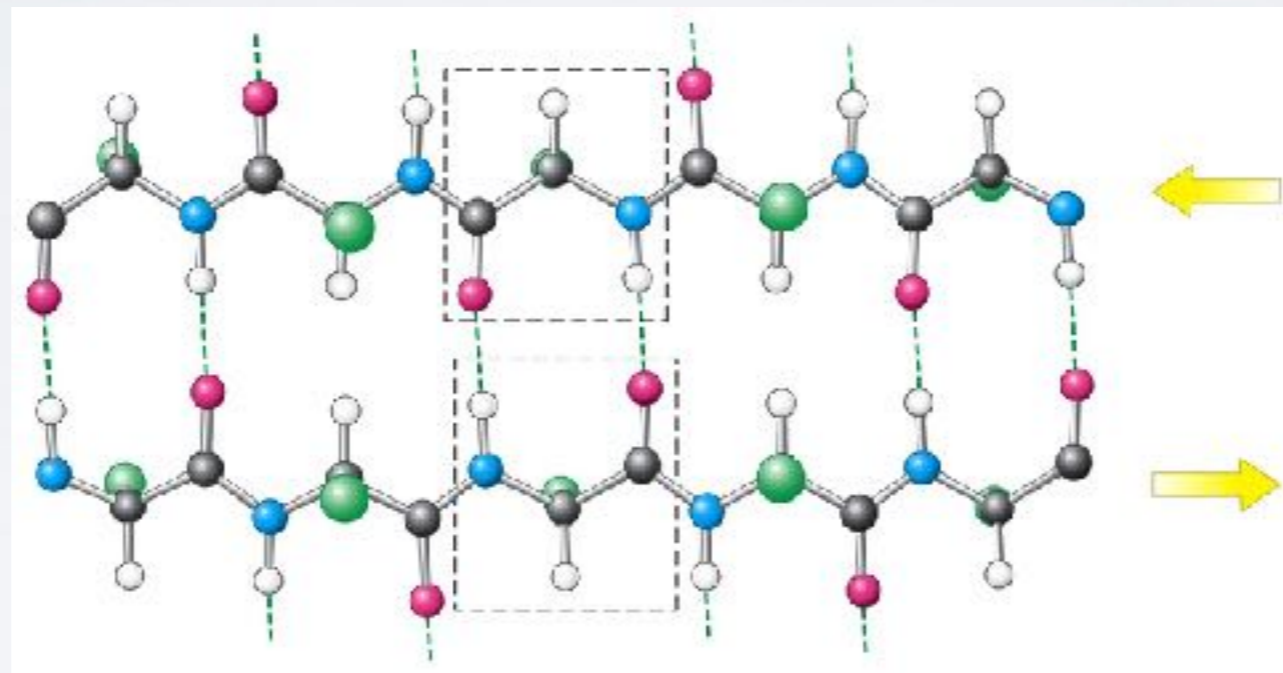


### **$\alpha$ -helix**

- Most common form has 3.6 residues per turn (number of residues in one full rotation)
- Hydrogen bonds (dashed lines) between residue  $i$  and  $i+4$  stabilize the structure
- The side chains (in green) protrude outward
- **$3_{10}$ -helix** and  **$\pi$ -helix** forms are less common

# MAJOR SECONDARY STRUCTURE TYPES

## ALPHA HELIX & **BETA SHEET**



### In antiparallel $\beta$ -sheets

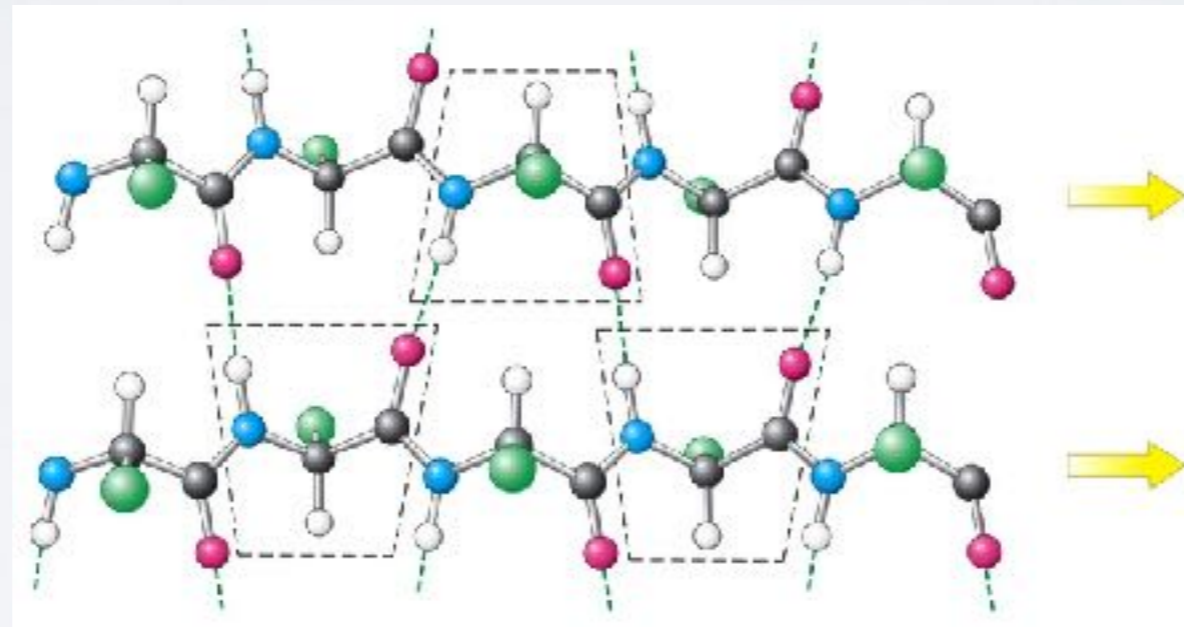
- Adjacent  $\beta$ -strands run in opposite directions
- Hydrogen bonds (dashed lines) between NH and CO stabilize the structure
- The side chains (in green) are above and below the sheet

Image from: <http://www.ncbi.nlm.nih.gov/books/NBK21581/>



# MAJOR SECONDARY STRUCTURE TYPES

## ALPHA HELIX & **BETA SHEET**

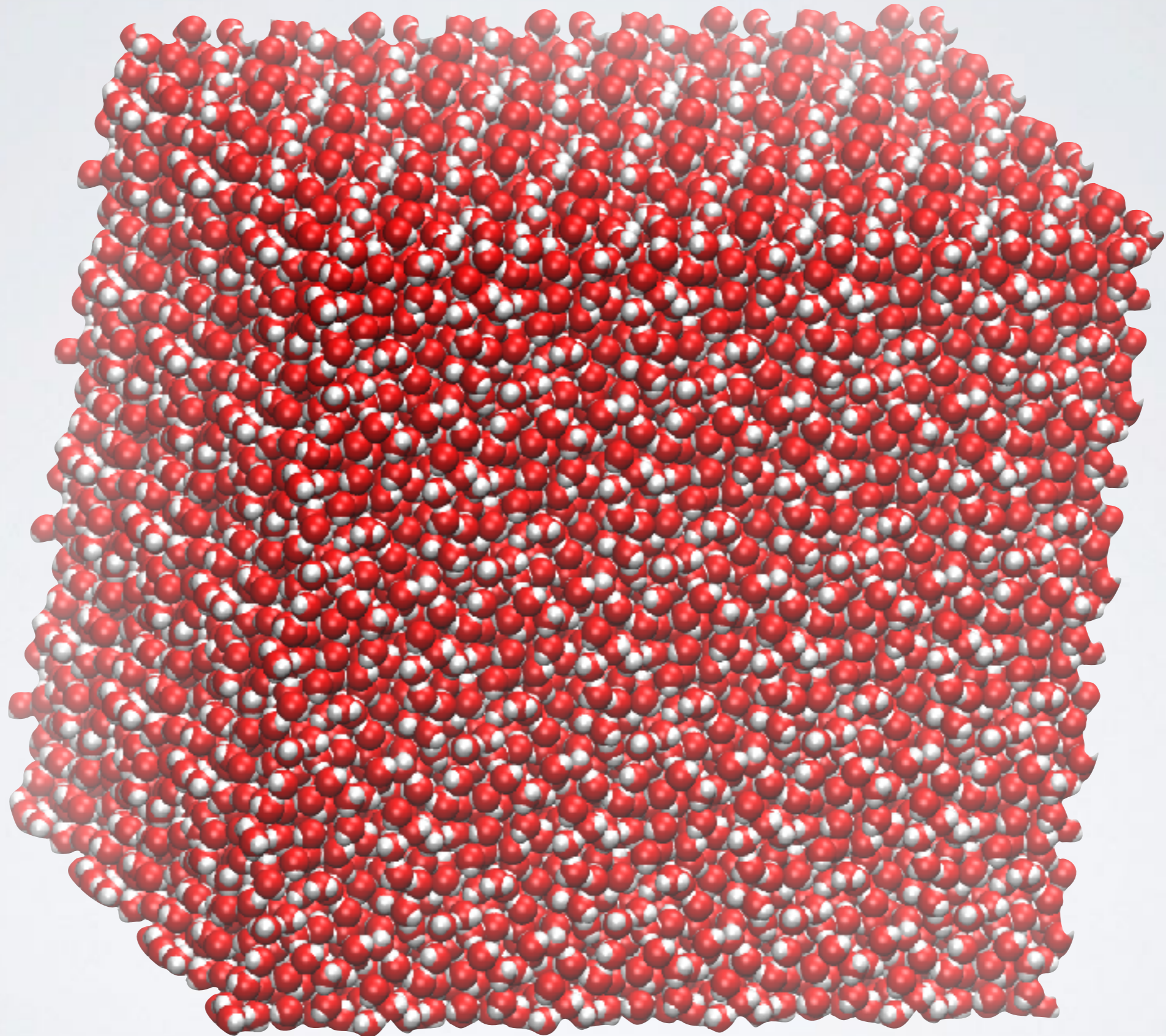


### In parallel $\beta$ -sheets

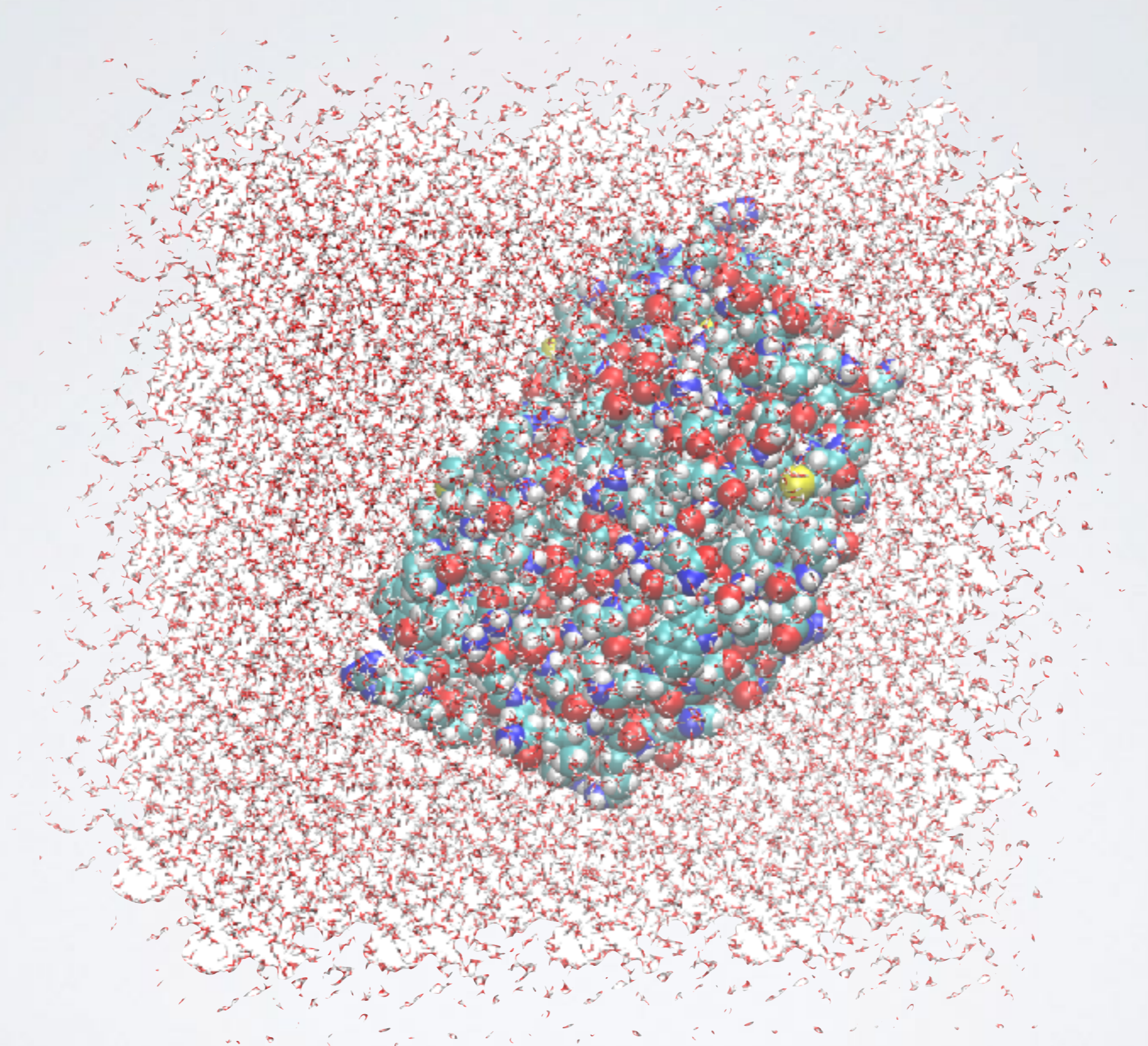
- Adjacent  $\beta$ -strands run in same direction
- Hydrogen bonds (dashed lines) between NH and CO stabilize the structure
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Image from: <http://www.ncbi.nlm.nih.gov/books/NBK21581/>

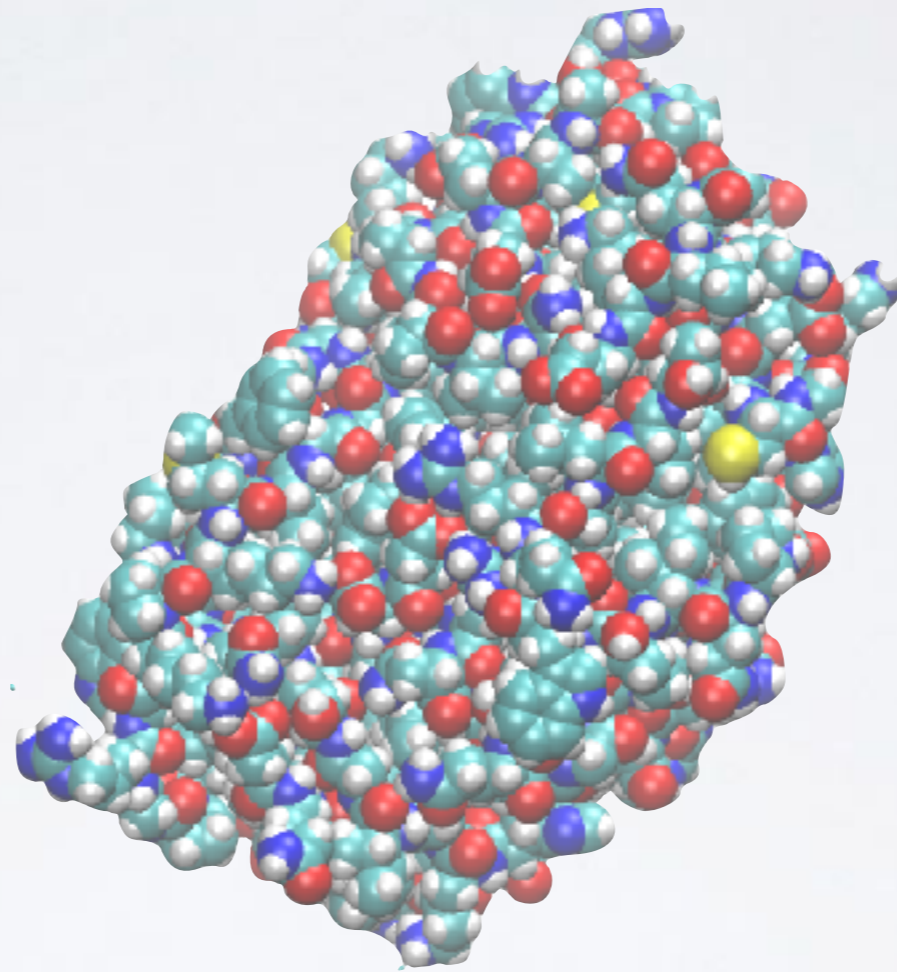
**What Does a Protein Look like?**



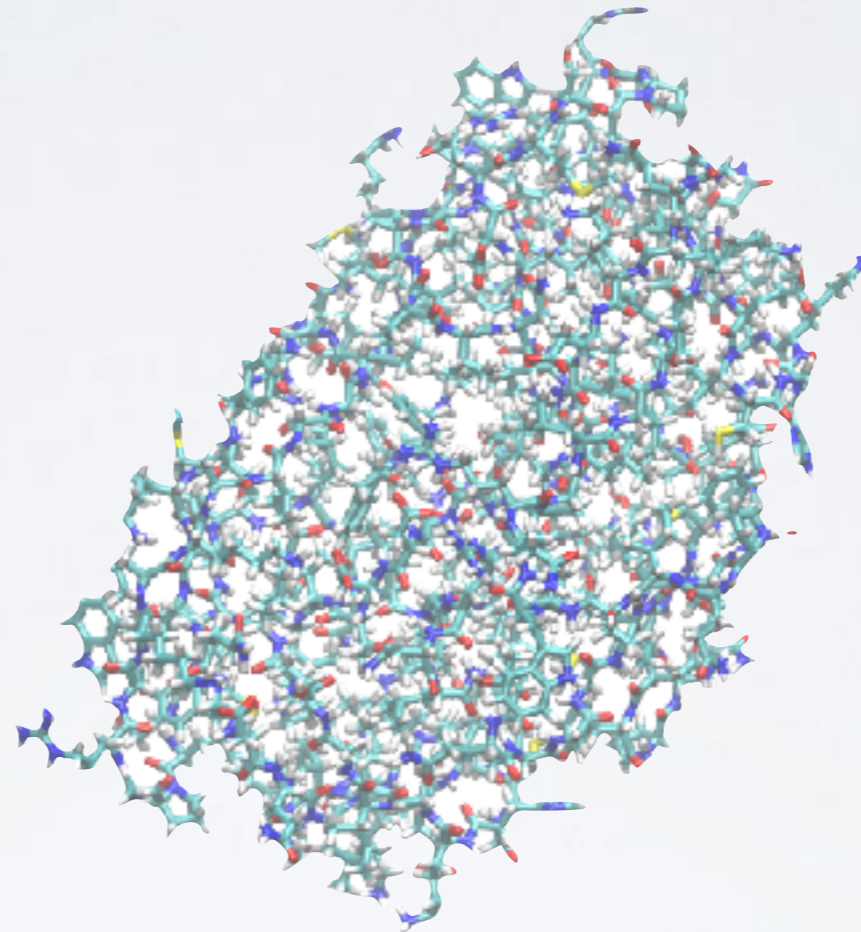
- Proteins are stable (and hidden) in water



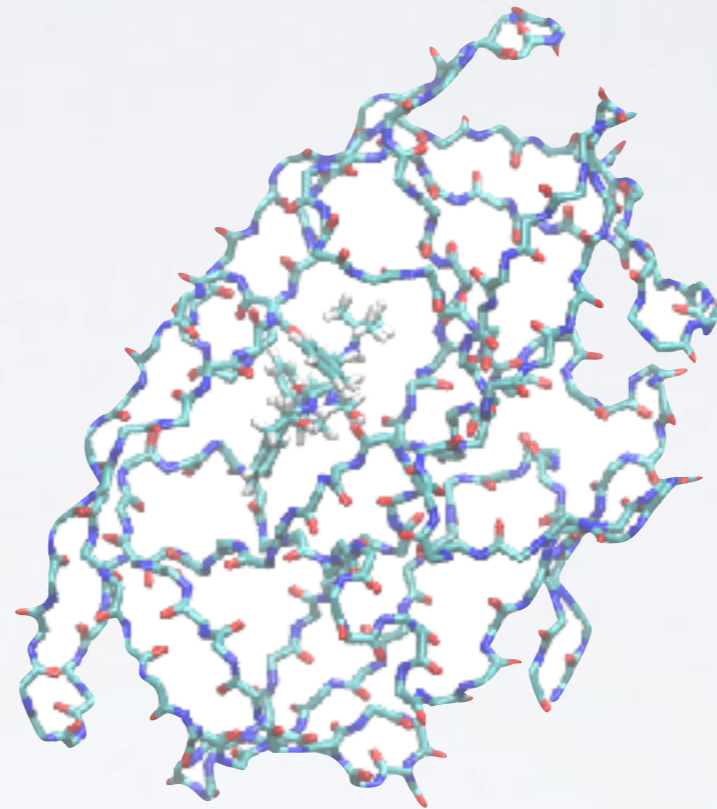
- Proteins closely interact with water



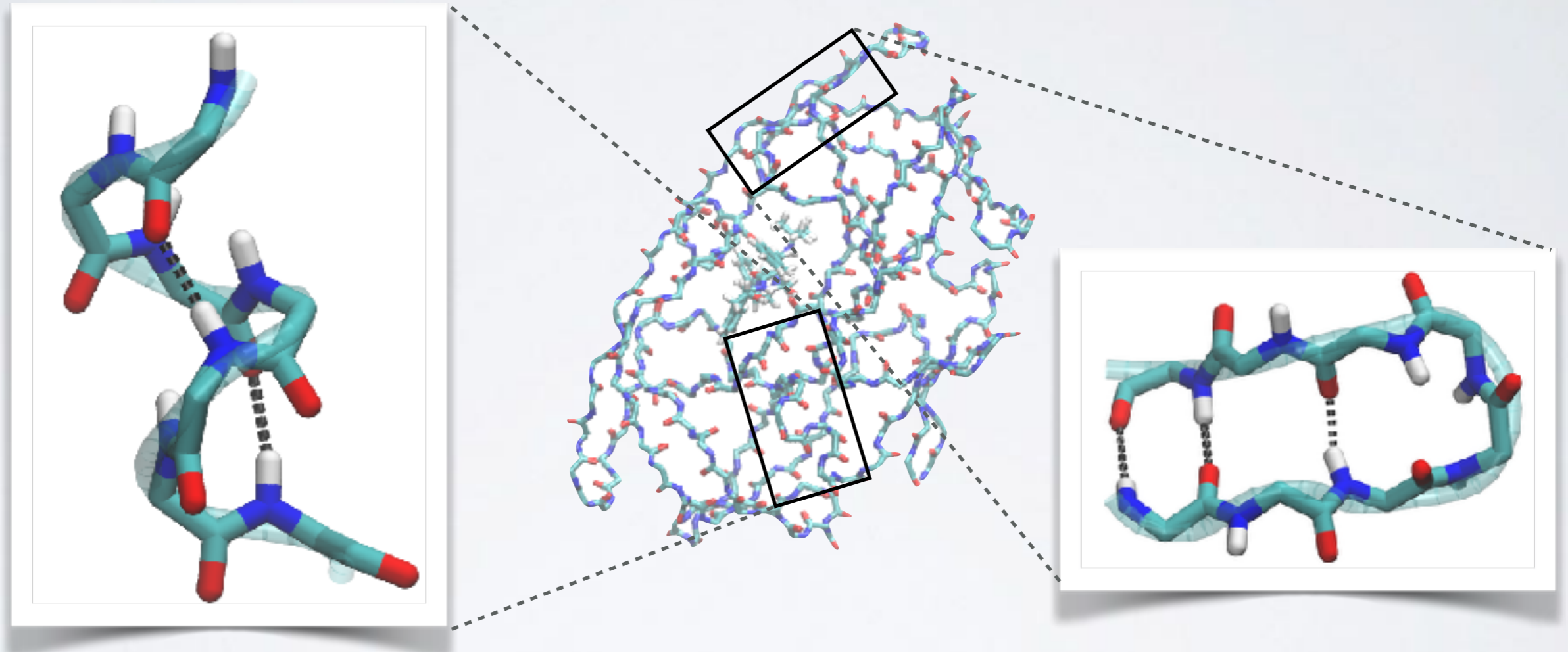
- Proteins are close packed solid but flexible objects (globular)



- Due to their large size and complexity it is often hard to see what's important in the structure

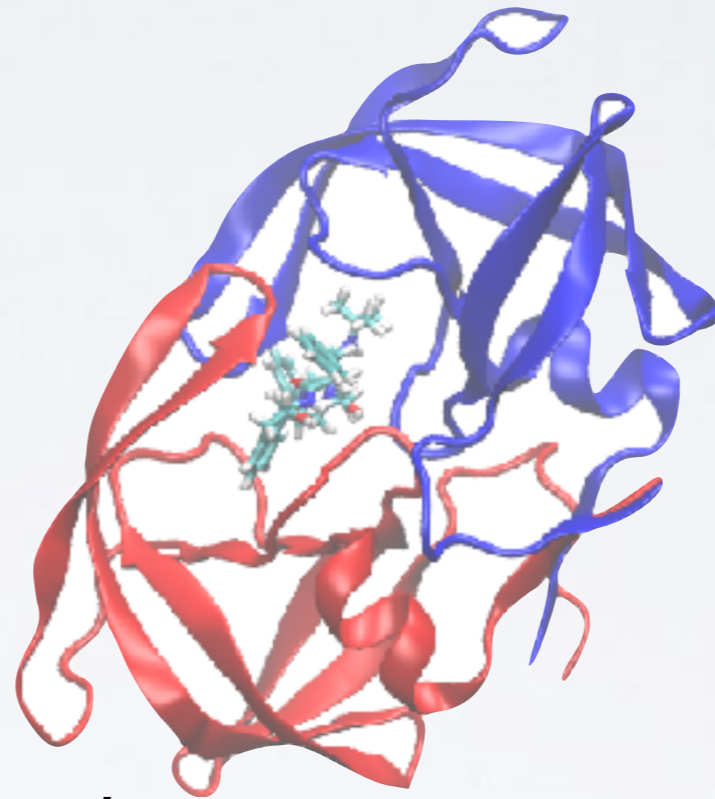


- Backbone or main-chain representation can help trace chain topology



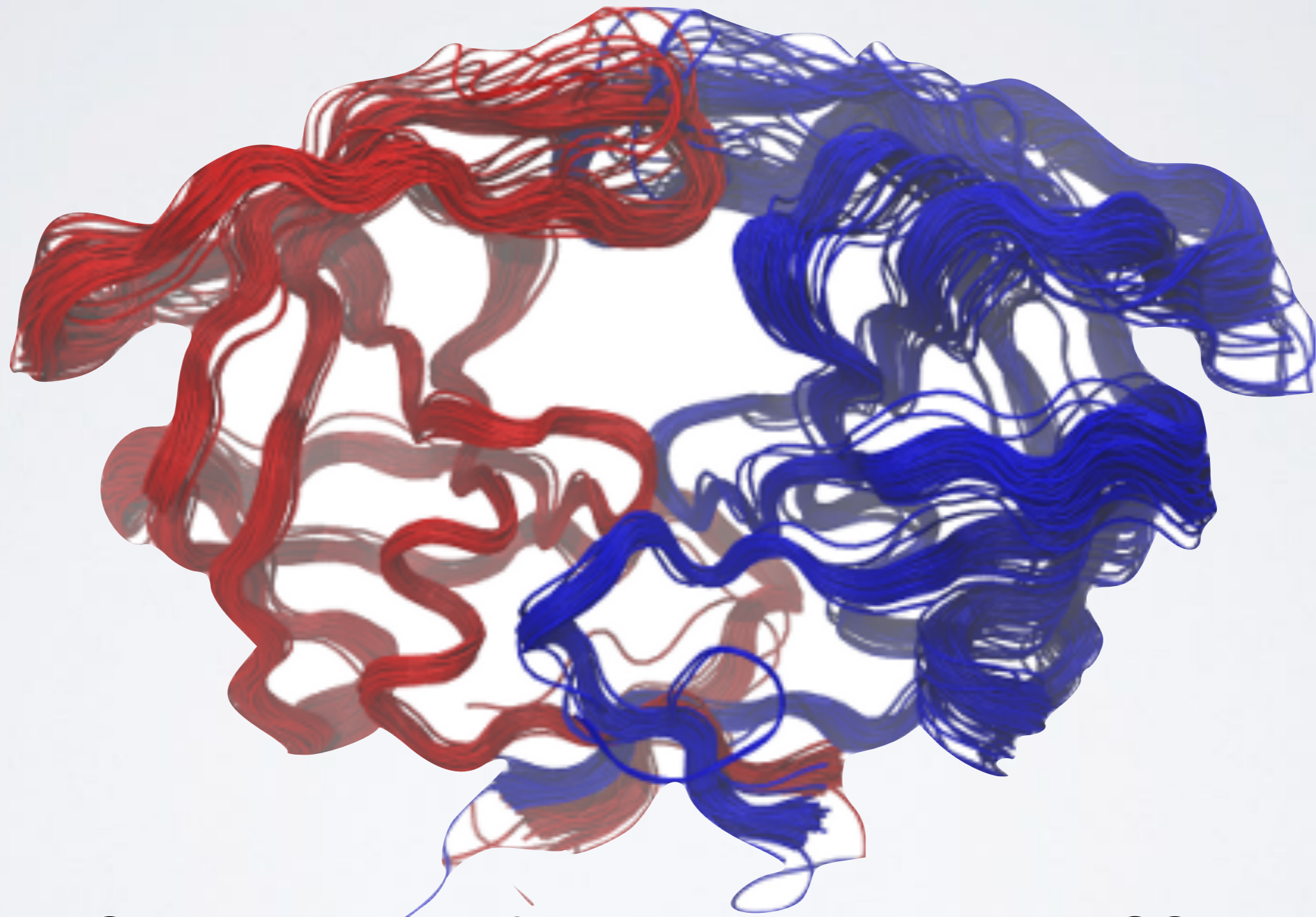
- Backbone or main-chain representation can help trace chain topology & reveal secondary structure





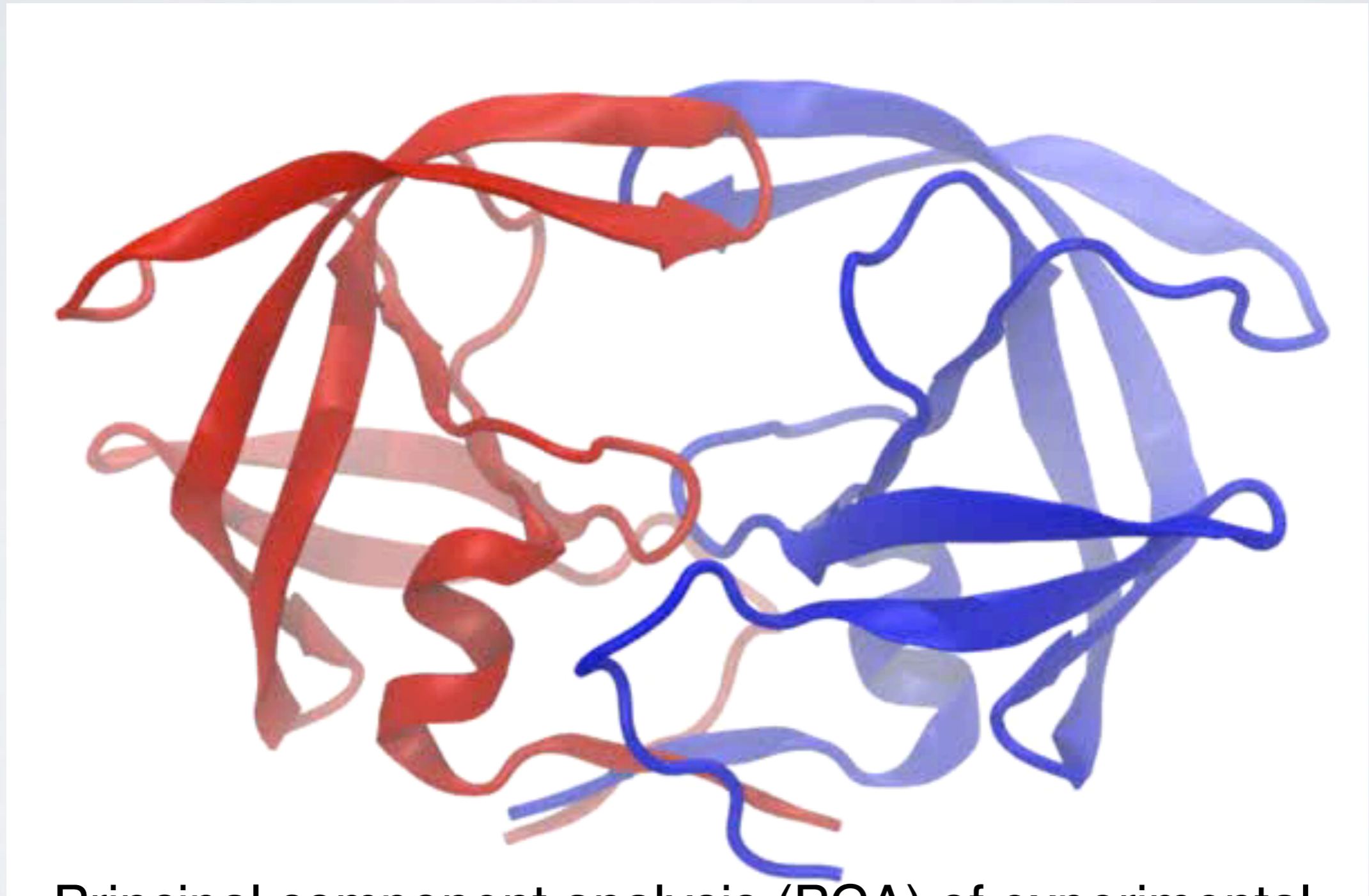
- Simplified secondary structure representations are commonly used to communicate structural details
- Now we can clearly see 2<sup>o</sup>, 3<sup>o</sup> and 4<sup>o</sup> structure
- Coiled chain of connected secondary structures

# DISPLACEMENTS REFLECT INTRINSIC FLEXIBILITY



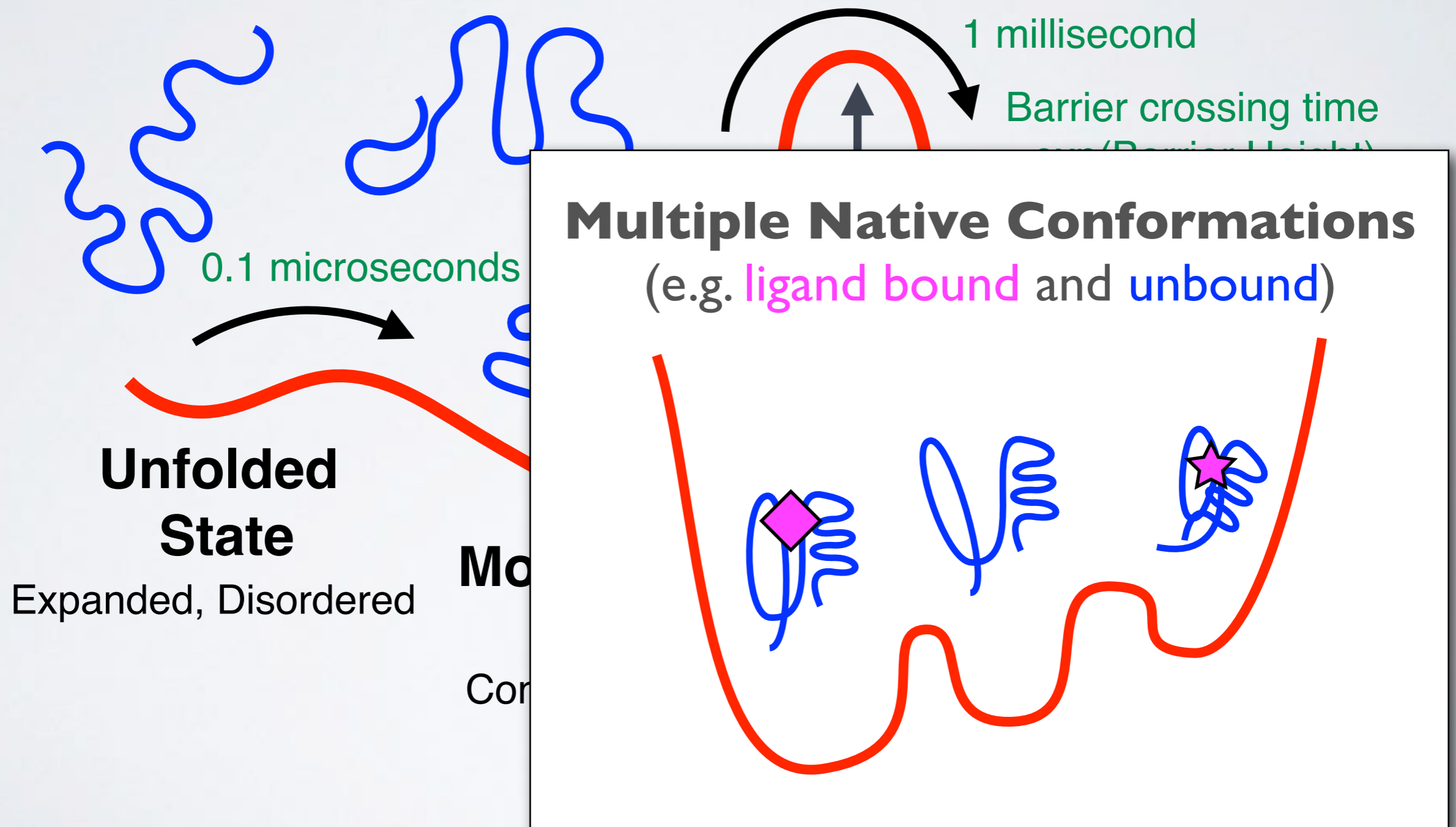
Superposition of all 482 structures in RCSB  
PDB (23/09/2015)

# DISPLACEMENTS REFLECT INTRINSIC FLEXIBILITY



Principal component analysis (PCA) of experimental structures

# KEY CONCEPT: ENERGY LANDSCAPE



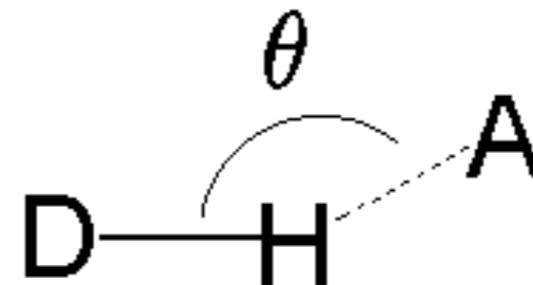
# Key forces affecting structure:

- H-bonding
- Van der Waals
- Electrostatics
- Hydrophobicity

Hydrogen-bond donor      Hydrogen-bond acceptor



← d →

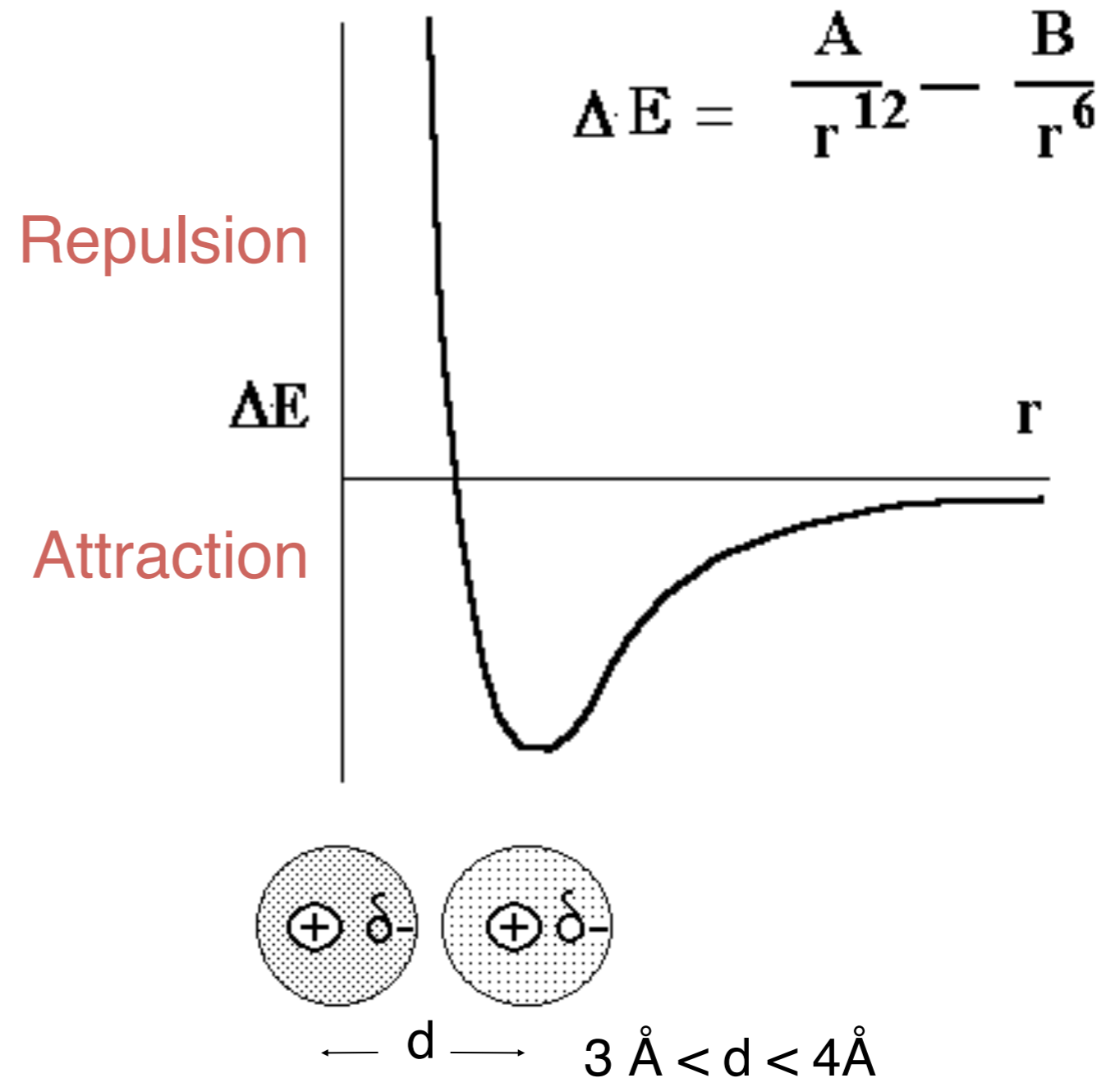


$$2.6 \text{ \AA} < d < 3.1 \text{ \AA}$$

$$150^\circ < \theta < 180^\circ$$

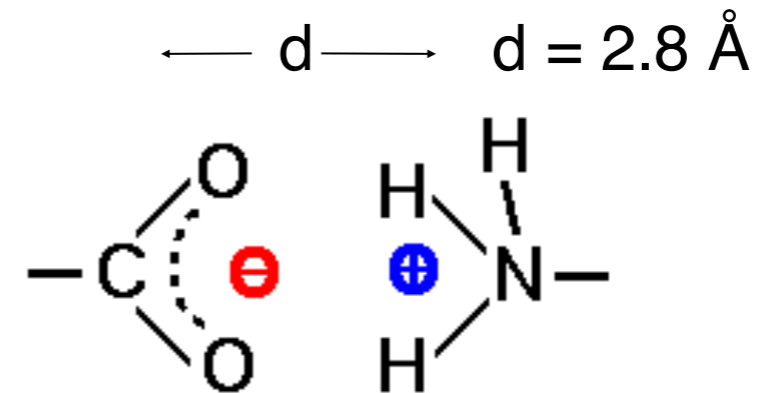
# Key forces affecting structure:

- H-bonding
- **Van der Waals**
- Electrostatics
- Hydrophobicity



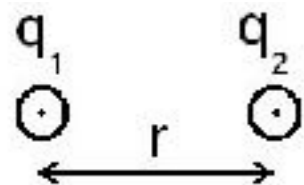
# Key forces affecting structure:

- H-bonding
- Van der Waals
- **Electrostatics**
- Hydrophobicity



carboxyl group and amino group

(some time called IONIC BONDS or SALT BRIDGES)



Coulomb's law

$$E = \frac{K q_1 q_2}{D r}$$

E = Energy

k = constant

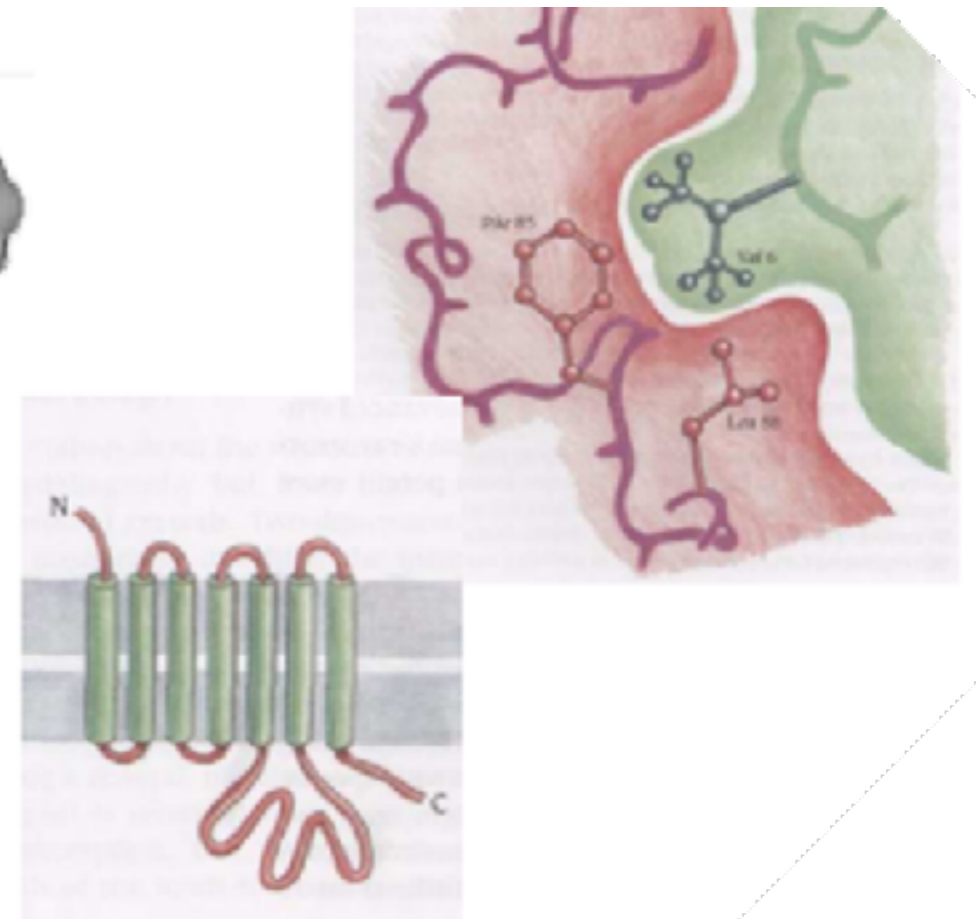
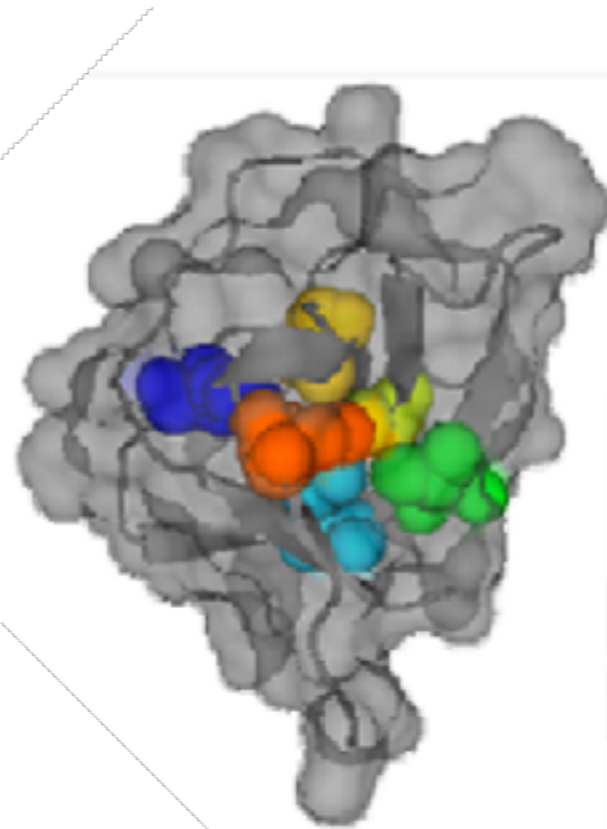
D = Dielectric constant (vacuum = 1; H<sub>2</sub>O = 80)

q<sub>1</sub> & q<sub>2</sub> = electronic charges (Coulombs)

r = distance (Å)

# Key forces affecting structure:

- H-bonding
- Van der Waals
- Electrostatics
- **Hydrophobicity**



The force that causes hydrophobic molecules or nonpolar portions of molecules to aggregate together rather than to dissolve in water is called Hydrophobicity (*Greek, "water fearing"*). This is not a separate bonding force; rather, it is the result of the energy required to insert a nonpolar molecule into water.



Do it Yourself!

# Hand-on time!

[https://bioboot.github.io/bimm143\\_S18/lectures/#11](https://bioboot.github.io/bimm143_S18/lectures/#11)

Focus on **section 1** to **3** only please!

# SIDE-NOTE: PDB FILE FORMAT

|      |         | Amino Acid |     | Chain name | Sequence Number | -----Coordinates----- |       |       |        |
|------|---------|------------|-----|------------|-----------------|-----------------------|-------|-------|--------|
|      | Element |            |     |            |                 | X                     | Y     | Z     | (etc.) |
| ATOM | 1       | N          | ASP | L          | 1               | 4.060                 | 7.307 | 5.186 | ...    |
| ATOM | 2       | CA         | ASP | L          | 1               | 4.042                 | 7.776 | 6.553 | ...    |
| ATOM | 3       | C          | ASP | L          | 1               | 2.668                 | 8.426 | 6.644 | ...    |
| ATOM | 4       | O          | ASP | L          | 1               | 1.987                 | 8.438 | 5.606 | ...    |
| ATOM | 5       | CB         | ASP | L          | 1               | 5.090                 | 8.827 | 6.797 | ...    |
| ATOM | 6       | CG         | ASP | L          | 1               | 6.338                 | 8.761 | 5.929 | ...    |
| ATOM | 7       | OD1        | ASP | L          | 1               | 6.576                 | 9.758 | 5.241 | ...    |
| ATOM | 8       | OD2        | ASP | L          | 1               | 7.065                 | 7.759 | 5.948 | ...    |

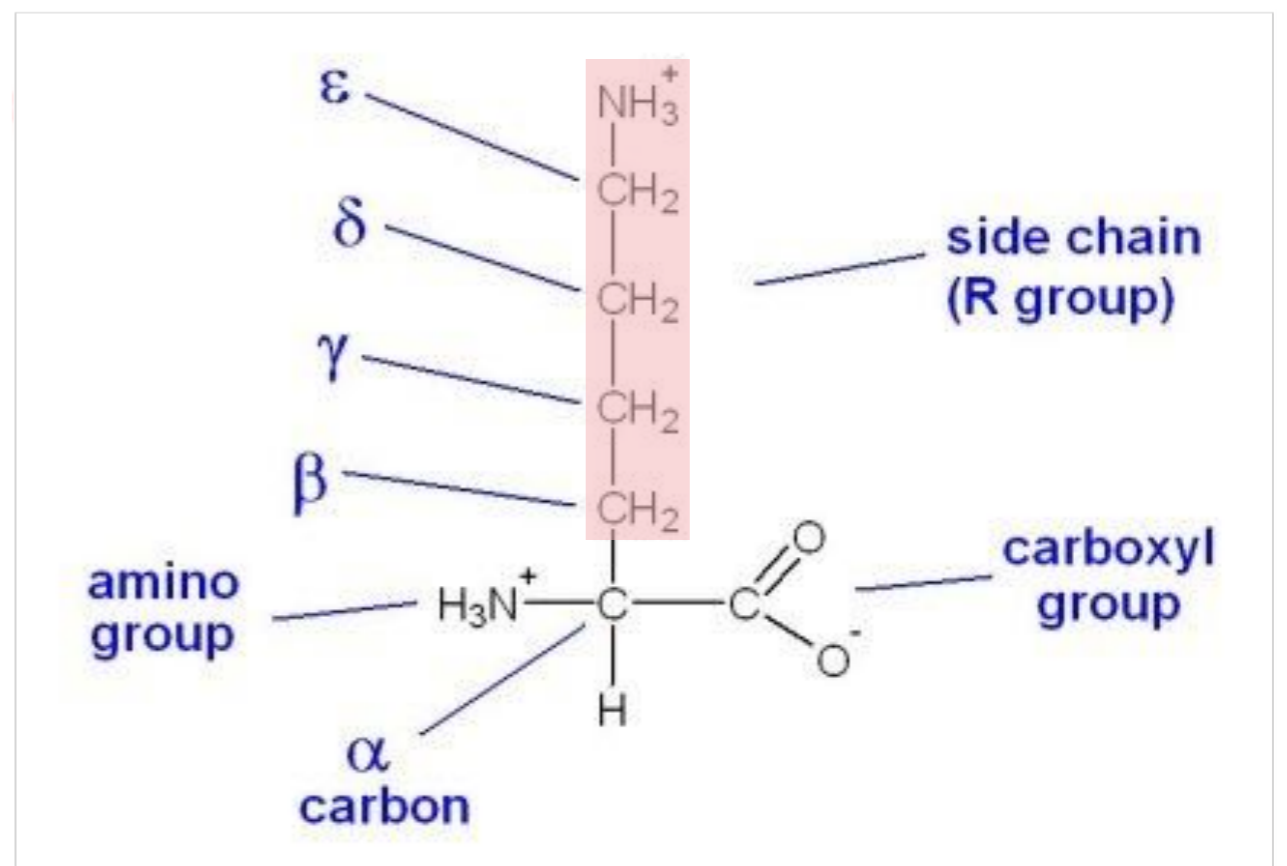
\\  
Element position within amino acid

- **PDB files** contains atomic coordinates and associated information.

# SIDE-NOTE: PDB FILE FORMAT

|      |   | Amino Acid |     |   | Chain |
|------|---|------------|-----|---|-------|
|      |   | Element    |     |   | Se    |
| ATOM | 1 | N          | ASP | L | 1     |
| ATOM | 2 | CA         | ASP | L | 1     |
| ATOM | 3 | C          | ASP | L | 1     |
| ATOM | 4 | O          | ASP | L | 1     |
| ATOM | 5 | CB         | ASP | L | 1     |
| ATOM | 6 | CG         | ASP | L | 1     |
| ATOM | 7 | OD1        | ASP | L | 1     |
| ATOM | 8 | OD2        | ASP | L | 1     |

\\  
Element position within amino acid



- **PDB files** contains atomic coordinates and associated information.

# NEXT UP:

- ▶ **Overview of structural bioinformatics**

- Major motivations, goals and challenges

- ▶ **Fundamentals of protein structure**

- Composition and dynamics

Change from here on!!!

Just do docking???

- ▶ **Representing and predicting protein structure**

- Modeling energy as a function of structure

- ▶ **Example application areas**

- Predicting functional dynamics & drug discovery

**KEY CONCEPT:** POTENTIAL FUNCTIONS  
DESCRIBE A SYSTEMS **ENERGY** AS A FUNCTION  
OF ITS **STRUCTURE**

Two main approaches:

(1). **Physics-Based**

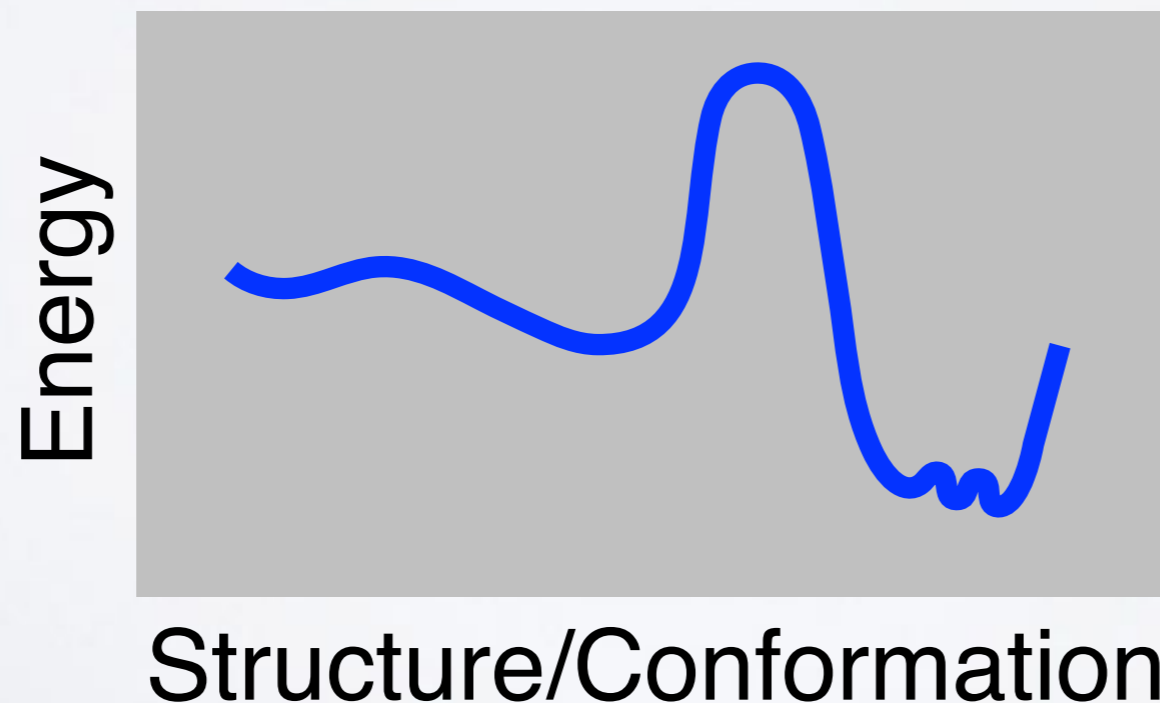
(2). **Knowledge-Based**

**KEY CONCEPT:** POTENTIAL FUNCTIONS  
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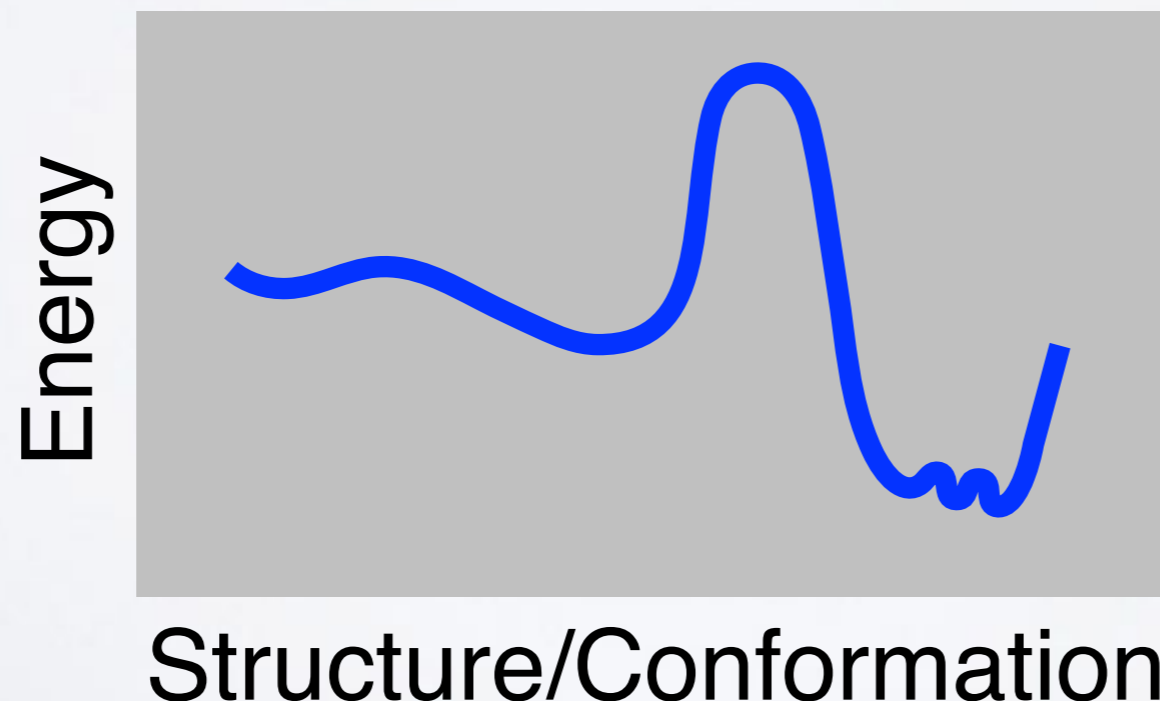


**KEY CONCEPT:** POTENTIAL FUNCTIONS  
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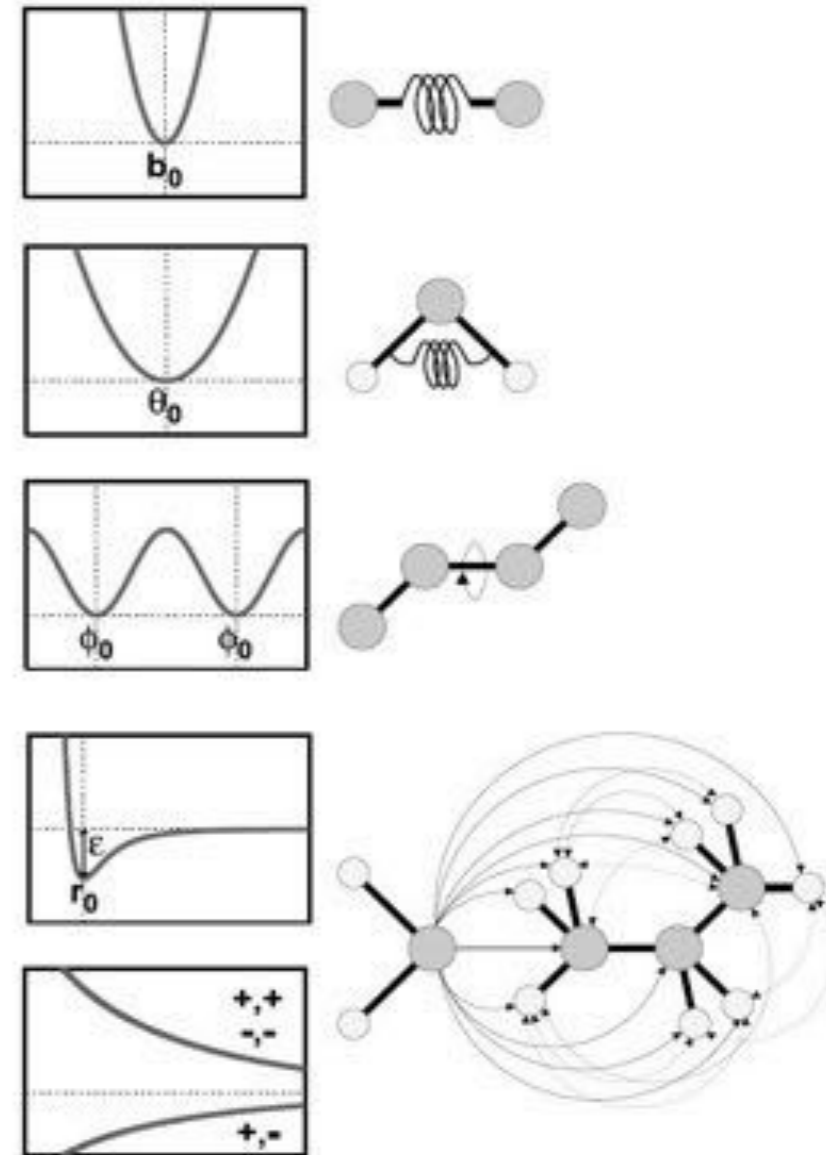
(2). Knowledge-Based



# PHYSICS-BASED POTENTIALS

## ENERGY TERMS FROM PHYSICAL THEORY

$$\begin{aligned}
 U(\vec{R}) = & \underbrace{\sum_{\text{bonds}} k_i^{\text{bond}} (r_i - r_0)^2}_{U_{\text{bond}}} + \underbrace{\sum_{\text{angles}} k_i^{\text{angle}} (\theta_i - \theta_0)^2}_{U_{\text{angle}}} + \\
 & \underbrace{\sum_{\text{dihedrals}} k_i^{\text{dihe}} [1 + \cos(n_i \phi_i + \delta_i)]}_{U_{\text{dihedral}}} + \\
 & \underbrace{\sum_i \sum_{j \neq i} 4\epsilon_{ij} \left[ \left( \frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{ij}}{r_{ij}} \right)^6 \right]}_{U_{\text{nonbond}}} + \sum_i \sum_{j \neq i} \frac{q_i q_j}{\epsilon r_{ij}}
 \end{aligned}$$



$U_{\text{bond}}$  = oscillations about the equilibrium bond length

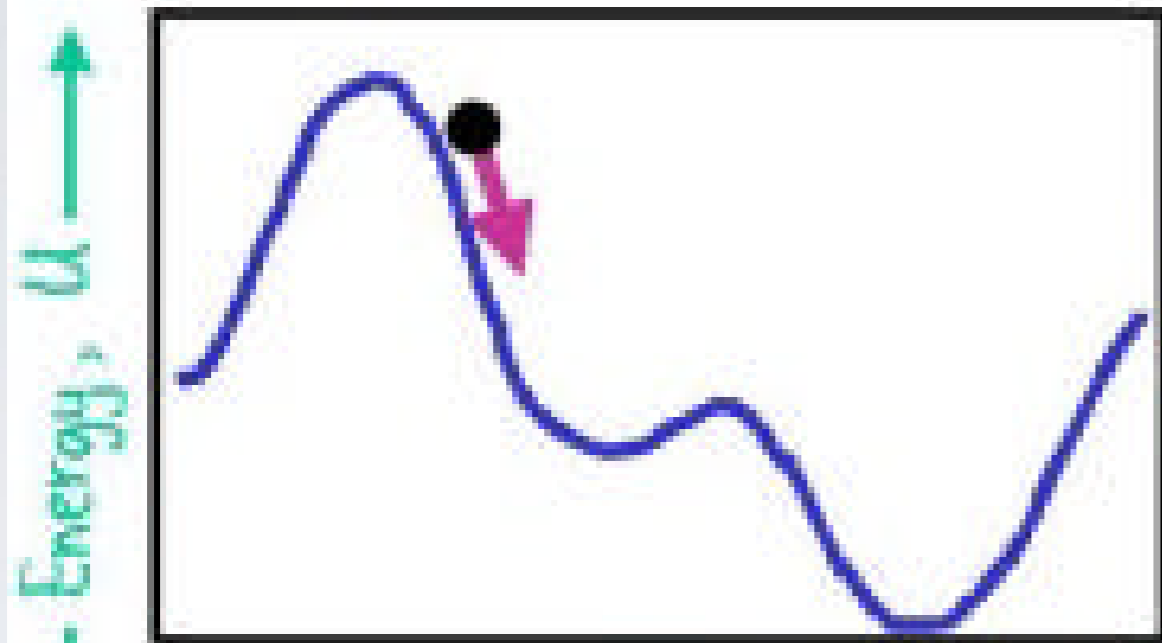
$U_{\text{angle}}$  = oscillations of 3 atoms about an equilibrium bond angle

$U_{\text{dihedral}}$  = torsional rotation of 4 atoms about a central bond

$U_{\text{nonbond}}$  = non-bonded energy terms (electrostatics and Lennard-Jones)



# TOTAL POTENTIAL ENERGY



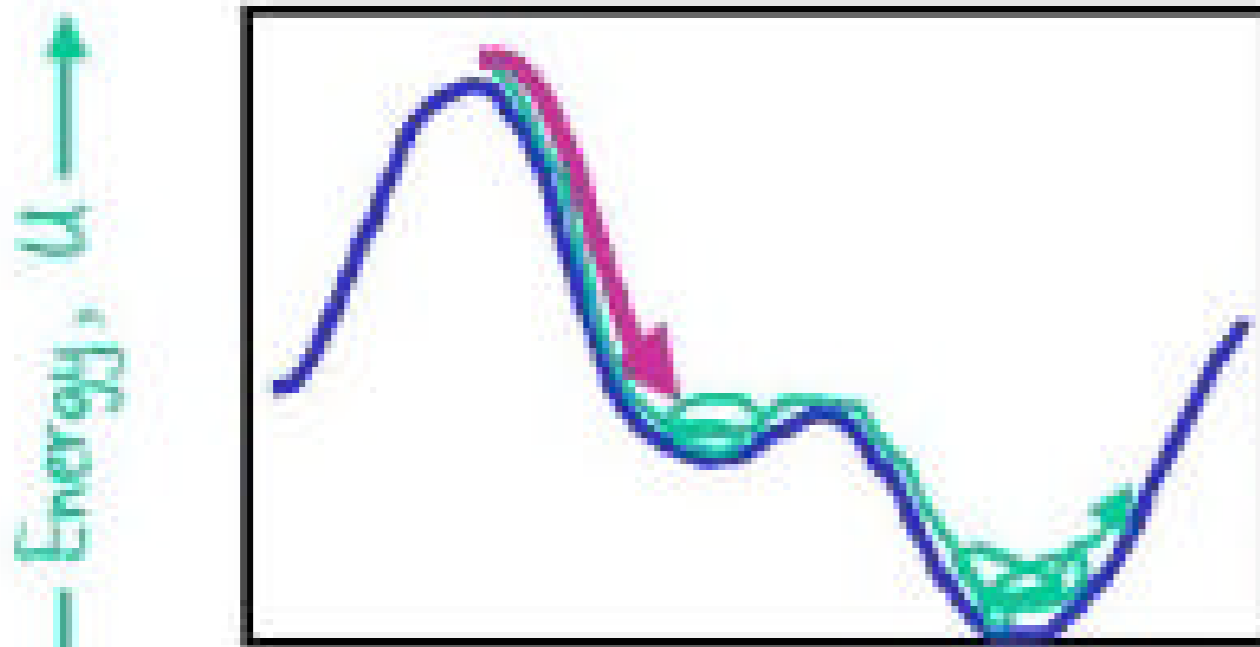
$$F(x) = -dU/dx$$



- The total potential energy or enthalpy fully defines the system,  $U$ .
- The forces are the gradients of the energy.
- The energy is a sum of independent terms for:  
Bond, Bond angles, Torsion angles and non-bonded atom pairs.

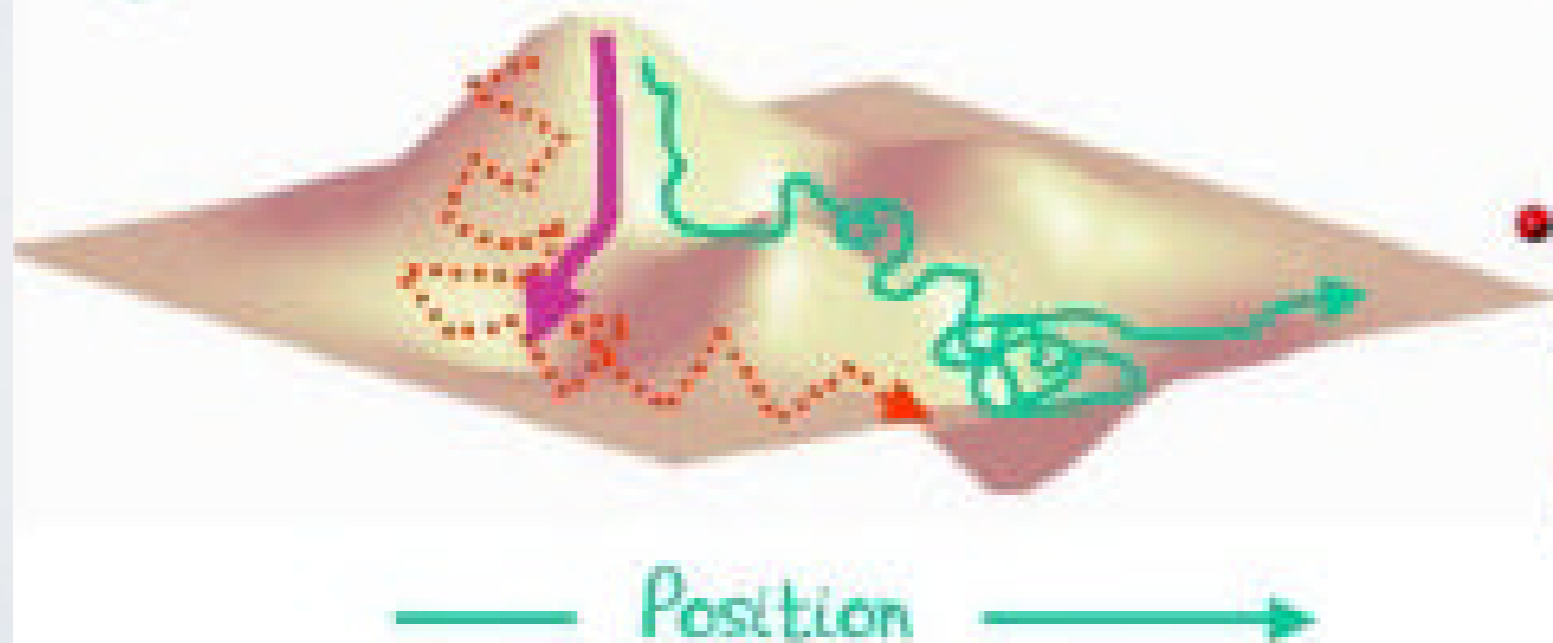
Slide Credit: Michael Levitt

# MOVING OVER THE ENERGY SURFACE



- Energy Minimization drops into local minimum.

- Molecular Dynamics uses thermal energy to move smoothly over surface.



- Monte Carlo Moves are random. Accept with probability  $\exp(-\Delta U/kT)$ .

Slide Credit: Michael Levitt

# PHYSICS-ORIENTED APPROACHES

## Weaknesses

Fully physical detail becomes computationally intractable

Approximations are unavoidable

(Quantum effects approximated classically, water may be treated crudely)

Parameterization still required

## Strengths

Interpretable, provides guides to design

Broadly applicable, in principle at least

Clear pathways to improving accuracy

## Status

Useful, widely adopted but far from perfect

Multiple groups working on fewer, better approxs

Force fields, quantum

entropy, water effects

Moore's law: hardware improving

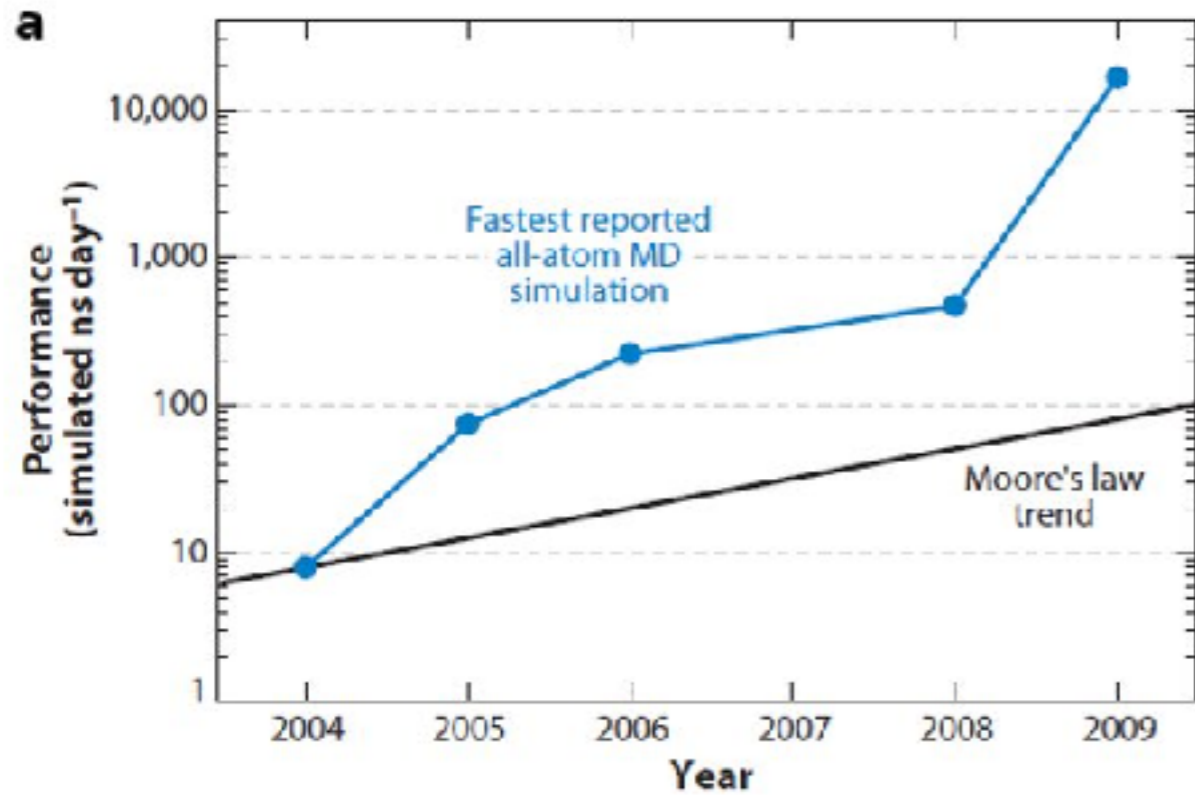
## HOW COMPUTERS HAVE CHANGED

| DATE   | COST    | SPEED   | MEMORY | SIZE   |
|--------|---------|---------|--------|--------|
| 1967   | \$40M   | 0.1 MHz | 1 MB   | WALL   |
| 2013   | \$4,000 | 1 GHz   | 10 GB  | LAPTOP |
| CHANGE | 10,000  | 10,000  | 10,000 | 10,000 |

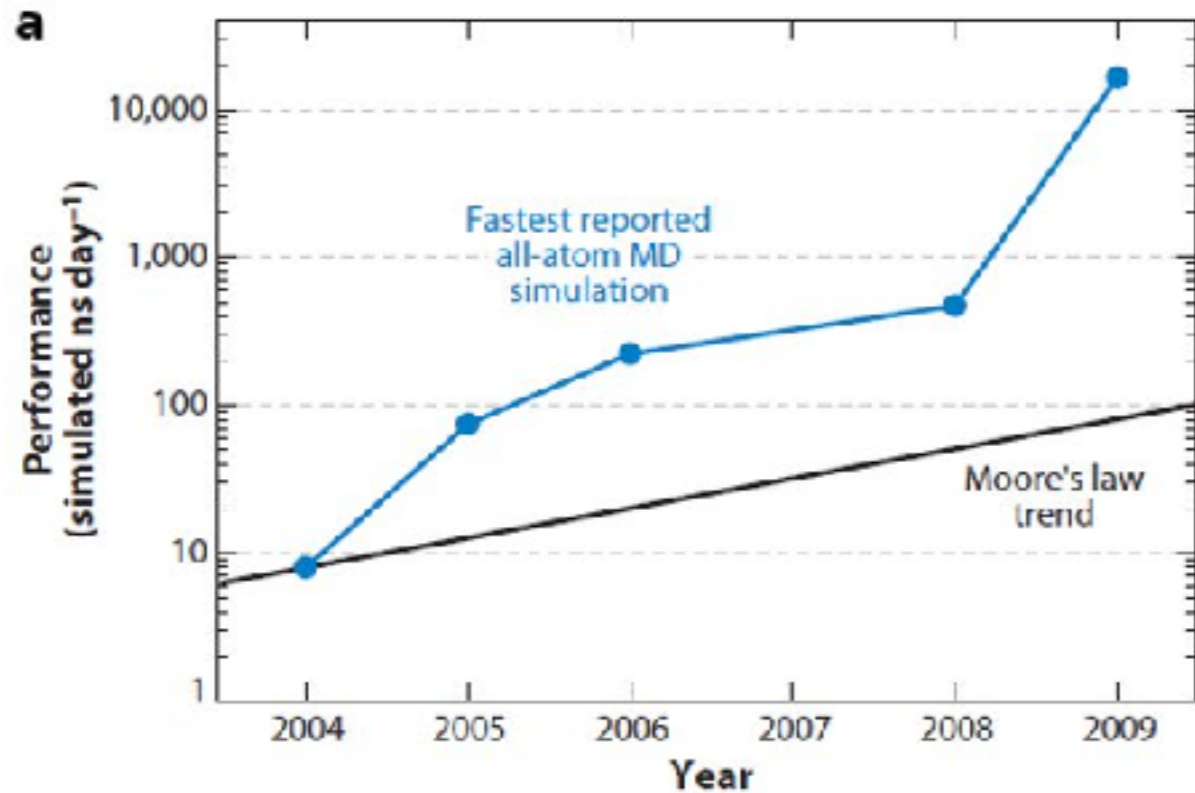
If cars were like computers then a new Volvo would cost \$3, would have a top speed of 1,000,000 km/hr, would carry 50,000 adults and would park in a shedbox



# SIDE-NOTE: GPUS AND ANTON SUPERCOMPUTER



# SIDE-NOTE: GPUS AND ANTON SUPERCOMPUTER



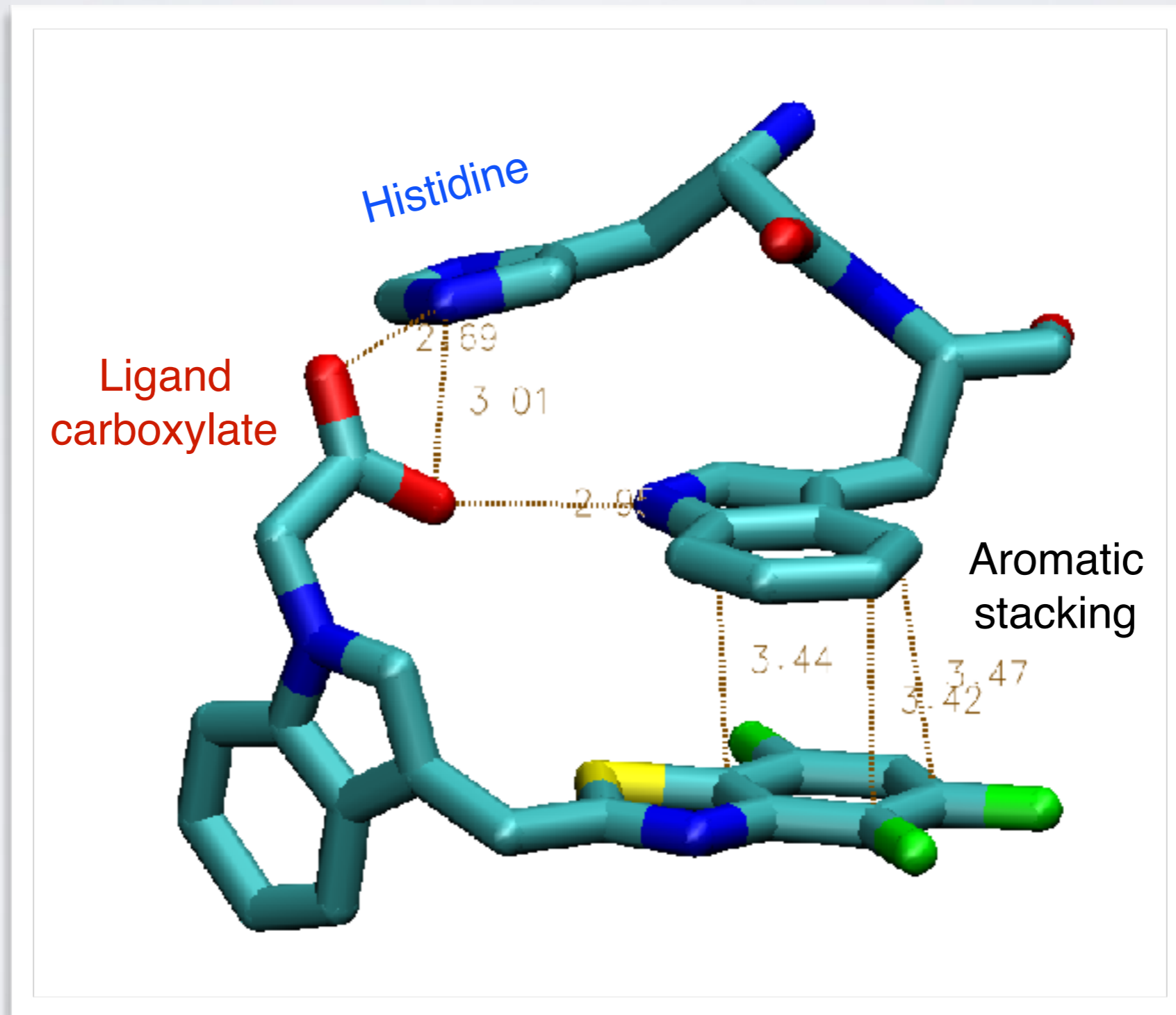
**KEY CONCEPT:** POTENTIAL FUNCTIONS  
DESCRIBE A SYSTEMS **ENERGY** AS A FUNCTION  
OF ITS **STRUCTURE**

Two main approaches:

(1). **Physics-Based**

(2). **Knowledge-Based**

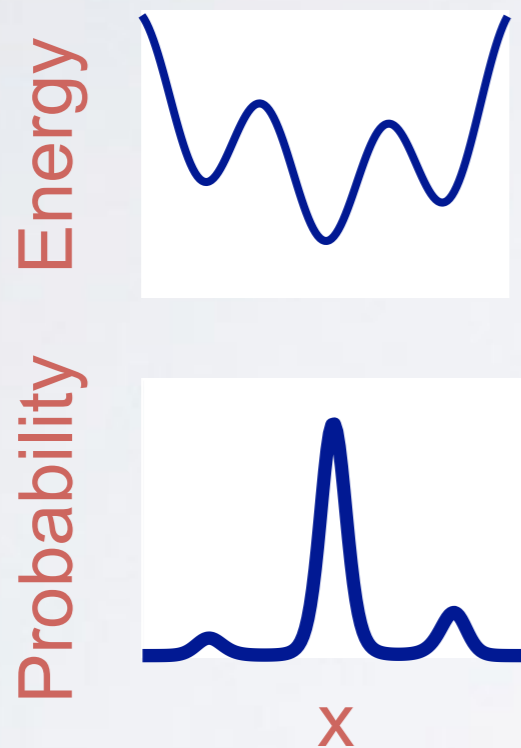
# KNOWLEDGE-BASED DOCKING POTENTIALS





# ENERGY DETERMINES **PROBABILITY** (STABILITY)

Basic idea: Use probability as a proxy for energy



Boltzmann:

$$p(r) \propto e^{-E(r)/RT}$$

Inverse Boltzmann:

$$E(r) = -RT \ln [p(r)]$$

Example: ligand **carboxylate O** to protein **histidine N**

Find all protein-ligand structures in the PDB with a ligand carboxylate **O**

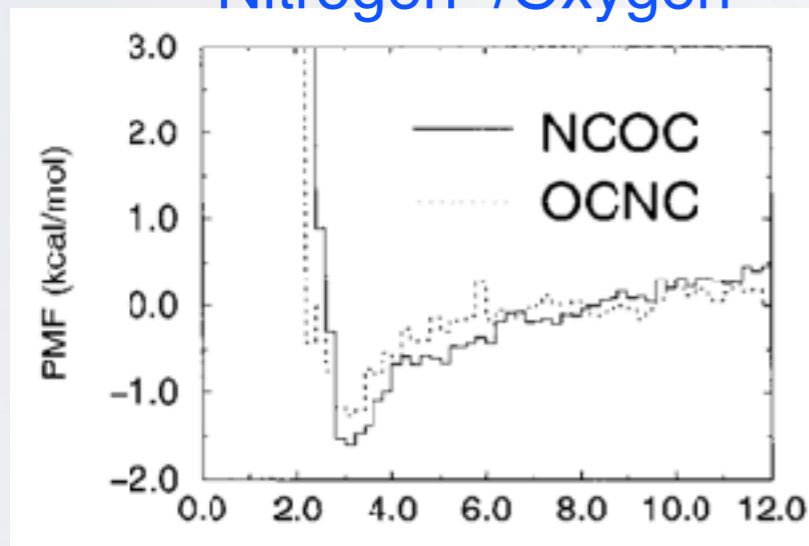
1. For each structure, histogram the distances from **O** to every histidine **N**
2. Sum the histograms over all structures to obtain  $p(r_{\text{O-N}})$
3. Compute  $E(r_{\text{O-N}})$  from  $p(r_{\text{O-N}})$

# KNOWLEDGE-BASED DOCKING POTENTIALS

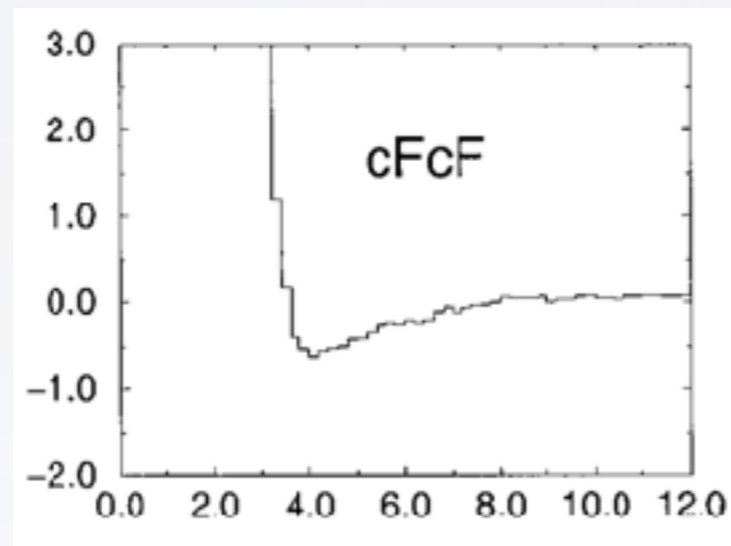
“PMF”, Muegge & Martin, J. Med. Chem. (1999) 42:791

A few types of atom pairs, out of several hundred total

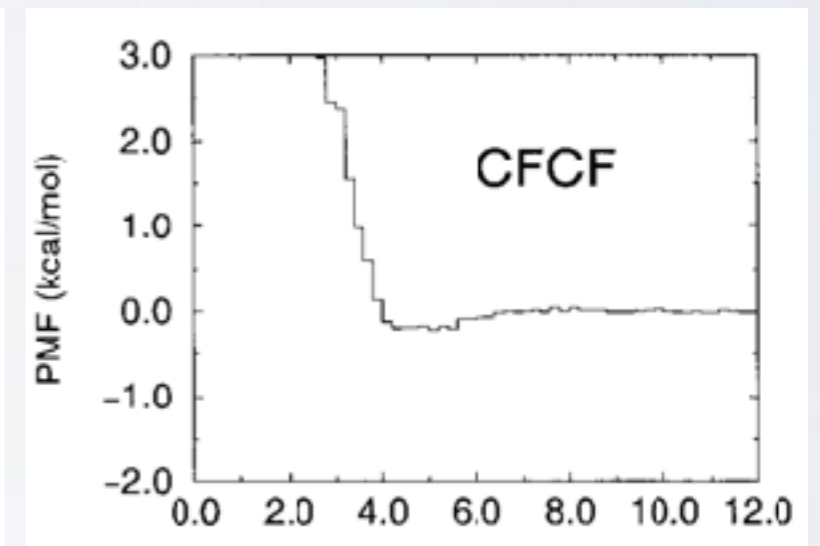
Nitrogen<sup>+</sup>/Oxygen<sup>-</sup>



Aromatic carbons



Aliphatic carbons



Atom-atom distance (Angstroms)

$$E_{prot-lig} = E_{vdw} + \sum_{pairs (ij)} E_{type(ij)}(r_{ij})$$

# KNOWLEDGE-BASED POTENTIALS

## Weaknesses

Accuracy limited by availability of data

## Strengths

Relatively easy to implement

Computationally fast

## Status

Useful, far from perfect

May be at point of diminishing returns

(not always clear how to make improvements)

Do it Yourself!

# Hand-on time!

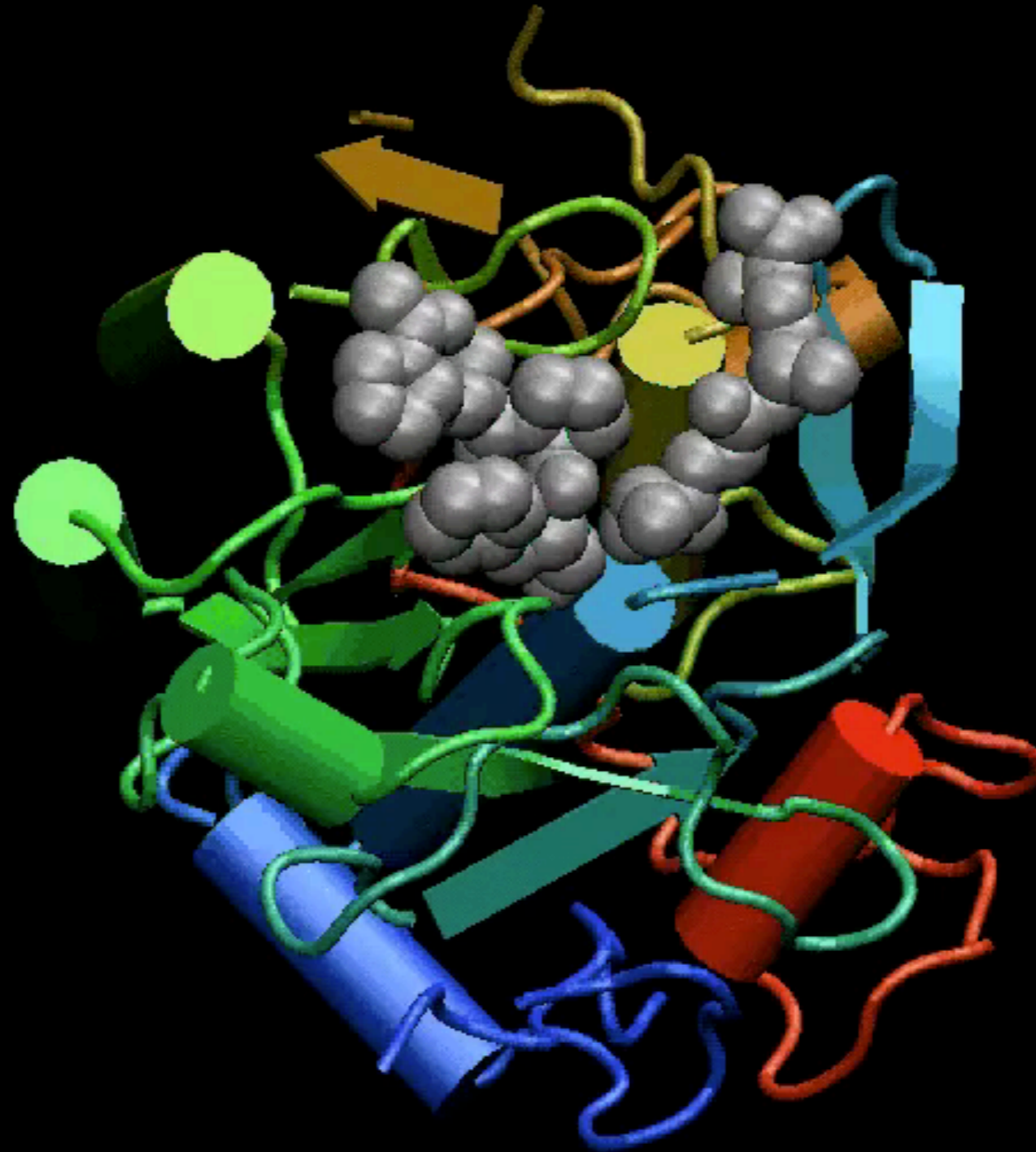
[https://bioboot.github.io/bimm143\\_S18/lectures/#11](https://bioboot.github.io/bimm143_S18/lectures/#11)

Focus on **section 4 & 5**

# NEXT UP:

- ▶ **Overview of structural bioinformatics**
  - Major motivations, goals and challenges
- ▶ **Fundamentals of protein structure**
  - Composition, form, forces and dynamics
- ▶ **Representing and interpreting protein structure**
  - Modeling energy as a function of structure
- ▶ **Example application areas**
  - Predicting functional dynamics & drug discovery

NMA models the protein as a network of elastic strings



Proteinase K

Do it Yourself!

# Hand-on time!

[https://bioboot.github.io/bimm143\\_S18/lectures/#11](https://bioboot.github.io/bimm143_S18/lectures/#11)

Focus on **section 6** to **7**

**Optional:**  
Stop here for Today!

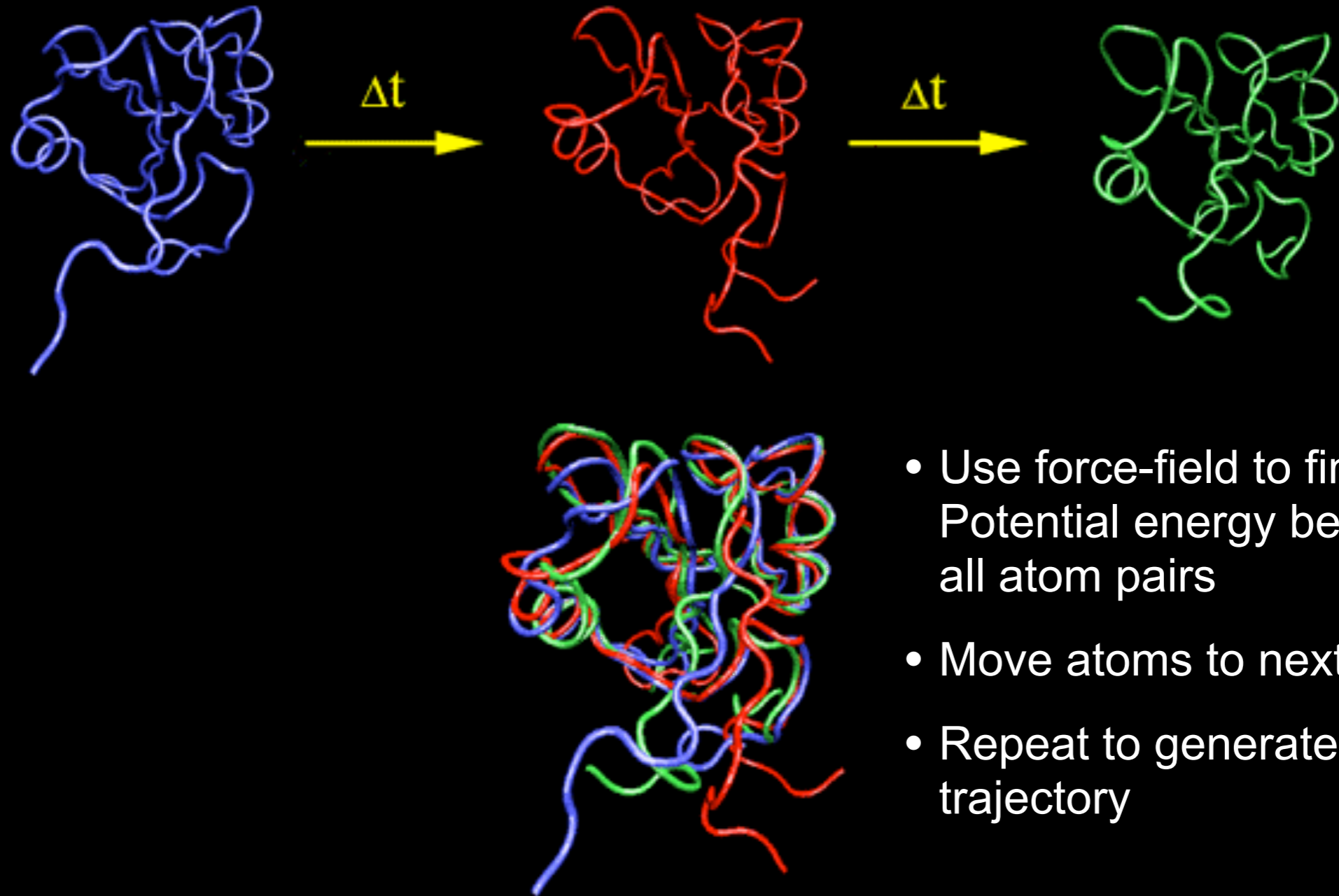
[ [Muddy Point Assessment](#) ]



# PREDICTING FUNCTIONAL DYNAMICS

- Proteins are intrinsically flexible molecules with internal motions that are often intimately coupled to their biochemical function
  - E.g. ligand and substrate binding, conformational activation, allosteric regulation, etc.
- Thus knowledge of dynamics can provide a deeper understanding of the mapping of structure to function
  - Molecular dynamics (MD) and normal mode analysis (NMA) are two major methods for predicting and characterizing molecular motions and their properties

# MOLECULAR DYNAMICS SIMULATION



McCammon, Gelin & Karplus, *Nature* (1977)

[ See: <https://www.youtube.com/watch?v=ui1ZysMFcKk> ]

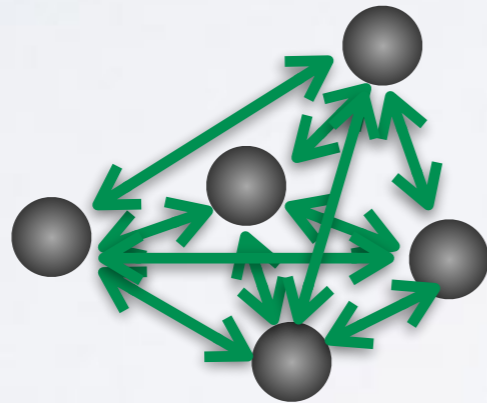
- ▶ Divide **time** into discrete ( $\sim 1$ fs) **time steps** ( $\Delta t$ )  
(for integrating equations of motion, see below)



- ▶ Divide **time** into discrete ( $\sim 1$ fs) **time steps** ( $\Delta t$ )  
(for integrating equations of motion, see below)



- ▶ At each time step calculate pair-wise atomic **forces** ( $F(t)$ )  
(by evaluating **force-field** gradient)



*Nucleic motion described classically*

$$m_i \frac{d^2 \vec{R}_i}{dt^2} = -\vec{\nabla}_i E(\vec{R})$$

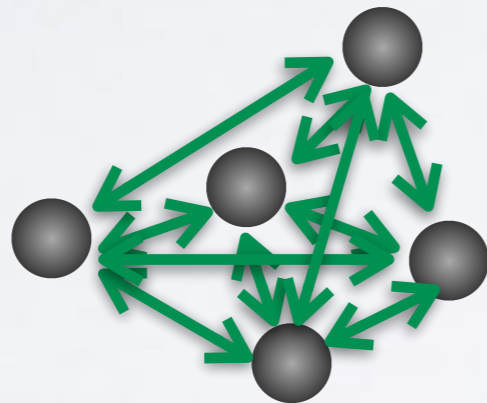
*Empirical force field*

$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_i(\vec{R})$$

- ▶ Divide **time** into discrete ( $\sim 1$ fs) **time steps** ( $\Delta t$ )  
(for integrating equations of motion, see below)



- ▶ At each time step calculate pair-wise atomic **forces** ( $F(t)$ )  
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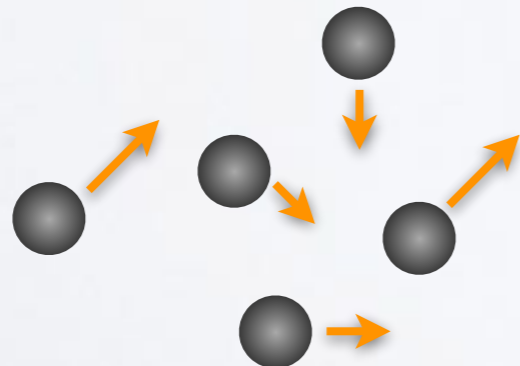
*Nucleic motion described classically*

$$m_i \frac{d^2 \vec{R}_i}{dt^2} = -\vec{\nabla}_i E(\vec{R})$$

*Empirical force field*

$$E(\vec{R}) = \sum_{\text{bonded}} E_i(\vec{R}) + \sum_{\text{non-bonded}} E_i(\vec{R})$$

- ▶ Use the forces to calculate **velocities** and move atoms to new **positions**  
(by integrating numerically via the “leapfrog” scheme)



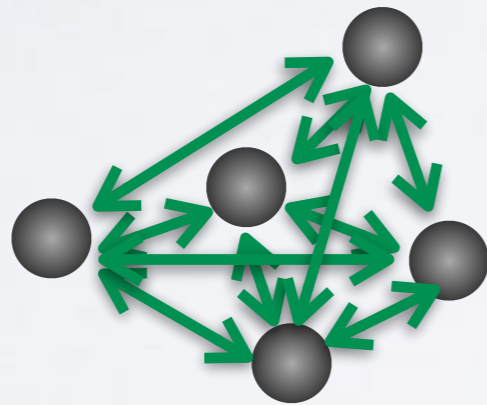
$$\begin{aligned} \mathbf{v}\left(t + \frac{\Delta t}{2}\right) &= \mathbf{v}\left(t - \frac{\Delta t}{2}\right) + \frac{\mathbf{F}(t)}{m} \Delta t \\ \mathbf{r}(t + \Delta t) &= \mathbf{r}(t) + \mathbf{v}\left(t + \frac{\Delta t}{2}\right) \Delta t \end{aligned}$$

# BASIC ANATOMY OF A MD SIMULATION

- ▶ Divide **time** into discrete ( $\sim 1$ fs) **time steps** ( $\Delta t$ )  
(for integrating equations of motion, see below)



- ▶ At each time step calculate pair-wise atomic **forces** ( $F(t)$ )  
(by evaluating **force-field** gradient)



*Nucleic motion described classically*

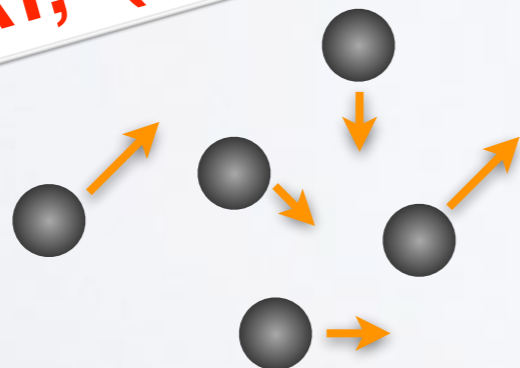
$$m_i \frac{d^2 \vec{R}_i}{dt^2} = -\vec{\nabla}_i E(\vec{R})$$

*Empirical force field*

$$E(\vec{R}) = \sum_{\text{non-bonded}} E_i(\vec{R})$$

- ▶ Use the forces to calculate **velocities** and move atoms to new **positions**  
(the integration is done numerically via the “leapfrog” scheme)

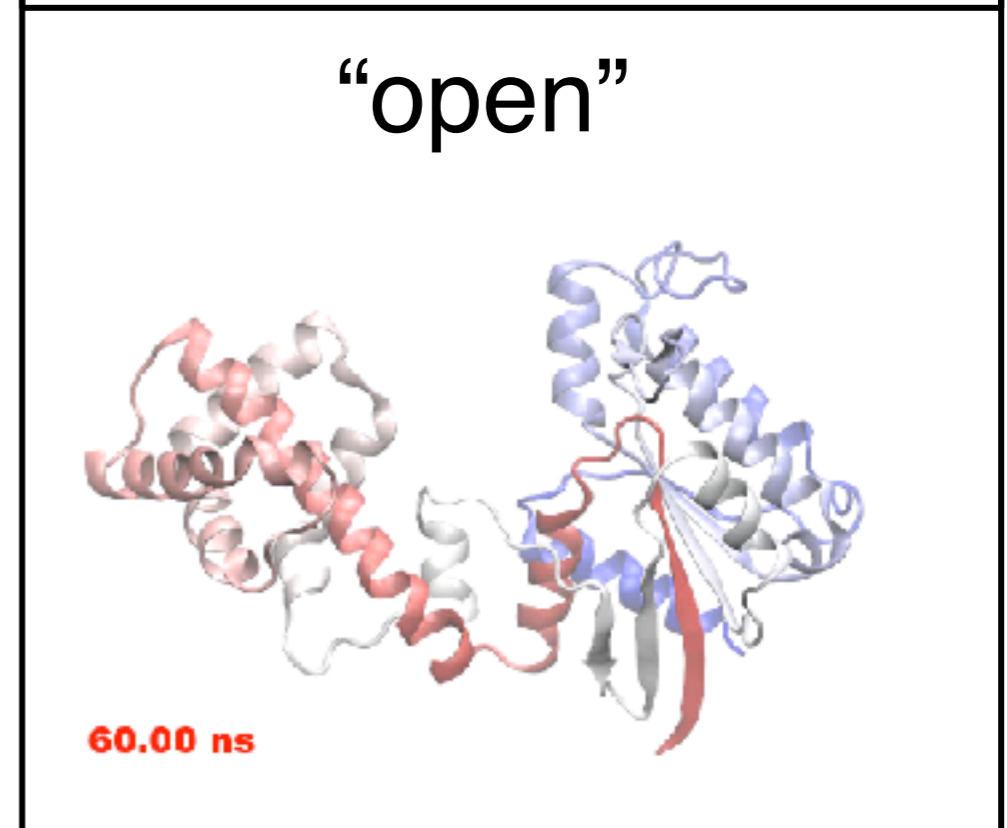
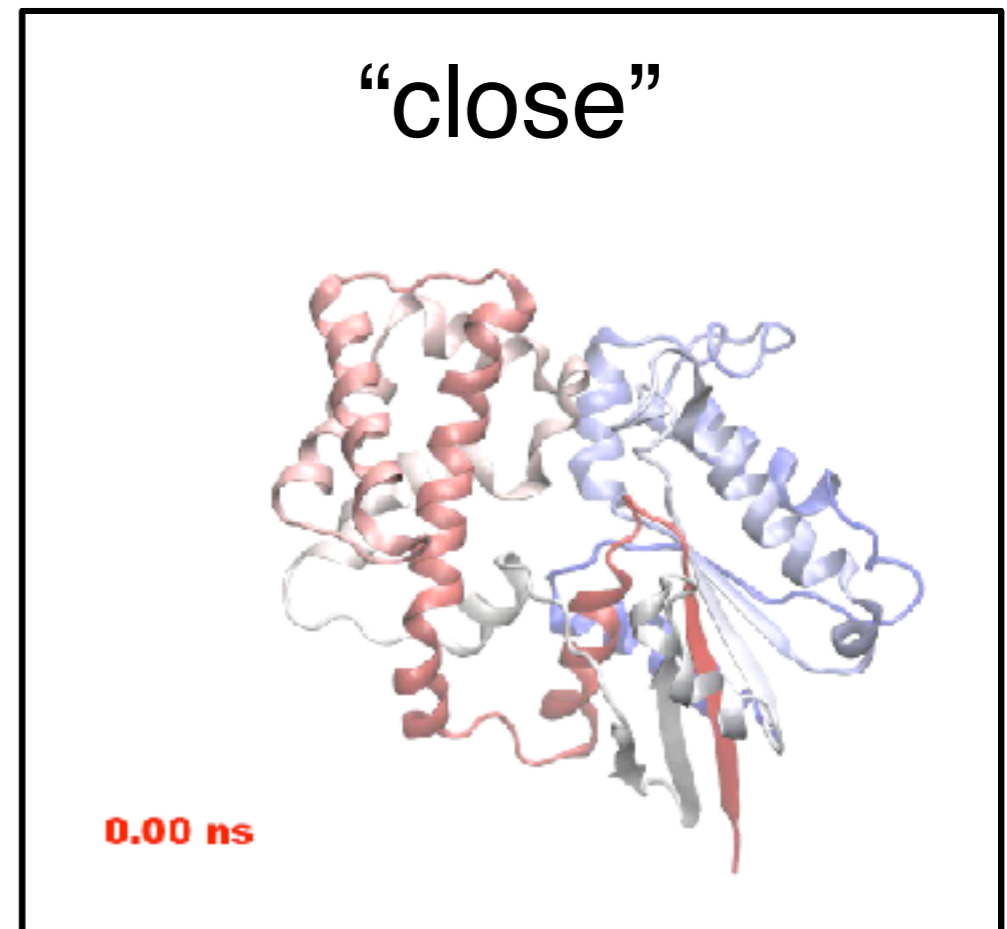
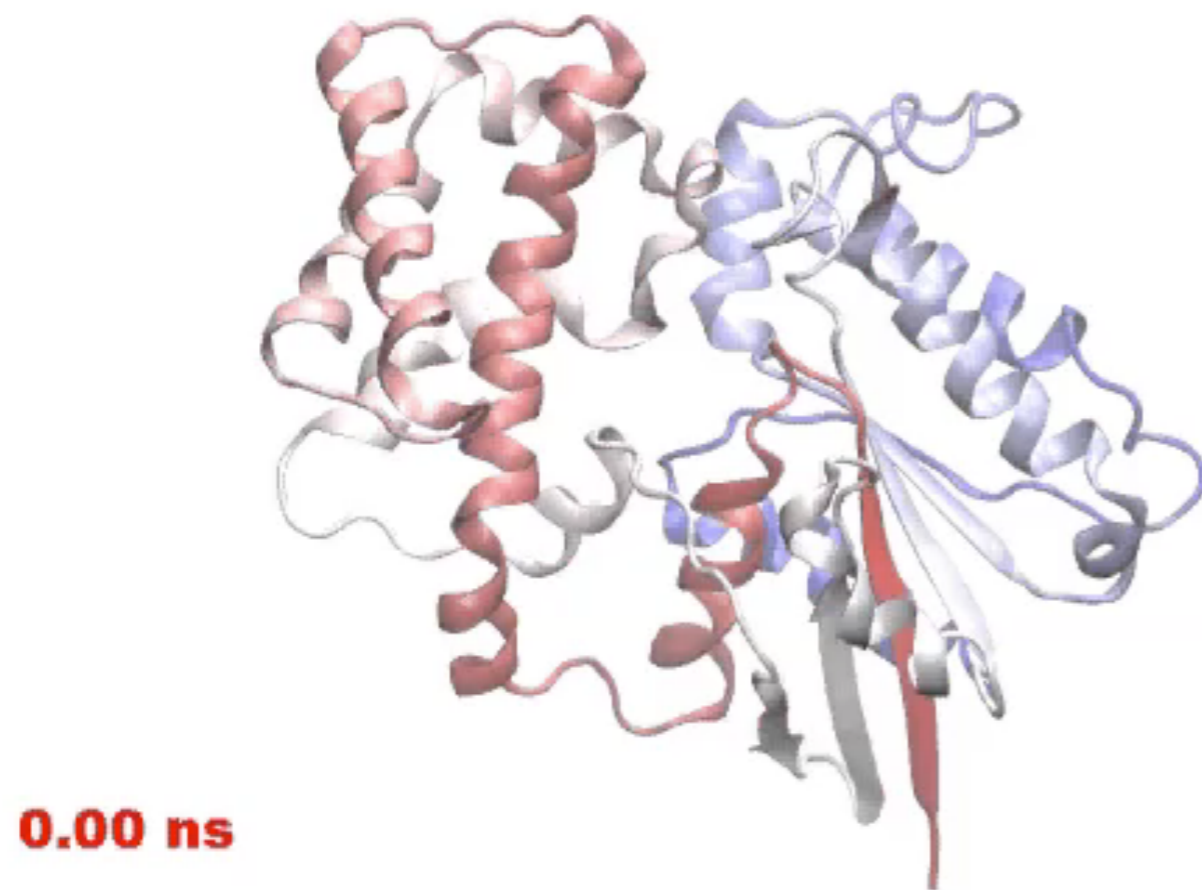
**REPEAT, (iterate many, many times... 1ms = 10<sup>12</sup> time steps)**



$$\begin{aligned} \mathbf{v}(t + \frac{\Delta t}{2}) &= \mathbf{v}(t - \frac{\Delta t}{2}) + \frac{\mathbf{F}(t)}{m} \Delta t \\ \mathbf{r}(t + \Delta t) &= \mathbf{r}(t) + \mathbf{v}(t + \frac{\Delta t}{2}) \Delta t \end{aligned}$$

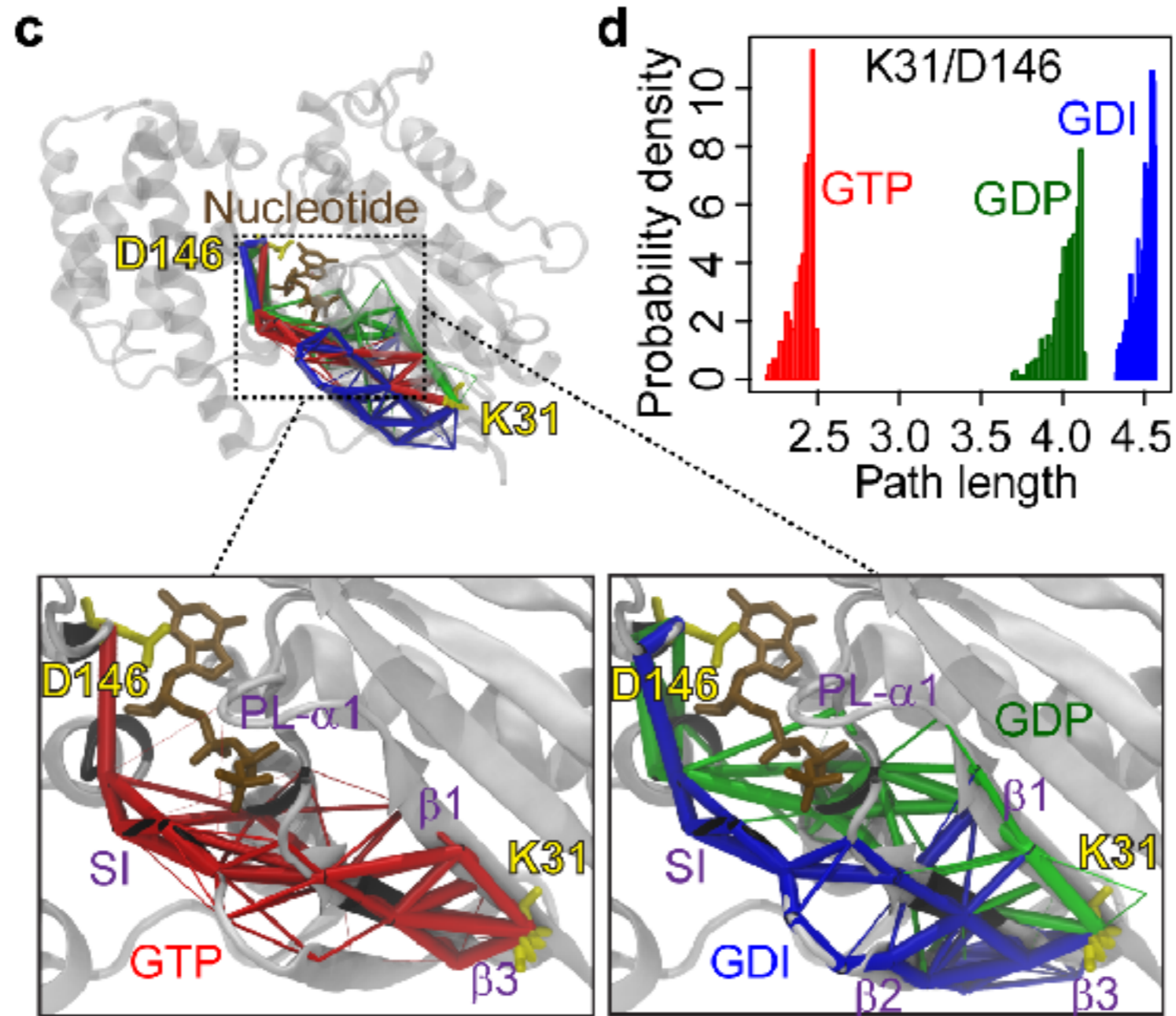
# MD Prediction of Functional Motions

Accelerated MD simulation of  
nucleotide-free transducin alpha subunit



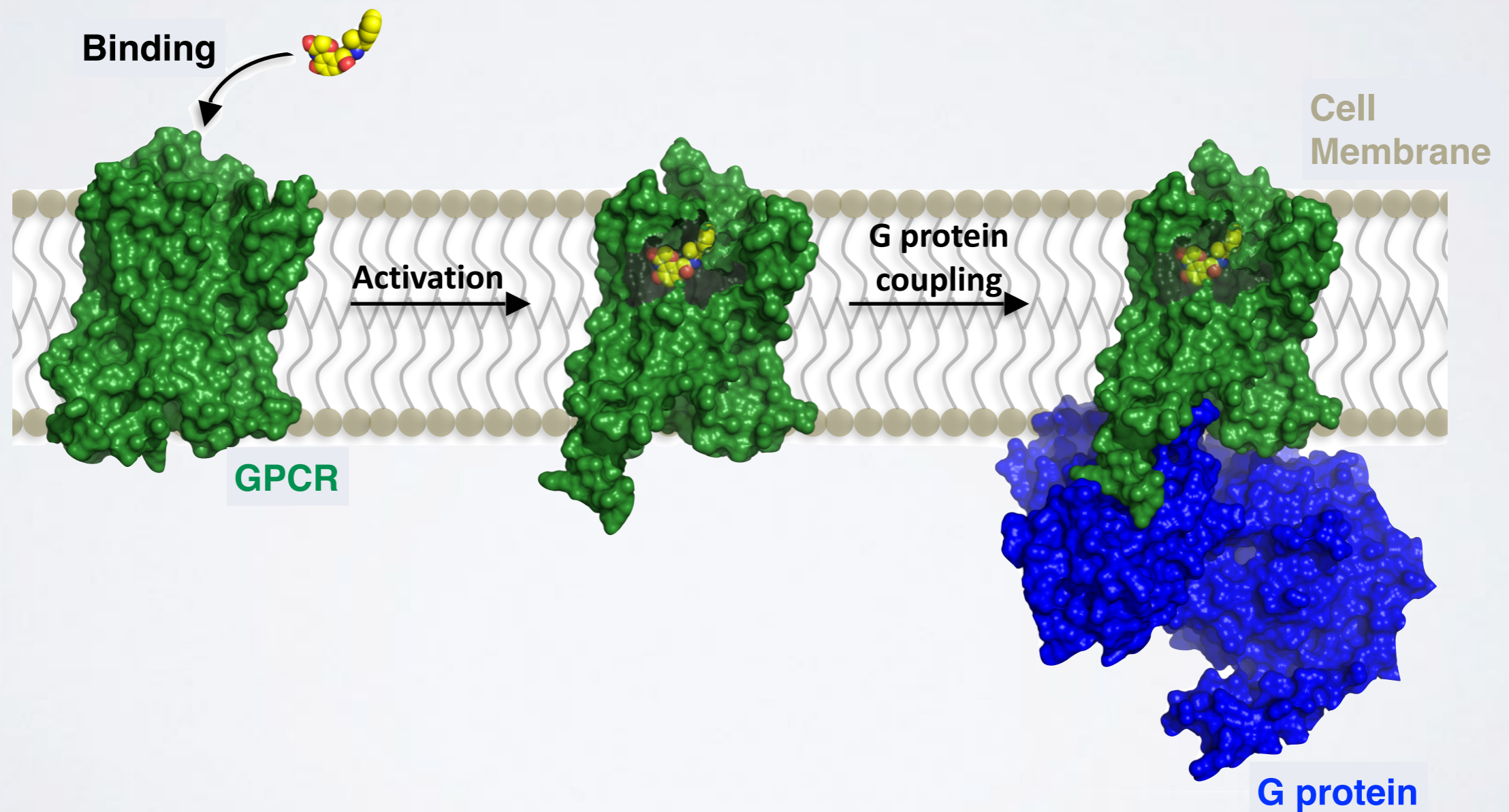
Yao and Grant, Biophys J. (2013)

# Simulations Identify Key Residues Mediating Dynamic Activation

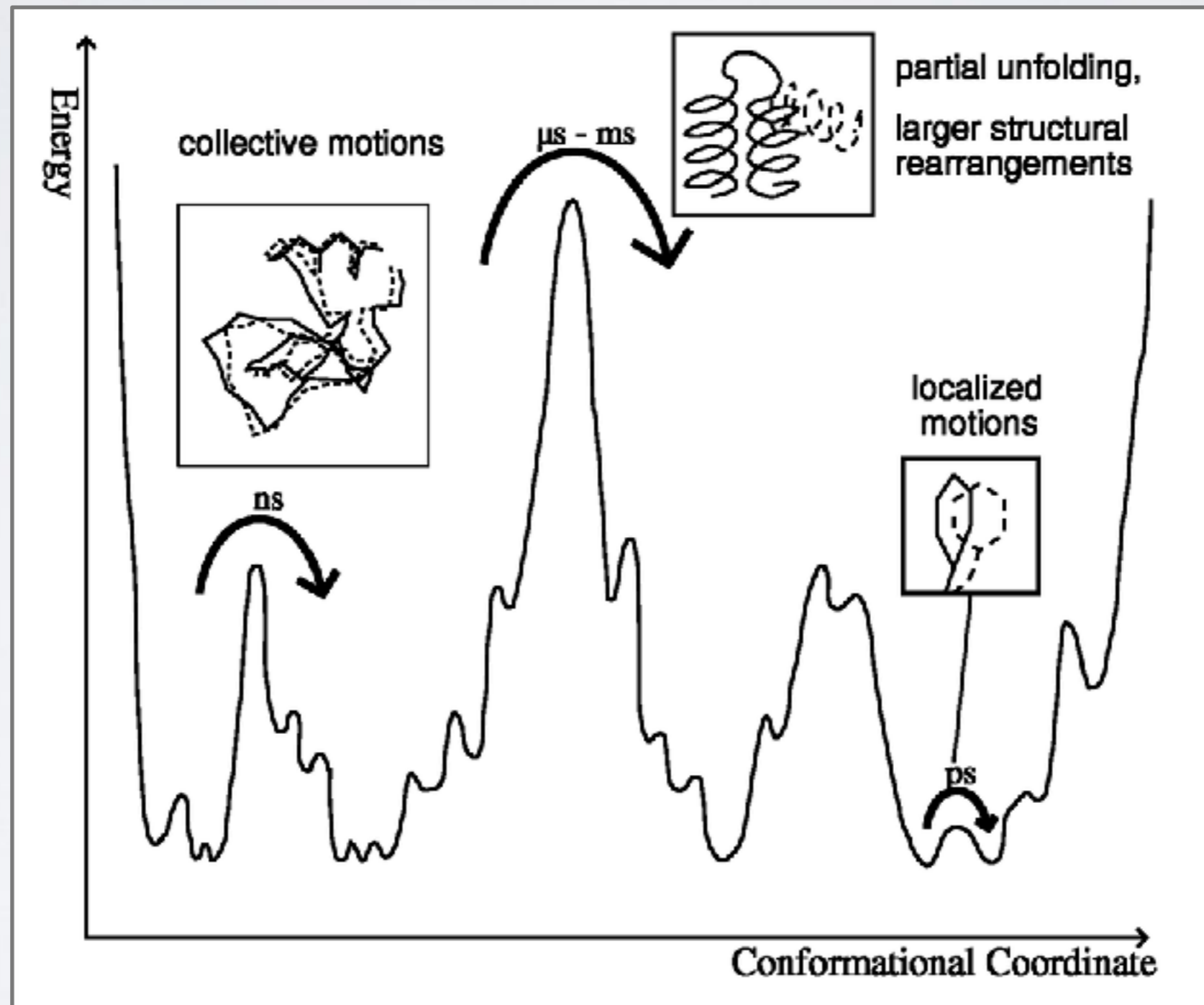




# EXAMPLE APPLICATION OF MOLECULAR SIMULATIONS TO GPCRS



# PROTEINS JUMP BETWEEN MANY, HIERARCHICALLY ORDERED “CONFORMATIONAL SUBSTATES”



H. Frauenfelder et al., *Science* **229** (1985) 337

# MOLECULAR DYNAMICS IS VERY

**Example:** F<sub>1</sub>-ATPase in water (183,674 atoms) for 1 nanosecond:

=>  $10^6$  integration steps

=>  $8.4 * 10^{11}$  floating point operations/step

[ $n(n-1)/2$  interactions]

Total:  $8.4 * 10^{17}$  flop

(on a 100 Gflop/s cpu: **ca 25 years!**)

**... but performance has been improved by use of:**

multiple time stepping ca. 2.5 years

fast multipole methods ca. 1 year

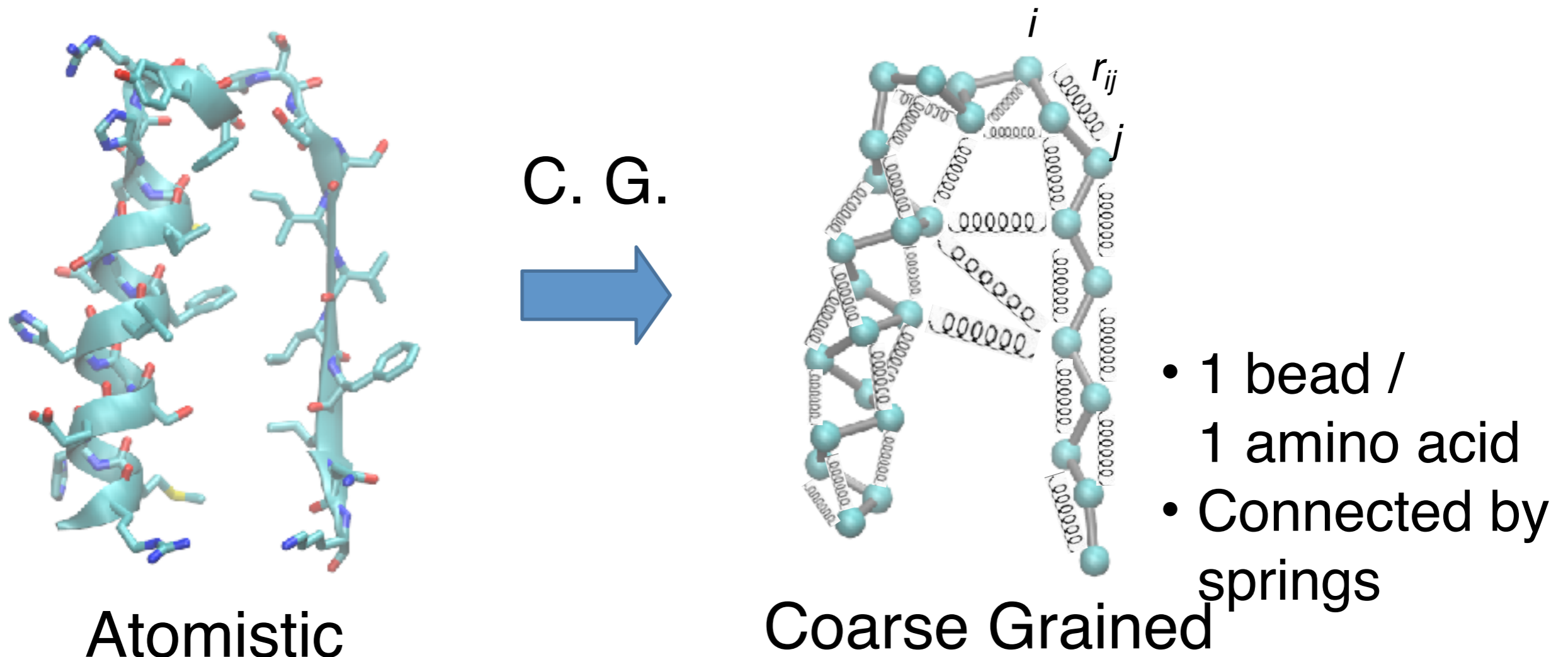
parallel computers ca. 5 days

modern GPUs **ca. 1 day**

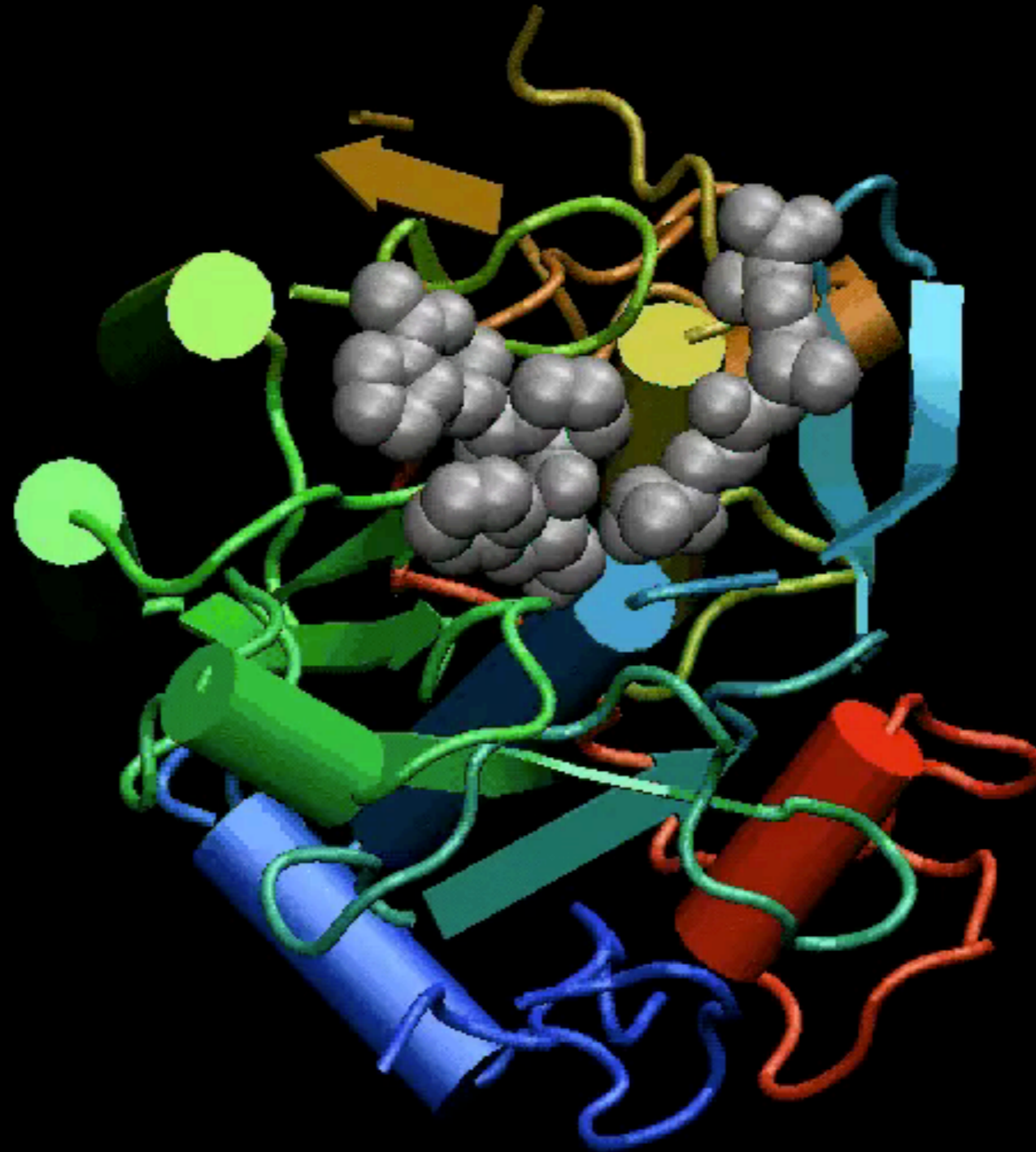
**(Anton supercomputer ca. minutes)**

# COARSE GRAINING: **NORMAL MODE ANALYSIS** (NMA)

- MD is still time-consuming for large systems
- Elastic network model NMA (ENM-NMA) is an example of a lower resolution approach that finishes in seconds even for large systems.



NMA models the protein as a network of elastic strings



Proteinase K

Do it Yourself!

# Hand-on time!

[https://bioboot.github.io/bimm143\\_S18/lectures/#11](https://bioboot.github.io/bimm143_S18/lectures/#11)

Focus on **section 6** to **7**

# SUMMARY

- Structural bioinformatics is computer aided structural biology
- Described major motivations, goals and challenges of structural bioinformatics
- Reviewed the fundamentals of protein structure
- Introduced both physics and knowledge based modeling approaches for describing the structure, energetics and dynamics of proteins computationally